

# **Plant Archives**

## Journal home page: www.plantarchives.org

DOI Url: https://doi.org/10.51470/PLANTARCHIVES.2021.v21.no1.221

#### VERSATILE ROLE OF AUXIN AND ITS CROSSTALK WITH OTHER PLANT HORMONES TO REGULATE PLANT GROWTH AND DEVELOPMENT

Hammad Ishtiaq, Savita Bhardwaj, Aaliya Ashraf and Dhriti Kapoor\*

Department of Botany, School of Bioengineering and Biosciences, Lovely Professional University, Delhi-Jalandhar Highway, Phagwara 144411, Punjab, India

\*E-mail: dhriti405@gmail.com

(Date of Receiving-20-01-2021; Date of Acceptance-14-04-2021)

ABSTRACT
Plant growth regulators are significant chemical compounds which are synthesized inside the plant cells and play vital role in plant growth and development. Such compounds are usually active at very low concentrations. These plant growth regulators act as a signalling molecule, which influences the growth of plants. Throughout the previous year's remarkable investigation have been done for understanding the synthesis of auxin and its effect on various physiological progressions. Auxin is a plant hormone that is involved in various physiological activities, including basic cellular processes such as cell enlargement, regulation of the cell cycle and distinction progress. Plants and several other microorganisms together produce auxin in order to carry out their cell cycle. The chemically synthesized auxins like NAA (naphthalene acetic acid) and IBA (Indole- butyric acid), also take part in various cellular processes. Against various types of biotic and abiotic stress conditions, these plant hormones significantly contribute in promoting acclimatization and adaptation in combination with other phytohormones. The present review highlights some of the important features of auxin role in regulation of plant growth either alone or in crosstalk with other plant hormones.

Keywords: Auxin, phytohormones, physiological, cell cycle, regulation, growth.

## INTRODUCTION

Plant growth regulators play an important part in various functional processes related to plant growth and development. These are biological constituents generally synthesized in advanced group of flora, regulating various evolutionary and biological purposes, distant from manufacturing sites and vigorous in tiny quantities. Generally, hormones are synthesized in a particular place and then these are distributed in different parts of plants. Plant hormones are functional message-carriers that must be used to alter the entire plant natural life, including propagation, radicalisation, fruit maturing, flowering, leaves and fall. Five groups of hormones are typically registered, the most common being auxins, gibberellins, abscisic acid and ethylene. Among, this Auxin is considered as the chief hormone for plants.

The word 'auxin' derived from the Grecian expression "Auxein" that involves increase in growth (Salehin *et al.*, 2015; Sauer *et al.*, 2013). Auxin acts as a plant growth regulator which is synthesized inside the plant body chemically as well as biologically. Synthesis of Indole acetic acid occurs biologically, as it is a natural plant growth regulator; whereas Naphthalene acetic acid and Indole-butyric acid are synthesized chemically. Auxin was acknowledged as the plant growth regulator due to its potential to promote variation progress in response to light inducements. In laboratory immunoassay, where auxin- carrying agar blocks stimulated the growth of oat coleoptile which identify Indole-3- acetic acid as an occurring auxin in plants. Reflective improvements in plant growth and production have been initiated by the application of IAA or synthetic auxins on plants. In plants, auxin biosynthesis is incredibly complex phenomenon. Several pathways may have engaged in the development of new auxins via hydrolytic breakdown of IAA methyl ester, IAA sugar.

All cellular activities in advanced plants are regulated by phytohormones and play an important role in coordinating several signal transduction pathways in plants (Pieterse *et al.*, 2009). Abscisic acid, salicylic acid, jasmonate and ethylene are known to provide resistance against different types of stress conditions (Pieterse *et al.*, 2012).

## Role of Auxin in plant Growth and Development

In plants, development is seen as an irreversible increase in the scale attained through expansion of discrete cells, a process occurring due to absorption of water. Auxin belongs to such class of phytohormones which is related to the optimal quantifiable growing changes throughout the plant life cycle. The effect of auxin was first realized when The Power of the Movement in plants was published by Charles and Francis Darwin. They discovered that an "effect is transferred" from one side of the grass coleoptile exposed to sun to another part as shown by the latter bending towards light. In 1926, auxin was extracted from plant tissue, and its ability for stimulation of growth was found. These days, the term auxin describes the group of vital molecules in plants that may be there in microorganisms and humans. One of the most important auxins, IAA is well known for its ability to regulate various attributes of plant growth. Artificial auxin, for example, 2, 4- dichlorophenoxyacetic acid is a vital herbicide. The amount and concentration of auxin used determines its influence on the plant development. Endogenous IAA is involved in embryonic and post-embryonic development, and tropisms for e.g. movement with respect to light and gravity.

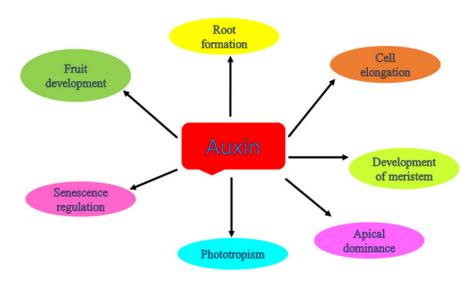


Figure 1: Role of auxin in regulating plant growth and development.

Although auxin plays an important role in phototropism and apical dominance, its main function remains the elongation of coleoptile and rooting (Went, 1934) (Fig 1). Auxin acts as a pivotal phytohormone for plant growth and regulation of different plant functions such as, pathogens stress (Fu and Wang, 2011), abiotic stress (Wang et al., 2010), senescence regulation (Elis et al., 2005), fruit forming (De Jong et al., 2009) and leaf abscission (Rubinstein and Leopold, 1963). Auxin also regulates the development of meristems, the division of the cells, and enhances the development of adventitious roots. Indole-3-acetic acid is the naturally occurring auxin, that triggers the synthesis coleoptile parts of oat (Bonner and Bandurski, 1952). Auxin is present in large amounts in early leaves, floral organs and developing fruits and seeds (Law and Davies, 1990). The growth and development of the plants is mainly regulated by phytohormone auxin. Auxin supports polarity development and preservation, apical superiority and tropical light and or gravity reaction (Woodward and Bartel, 2005). It regulates cell division at the cellular level, e.g. the organisations of meristem development results in new structures like lateral and adventitious roots and elongation of cell through changes in cell wall plasticity. In addition, auxin also crosses talks with other plant hormones through various biochemical pathways (Vanstraelen and Benkova, 2102).

#### Physiological role of auxin in plants

Stimulatory effect of auxin on cell elongation could be identified in parts of coleoptiles or stems

with physiologically significant IAA absorptions. Auxin shows its maximal effect in the concentration ranging from 0.1-10µm in most of the cases (Funada et al., 2001). Initiation, development and regulation of the cell division in buds monitored by auxin. Similarly, auxin takes part in mitosis and DNA replication (Jouanneau, 1971), is involved in the dissimilarity between buds and roots and stimulates separation of vascular bundles (Aloni, 2010). Development of adventitious roots is promoted by auxin. In the normal growth of the plants, auxin promotes

cell elongation and can promote the effect of light and gravity on the plant growth (Bernier and Kinet, 1986). Phototropism may be considered as the cause of the lateral reorganisation of auxin. The apical dominance is conveyed downhill through the stem to the neighbouring buds and hampers development under the influence of the auxin synthesized at the terminal bud. (Badescu and Naper, 2006). Higher auxin concentration hinders elongation of root, but the number of lateral roots is highly increased, i.e. (Benjamins and Scheres, 2008). Respiration rate in plants is increased obliquely by rapid usage of ATP in growing cells that brings an increase in the amount of ADP. Auxin plays an important role in development of callus. Several synthetic auxins, in particular 2,4-D and 2,4,5-T, are effective at higher concentrations in marijuana abolition (Zhao, 2010). Auxin however inhibits flowering, but it enables unchanging flowering in Lettuce and pineapple (Petrasek and Friml, 2009).

#### Auxin and its crosstalk with other Phytohormones

The plant hormones function together by stimulating

various biological processes, either during the release of chemicals when prone to external stimulus or during the addition of phosphate to ADP molecule. These chemical signals control DNA, produce functional proteins, which then influences the production or activity of various hormones and growth of plant in synchronization with various stimuli generating external factors. Plant hormonal signalling procedures manage the developmental aspects and vigour of plants, either alone or crosstalk between them. The content of hormones, released by plants such as auxin, ethylene, cytokinin, abscisic acid, gibberellin, and brassinosteroids vary at the cellular level and exhibit their functions either individually or in crosstalk with each other.

## Auxin and gibberellic acid

Gibberellins, agroup of plant hormones influence the several aspects to influence the development of plant and inhibition in their biosynthesis results in dwarf plants (Fleetand Sun, 2005; Ninnemann et al., 1964). Moubayidin et al., (2010) showed that in the early stages of meristem development, the high concentrations of gibberellic acid block the regulation of transcription factor ARR1. The ARR1stimulates the SHY2 gene, that shows antagonistic control over the auxin carrying PIN genes. So gibberellic acid forms a route to control the stability occurring during auxin signalling. During the tuber development, both GAs and auxin participate together. During the stolen developmental phase of plant life cycle, the concentration of GA is fairly elevated which result in the increase of stolen length whereas, the auxin concentration is less and has the role to maintain the apical dominance of plant stolen (Roumeliotis et al., 2012). Supplementation of GA together with auxin caused fruit developments in which cell expansion and quantity of pericarp cells were alike asthat in seeded fruits, signifying that GA and auxin are needed for standard fruit formation (Serrani et al., 2007). Whereas, auxin triggered fruit formation was considerably declined by the instantaneous supplementation of GA biosynthetic inhibitors, which showing that influence of auxin on fruit development is mediated by GA (Serrani et al., 2008). GAs network with auxin to stimulate lateral root formation where firstly, auxin amount significantly increased in the roots of the GA-deficient and GA-insensitive lines and then functioning of the gene expressing PIN9, was greatly escalated in both transgenic plants primarily in the roots (Farquharson, 2010).

## Auxin and Cytokinin

The balance between auxin and cytokinin signaling is critical at initial growth stages: for example, auxin promotes cell division in Arabidopsis root meristems (DelloLoio et al., 2008) whereas cytokinin activates cell differentiation in shoot (Perilli et al., 2010). Cytokinin and auxin association significantly helpful in stimulating the root development (Dello et al., 2008; Bielach et al., 2012). In Arabidopsis thaliana, the cytokinin endorses the cell differentiation, suppressing transport of auxin and signalling across the cell. At the molecular level, the auxin and cytokinin do show interdependent relations to control the direction of the root towards the gravity experienced by it (Buer et al., 2006), the development of the root (Rahman et al., 2001), the growth of lateral root (Ivanchenko et al., 2008; Negi et al., 2008, 2010) and the variation shown by the relative humidity (Pitts et al., 1998). These two hormones interact to mediate the meristem size of the root and also mediate the root growth. Crosstalk between auxin and cytokinin responsible for maintaining stability between cell differentiation and cell division, required for regulating root meristem dimensions and root development; which happens via a direct controlling network uniting on the SHY2 (SHORT HYPOCOTYL 2) gene, a member of the Aux/IAA gene group (Moubayidin et al., 2009).

## Auxin and ethylene

The auxin in association with ethylene conjointly controls a diversity of biological progressions that lead to growth in plants. Auxin which is a crucial regulator of plant development, induce ethylene synthesis, cellular signal transduction and regulates the response of growth to ethylene. The crosstalk between auxin and ethylene is much significant to regulate several aspects of plant morphogenesis, cell division within the root cells, emergence of root hairs, the development of lateral root and seed germination. The 1-aminocyclopropane- 1-carboxylate synthase (ACS) gene is under the control of auxin, that encrypts the main enzyme during the synthesis of ethylene (Muday et al., 2012). Ethylene mediated1° root growth need auxin production, passage, and signalling (Qin and Huang, 2018).In Arabidopsis, treatment with ethylene improve the functioning of IAA production genes and IAA concentration and also enhanced the whole auxin action at the root tip (Růžička et al., 2007).TFs which involve in the ethylene signalling, like EIN3, ERF1 and PIF4, acts

as crosstalk branches amongst ethylene and auxin in root development (Liu *et al.*, 2016; Mao *et al.*, 2016).Auxin and ethylene show contrary roles in lateral root growth for instance, ethylenedecline DR5erv:GFPfunctioning in those areas where lateral roots arise, showing decrease dauxin reaction in the vicinity of cell (Lewis *et al.*, 2011).OsERF3, which encodes an AP2/ERF protein, affect ethylene production and functions as a WOX11-association moiety in rice crown root growth. Furthermore, auxin and ethylene application stimulate the functioning of OsERF3, exhibiting synergistic association between ethylene and auxin during crown root growth (Qin and Huang, 2018).

## Auxin and other plant hormones

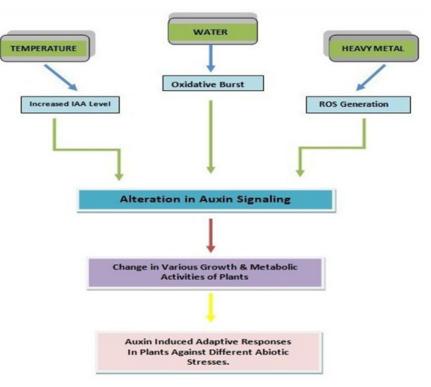
ABA is an isoprenoid hormone, concerned with controlling dormancy of seeds as well as plant response to various environmental stresses (Frankelstein and Rock, 2002). The gene VIVIPAROUS 1 (VP1) in maize and its Arabidopsis ortholog AB13, which encrypts a transcription factor tangled in ABA signalling, has been experimentally discovered to be auxin-inducible (Suzuki et al., 2001; Brady et al., 2003). ABA thus interrelates with the framework of crosstalk via auxin action. WRKY46 recognized to regulate the stimulation of lateral root development by triggering auxin stability and ABA perception in response to osmotic stress (Ding et al., 2015). Auxin associated with ABA triggered tolerance to limited water supply, hence, improve the functioning of ABA mediated gene expression of RAB18, RD22,

RD29A, RD29B, Dehydration responsive element binding factor 2A, and 2B) (Shi al., 2014).Brassinosteroids (BRs) et interrelate with auxin, for example, the appearance of multiple auxin- responsive genes is coordinated by both BRs and auxin pathways in combination (Mouchel et al., 2006). ARF2 considerably influence auxin biosynthetic genes, which signifies that BRs might regulate auxin production (Vert et al., 2008). BRs trigger the development of dark grown hypocotyls and cause phenotypic alterations, thar are triggered by IAA19 and ARF7, 2 crucial constituents of auxin signalling, via precise modulation of BZR1 (Zhou et al., 2013). OsMYB-R1 TFsregulate the cross-talk between auxin &salicylic acid perception as well as additional stress related genes by amending genetic

and root apparatus (Tiwari et al., 2020).

## Role of Auxin under abiotic stress conditions

Plants are often exposed to several abiotic stresses during their life cycle. The commonly occurring abiotic stresses that a plant experience, includes limited water supply, salinity, high or low temperature, heavy metal stress(Ahmad and Prasad, 2011; Iqbal et al., 2018). To cope with these unfavorable environmental conditions, plants develop advanced mechanisms to maintain equilibrium between their development, reproduction and survival (Fig 2). Reactive oxygen species (ROS), reactive nitrogen species (RNS), as well as phytohormones, are the essential signaling assets in plant tolerance responses to ever changing ecosystem (Kazan, 2013; Baxter et al., 2014; Kolbert, 2016). These signaling assets crosstalk with each other to improve plant tolerance to stress circumstances through various defense mechanisms (Mittler et al., 2011; Corpas and Barossa, 2013; Xia et al., 2015; Tognetti et al., 2017; Choudhary et al., 2017; Raja et al., 2017). Review of genome- wide induction suggested thatunder abiotic stresses such as drought, action of auxin sensitive genes stimulated (Jain and Khurana, 2009; Van Ha et al., 2013). ABA mediated inhibition of lateral root growth is opposed by the auxin positive regulation. However, it was also shown that ABA can regulate the auxin transport to manage the root development under water stress by stimulating proton gradient at root tips (Yamaguchi and Sharp, 2010; Xu et al., 2013).



perception, endogenous tissue stability Figure 2: Auxin induced plant responses to various abiotic stresses.

## **Drought stress**

Increased amount of IAA resulted in inhibition of plant growth, which is due to the altered hormonal balance in response to stress environments (Fahad et al., 2015). IAA supplementation controlled the apo plastic H<sub>2</sub>O transport flow in non- arbuscular and arbuscular mycorrhizal maize plants under limited water supply conditions (Quiroga et al., 2020). Application of auxin significantly provided tolerance to drought stress in Arabidopsis by improving the root architecture, particularly lateral root number, functioning of ABA-mediated genes, and also regulated the level of metabolites like amino acids, organic acids, aromatic amines, carbohydrates, sugar alcohols; ultimately diminished the ROS level by decreasing amount of  $H_2O_2$  and  $O_2^{-}$ , and increased the antioxidative activities of SOD, CAT, POD, and GR enzymes(Shi et al., 2014).Auxin supplementation considerably elevated the relative water content, chlorophyll amount, the amount of endogenous ABA & JA phytohormones, the functioning of auxin responsive genes, and also altered the expression of genes associated with leaf senescence to confer drought tolerance to white clover plant genotypes (Zhang et al., 2020).

## Salinity stress

Auxin (IAA) treatment improved the level of photosynthetic pigments and functioning of antioxidative enzyme i.e. GPX, CAT, SOD in okra plants under salinity stress, therefore it was found that application with IAA in okra seeds could be a promising strategy for cultivar upgrading under high salt concentrations (Esan et al., 2017). IAA supplementation improved the salt tolerance in rice cultivars through increasing grain yield, filled grain %age, which are related with elevation in the amounts of carbohydrates moieties such as starch, sucrose, glucose and fructose in rice grains via IAA treatment (Javid et al., 2011). Exogenous supplementation of IAA significantly increased the plant growth by increasing the level of photosynthetic pigments, decreased the Na<sup>+</sup> amount and increased the amount of  $K^+$ ,  $Ca^{2+}$ , and however, increased the leaf  $Na^+/K^+$ ratio. It also declined the membrane permeability, and changed the functioning of antioxidative enzymes such as SOD, CAT under salinity stress inZea maysplants (Kaya et al., 2013).

## Heavy metal stress

Abiotic stress circumstances result in stress-triggered

morphological changes, for instance, suppression of 1° root length, improved lateral root development and escalated root hairs formation. In this context, auxin play crucial role in regulating these morphological alterations, and modify plant reaction to heavy metal stress by altering auxin homeostasis involving auxin relocation, passage and steadiness (Yuan et al., 2013). Treatment with IAA at small amounts alleviated Cd toxicity, through considerable decrease in Cd uptake, as well as by escalating the plant growth by improving the functioning of photosynthetic apparatus and antioxidative enzymes activities in Trigonella foenum-graecums eedlings grown under Cd stress (Bashri and Prasad, 2015). Application of IAA improved the root and shoots growth and elevated the rate of Cd, Pb and Zn phytoextraction in sunflower and maize (Fässler et al., 2010; Hadi et al., 2010). Exogenous application of IAA, GA<sub>3</sub>, and citric acid increased plant shoot dry weight Panicum virgatum in lead contaminated soil (Aderholt et al., 2017). Treatment with IAA considerably enhanced stem length, amount of photosynthetic pigments, and antioxidative enzymes activities, and also decreased the proline level in leaves, obstructed the Cd passage from roots to shoot in Triticum aestivum under Cd toxicity (Agami and Mohamed, 2013).

IAA application obstructed the Cd accumulation in roots& its transport from roots to shoot, improved plant growth, functioning of photosynthetic apparatus, proline content, strengthens the plant antioxidant defense system by improving the activity of antioxidative enzymes, regulated the C and N metabolism to alleviate adverse effects of Cd in tea plants (Zhang et al., 2020b).IAA treatment improved the plant growth, biomass, amount of photosynthetic pigments & net photosynthetic rate, soluble sugars content, decreased the endogenous amount of Cd and escalated the activities of POX, SOD enzymes while reduced the protein amount in Cyphomandrabetacea seedlings to alleviate Cd toxicity (Li et al., 2020). Auxin alleviated the Cd stress by significantly enhancing the plant growth, amount of photosynthetic pigments, relative water content, diminished the ROS induced oxidative stress through improving functioning of antioxidative enzymes i.e. SOD, CAT and POX to mitigate Cd toxicity in wheat (Agami and Mohamed, 2013).

## CONCLUSION

Phytohormones are essential internal messengers that involve germination, rooting, development, flowering and fruit ripening etc. to control the entire plant life cycle. Among various plant hormones like auxins, gibberellins, cytokinin's, abscisic acid and ethylene, auxin is of critical importance. Auxin plays a vital role in controlling cell elongation, cell division, proliferation, root initiation, apical dominance and tropical responses, and is crucial regulator for plant growth and development. It is also quite important that auxin plays a vital role in generating & sustaining primary meristems, and in the development of axillary meristems. Therefore, it is proved that auxin is an essential phytohormone which play vital role in numerous cellular processes along with providing resistance against different biotic as well as abiotic stress conditions.

#### REFERENCES

- Aderholt, M., D. L. Vogelien, M., Koether, and S. Greipsson (2017). Phytoextraction of contaminated urban soils by Panicum virgatum L. enhanced with application of a plant growth regulator (BAP) and citric acid. *Chemosphere*, 175: 85-96.
- Agami, R. A., and G. F. Mohamed (2013). Exogenous treatment with indole-3-acetic acid and salicylic acid alleviates cadmium toxicity in wheat seedlings. *Ecotoxicol. Environ. Saf.*, 94: 164-171.
- Bashri, G., and S. M. Prasad (2015). Indole acetic acid modulates changes in growth, chlorophyll a fluorescence and antioxidant potential of Trigonella foenum-graecum L. grown under cadmium stress. *Acta Physiol. Plant.*, 37(3): 49.
- Benjamins R, and B Scheres (2008). Auxin: the looping star in plant development. *Annu. Rev. Plant Biol.* 59: 443-465.
- Bielach A, J Duclercq, P Marhavy', and E Benkova (2012). Genetic approach towards the identification of auxin– cytokinin crosstalk components involved in root development. *Phil Trans R Soc B*.367:1469–1478.
- Bonner J, and RS Bandurski (2005). Studies of the physiology, pharmacology, and biochemistry of the auxins. *Annu. Rev. Plant Physiol.*,**3**(1): 59-86.
- De Jong M, C Mariani, and W.H. Vriezen (2009). The role of auxin and gibberellin in tomato fruit set. *J.Exp.Bot.*, ERP: 094.
- Dello Ioio R, K Nakamura, L Moubayidin, S Perilli, M Taniguchi, MT Morita, T Aoyama, P Costantino and S Sabatini (2008). A genetic framework for the control of cell division and differentiation in the root meristem. *Science*, 322: 1380–1384.
- Ding, Z. J., J. Y., Yan, C. X. Li, G. X., Li, Y. R., Wu and S. J. Zheng (2015). Transcription factor WRKY 46 modulates the development of Arabidopsis lateral roots in osmotic/salt stress conditions via regulation of ABA signaling and auxin homeostasis. *Plant J.*, 84 (1): 56-69.

Ellis C.M., P. Nagpal, J.C. Young, G. Hagen, T.J. Guilfoyle and

J.W. Reed (2005). Auxin Response Factor 1 and Auxin Response Factor 2 regulate senescence and floral organ abscission in Arabidopsis thaliana. *Development*, 132 (20): 4563-4574.

- Esan, A. M., K. Masisi, F.A. Dada and C. O. Olaiya (2017). Comparative effects of indole acetic acid and salicylic acid on oxidative stress marker and antioxidant potential of okra (Abelmoschus esculentus) fruit under salinity stress. *Sci Hortic.*, 216: 278-283.
- Fahad, S., S. Hussain, A. Bano S. Saud, S. Hassan, D. Shan and M.A. Tabassum (2015). Potential role of phytohormones and plant growth-promoting rhizobacteria in abiotic stresses: consequences for changing environment. *Environ. Sci. Pollut. Res.*, 22(7): 4907-4921.
- Farquharson, K. L. (2010). Gibberellin-auxin crosstalk modulates lateral root formation. *Plant Cell*,22(3): 540.
- Fässler, E., M. W. Evangelou, B. H., Robinson, and R. Schulin (2010). Effects of indole-3-acetic acid (IAA) on sunflower growth and heavy metal uptake in combination with ethylene diamine disuccinic acid (EDDS). *Chemosphere*, 80(8): 901-907.
- Fleet, C.M., andT.P. Sun (2005). A Delicate balance: The role of gibberellin in plant morphogenesis. *Curr. Opin. Plant Biol.*,8: 77–85.
- Funada R, T Kubo, M Tabuchi, T Sugiyama, andM Fushitani (2001). Seasonal variations in endogenous indole-3-acetic acid and abscisic acid in the cambial region of Pinus densiflora Sieb. et Zucc. stems in relation to earlywood-latewood transition and cessation of tracheid production. Holzforschung,55(2): 128-134.
- Hadi, F., A., Bano, and M. P. Fuller (2010). The improved phytoextraction of lead (Pb) and the growth of maize (Zea mays L.): the role of plant growth regulators (GA3 and IAA) and EDTA alone and in combinations. *Chemosphere*, 80(4): 457-462.
- Ivanchenko MG, GK Muday, andJG Dubrovsky (2008). Ethylene–auxin interactions regulate lateral root initiation and emergence in Arabidopsis thaliana. *Plant J.*,55:335–347
- Javid, M. G., A. Sorooshzadeh, S. A. M. Sanavy, M. I. Allahdadi, and F. Moradi (2011). Effects of the exogenous application of auxin and cytokinin on carbohydrate accumulation in grains of rice under salt stress. *Plant Growth Regul.*, 65(2): 305-313.
- Kaya, C., M., Ashraf, M., Dikilitas, and A. L. Tuna (2013). Alleviation of salt stress-induced adverse effects on maize plants by exogenous application of indoleacetic acid (IAA) and inorganic nutrients-A field trial. *Aust. J. Crop Sci.*, 7(2): 249.
- Law DM, and P.J. Davies (1990). Comparative indole-3-acetic acid levels in the slender pea and other pea phenotypes. *Plant Physiol.*,93(4): 1539-1543.
- Lewis, D. R., S., Negi, P., Sukumar, and G. K. Muday (2011). Ethylene inhibits lateral root development, increases IAA transport and expression of PIN3 and PIN7 auxin efflux carriers. *Development*, 138(16): 3485-3495.

- Li, Z., J., Zhu, Y., Wang, L., Lin, M. A., J., Liao Wang, and J.Wang (2020). Effects of exogenous indole acetic acid on growth and cadmium accumulation of Cyphomandrabetacea seedlings. *Int. J. Environ. An. Chem.*, 1-9.
- Liu, G., S., Gao, H., Tian, W., Wu, H. S., Robert, and Z.Ding (2016). Local transcriptional control of YUCCA regulates auxin promoted root-growth inhibition in response to aluminium stress in Arabidopsis. *PLoS genetics*, 12(10): e1006360.
- Mao, J. L., Z. Q., Miao, Z., Wang, L. H., Yu, X. T., Cai, and C. B. Xiang (2016). Arabidopsis ERF1 mediates cross-talk between ethylene and auxin biosynthesis during primary root elongation by regulating ASA1 expression. *PLoS Genet.*, 12(1): e1005760.
- Moubayidin, L., R., Di Mambro, and S. Sabatini (2009). Cytokinin–auxin crosstalk. *Trends Plant Sci*, 14(10): 557-562.
- Moubayidin, L., S Perilli. R Dello Ioio., R Di Mambro. P Costantinoand, and S. Sabatini (2010). The rate of cell differentiation controls the Arabidopsis root meristem growth phase. *Curr. Biol.*,20: 1138–1143.
- Mouchel, C. F., K. S Osmont, and C.S. Hardtke (2006). BRX mediates feedback between brassinosteroid levels and auxin signalling in root growth. *Nature*,443: 458–461.
- Negi S, MG Ivanchenko, and GK Muday (2008). Ethylene regulates lateral root formation and auxin transport in Arabidopsis thaliana. *Plant J.*,55:175–187
- Ninnemann, H., J.A.D, Zeevaart, H., Kendeand, and A Lang (1964). The plant growth retardant CCC as inhibitor of gibberellin biosynthesis In Fusarium moniliform. *Planta*,61: 229–235.
- Petrášek J, and J. Friml (2009). Auxin transport routes in plant development. *Development*, 136(16): 2675-2688.
- Qin, H., and R. Huang (2018). Auxin controlled by ethylene steers root development. *Int.Journal Mol.Sci.*, 19(11): 3656.
- Quiroga, G., G. Erice, R. Aroca, Á. M. Zamarreño, J. M García-Mina, and J. M Ruiz-Lozano (2020). Radial water transport in arbuscular mycorrhizal maize plants under drought stress conditions is affected by indole-acetic acid (IAA) application. J. Plant Physiol., 246: 153115.
- Roumeliotis, E., R. G., Visser, and C. W. Bachem (2012). A crosstalk of auxin and GA during tuber development. *Plant Signal.Behav.*, 7(10): 1360-1363.
- Růžička, K., K., Ljung, S., Vanneste R., Podhorská, T., Beeckman J., Friml, and E Benková (2007). Ethylene regulates root growth through effects on auxin biosynthesis and transport-dependent auxin distribution. *Plant Cell.*, 19(7): 2197-2212.

- Sauer M, S Robert, and J Kleine-Vehn (2013). Auxin: simply complicated. J. Exp. Bot.,64(9): 2565-2577.
- Serrani, J. C., M., Fos, A., Atarés and J. L. García-Martínez (2007). Effect of gibberellin and auxin on parthenocarpic fruit growth induction in the cv Micro-Tom of tomato. J. Plant Growth Regul., 26(3): 211-221.
- Serrani, J. C., O., Ruiz-Rivero, M., Fos and J. L. García-Martínez (2008). Auxin-induced fruit-set in tomato is mediated in part by gibberellins. *Plant J.*, 56(6): 922-934.
- Shi, H., L Chen, Ye, T Liu, X., Ding, K., and Z Chan (2014). Modulation of auxin content in Arabidopsis confers improved drought stress resistance. *Plant Physiol. Biochem.*, 82: 209-217.
- Tiwari, P., Y., Indoliya, A. S., Chauhan, P., Singh, P. K., Singh, P. C., Singh, and D. Chakrabarty (2020). Auxinsalicylic acid cross-talk ameliorates OsMYB–R1 mediated defense towards heavy metal, drought and fungal stress. J. Hazard. Mater., 122811.
- Vanstraelen M, and E Benková (2012). Hormonal interactions in the regulation of plant development. *Annu.Rev.Cell Dev.Biol.*,28: 463-487.
- Vanstraelen, M., and E Eva Benkova (2012). Hormonal interactions in the regulation of plant development. *Annu. Rev. Cell Dev. Biol.*, 28: 463-486.
- Vert, G., C. L., Walcher, J., Chory, and J. L. Nemhauser (2008). Integration of auxin and brassinosteroid pathways by Auxin Response Factor 2. *Proc. Natl. Acad. Sci.*, 105(28): 9829-9834.
- Wang S, Y Bai, C Shen, Y Wu, S Zhang, D Jiang et al (2010). Auxin-related gene families in abiotic stress response in Sorghum bicolor. Funct.Integr. Genomics.,10(4):533-546.
- Yuan, H. M., W. C., Liu, Y., Jin and Y. T. Lu (2013). Role of ROS and auxin in plant response to metal-mediated stress. *Plant signal.Behav.*, 8(7): e24671.
- Zhang, Y., Li, Y., Hassan, M. J., Li, Z., & Y. Peng (2020a). Indole-3-acetic acid improves drought tolerance of white clover via activating auxin, abscisic acid and jasmonic acid related genes and inhibiting senescence genes. *BMC Plant Biol.*, 20: 1-12.
- Zhang, C., He, Q., M., Wang, X., Gao, J., Chen and C. Shen (2020b). Exogenous indole acetic acid alleviates Cd toxicity in tea (Camellia sinensis). *Ecotoxicol. Environ. Saf.*, 190: 110090.
- Zhou, X. Y., L., Song and H. W. Xue (2013). Brassinosteroids regulate the differential growth of Arabidopsis hypocotyls through auxin signaling components IAA19 and ARF7. *Mol.Plant.*, 6(3): 887-904.