

ABSTRACT

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MORPHOLOGICAL RESPONSES OF SUGARCANE CULTIVARS AGAINST WATER DEFICIT STRESS IN NORTHERN KARNATAKA REGION OF INDIA

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The morphological responses of sugarcane cultivars against water deficit stress were carried out in twenty-three cultivars of sugarcane grown in Yaragatti Farm of S. Nijalingappa Sugar Institute, Belagavi in Karnataka, India. Data was recorded at different time intervals, i.e., control, moderate and severe water deficit conditions. During these different time intervals, parameters such as the number of tillers, total plant height, stalk length, number of internodes and cane diameter were recorded to have a comparative study among varieties of sugarcane against water deficit stress. Thus, these distinct characters help to understand those varieties that are better suited for water deficit stress and can further be useful for selective breeding programme for commercial production of sugarcane cultivars. Therefore, cultivars which are resistant to water deficit stress such as Co 09004, Co 14011, Co 13003, Co 95020, Co 08020, Co 86032, Co 05001 and Co 671 showed increase in numbers of tillers, total plant height, stalk length, number of internodes, and cane diameter. Further, through these morphological studies, principal component analysis was done to identify the genotypes which are resistant to water deficit stress.

Keywords : Morphological parameters; Principal component analysis; Sugarcane; Water deficit stress.

INTRODUCTION

Sugarcane belongs to family Poaceae, genus Saccharum, tribe Andropogoneae. It has four different phases of growth such as germination, formative, grand growth and maturation phase (Gasho & Shih, 1983; Endres et al., 2018). It is cultivated about 19.38 million hectares in 2000 and an increase of 25.98 million hectares in 2017 was seen (Ram & Ramaiyan, 2019). It is grown in more than 120 countries among which Brazil (420,121,000 MT), India (232,320,000 MT), China (88,730,000 MT), Thailand (49,572,000 MT), Pakistan (47,244,100), Mexico (45,126,100 MT), Columbia (39,849,240 MT), Australia (38,246,100 MT), Philippines (31,000,100 MT) and USA (25,803,960 MT) are the top ten countries in production (FAO, 2005; Srivastava & Rai, 2012). Brazil ranks first in the world with 33% of global sugar cane production followed by India (23%) China (7%) and Pakistan (4%) (FNP 2009; Srivastava & Rai, 2012). Among which India occupies second position with respect to the area of 4.2 million hectares, having production of 314 million tons. Major sugarcane growing states in India which has the highest area of cultivation viz. Uttar Pradesh (47.05%) followed by Maharashtra (17.52%), Karnataka (7.76%), Tamil Nadu (7.47%), Gujarat (4.57%), Andhra Pradesh (3.76%) contributing 88% of total area (Srivastava and Rai, 2012) and remaining 12% sugarcane growing states are Bihar, Uttarakhand, Haryana, Madhya Pradesh,

Chhattisgarh and Punjab (ICRISAT, 2009; Srivastava & Rai, 2012). Saccharum barberi Jeswiet, the sugarcane of India, and Saccharum sinense Roxb. of China was known through folklore and mythology from 1000 to 500 B.C. These original sugarcanes were delivered to the Americans by Columbus in 1493 and spread by human from the Orient through the Middle East, Northern Africa, and the Mediterranean (Hussain et al., 2004). Interspecific crosses were done between S. officinarum having chromosome number '10', with high sugar content and S. spontaneum chromosomes number '8' having high plant vigor and resistance towards pest. These sugarcanes (S. officinarum X S. spontaneum) of distinct chromosomal organization and superiority provided the new approach in production of modern cultivars of sugarcane (Bordonal et al., 2018). Different sugarcane cultivars resemble each other in their appearance but it has different morphological traits. These traits are also influenced by the environmental factors (Chidambaram and Sivasubramaniam, 2017).

One of the key factors responsible for growth of crop is a change in climate which increases the rate of transpiration in leaves. As it develops different mechanism of resistance to deal with stress by escaping drought period and by reducing their life cycle (Cia *et al.*, 2012). However, up to 60% of total loss in production may be due to drought at early growth stage and midseason. However, tillering combined with early grand growth stage which is the most important phase during which 70-80% of the cane yield takes place. Any change caused due to environmental factor during this period leads to reduction in the total plant height, stalk diameter, number of tillers, leaf area and total biomass which in turn affects the yield (Devi et al., 2018; Dinh et al., 2017; Ferreira et al., 2017; Hemaprabha et al., 2013; Misra et al., 2020; Ramesh, 2000). Thus, studies revealed that those sugarcane varieties susceptible to drought wilted by having lesser cane production when compared to other varieties which are tolerant to stress that remain turgid. Hence this factor helps in identification and evolution of water stress tolerant varieties from that of susceptible ones (Hemaprabha et al., 2006, 2013). The present study focuses on the impact of water deficit stress on morphological parameters of 23 sugarcane cultivars during the early growth period.

MATERIALS AND METHODS

Experimental Layout and Design

Twenty-three sugarcane cultivars were selectively grown in Yargatti farm of S. Nijalingappa Sugarcane Institute, Belagavi, Karnataka, India. These includes Co 09004, Co 0303, Co 14011, Co 98017, Co 13003, Co 93009, Co 95020, Co 92013, Co 07015, Co 12007, Co 08020, Co 86032, Co 85019, Co 13006, Co 90003, Co 92002, Co 92020, Co 06015, Co 98008, Co 94005, Co 10033, Co 671 and Co 05001 were selected for water deficit stress. The plot size for each cultivar represented three rows of 1.5 cm spaced. The experiment consisted of 4 data collection (2 under water stress and other 2 after stress). The irrigation was given up to 60 days and then water stress was initiated by withholding irrigation from 60-120 Days After Planting (DAP) followed by normal irrigation until harvest for recovery studies. Periodical field visits were done satisfactorily to collect data during the period of February to December 2019. Measurable morphological characters such as, the number of tillers, total plant height (cm), stalk length (cm), number of internodes and cane diameter were taken into consideration for random stalks from each row. Stalk length was taken from last visible dewlap in cm. Cane diameter was measured from middle portion of stalk by using Vernier caliper in cm (Table 1). Internodal distance of each internode was measured by the measuring scale. Thus, estimation of the growth parameters of the plant samples, was done by taking three replications which were randomly selected and average of three replicas was calculated.

Statistical Analysis

The principal component analysis (PCA) was carried out on morphological parameters of sugarcane cultivars with different time intervals. Dendrogram analysis was performed for grouping patterns of sugarcane cultivars using SPSS Statistical Software (2011).

RESULTS AND DISCUSSION

Morphological Characterization

The yield of sugarcane mainly depends upon some of the morphological traits such as number of tillers (NOT), total plant height (TPH), stalk length (SL), number of internodes (NI) and cane diameter (CD) which were considered for the present study with different time intervals such as 90th (T₁), 120th (T₂), 150th (T₃), 270th (T₄) (DAP). Where 90th and 120th DAP was considered as water stress

period. Among the twenty-three cultivars, the tolerant cultivars showed lesser number of tillers at 120th DAP in cultivars Co 09004 (25), Co 671 (25), Co 14011 (24), Co 95020 (23), Co 92013 (27), Co 93009 (26), when compared recovery 270th DAP Co 09004 (88), Co 14011 (86), Co 95020 (85), Co 93009 (72), Co 92013 (71), Co 671 (89); whereas, the genotypes which are susceptible to stress showed reduction in number of tillers at 120th DAP genotypes such as Co 06015 (12), Co 12007 (16), Co 13006 (15), Co 92020 (14) when compared to 270 DAP Co 06015 (63), Co 12007 (72), Co 13006 (68), Co 92020 (67). Internode length in tolerant cultivars at 120th DAP in Co 09004 (3 cm), Co 14011 (4 cm) and Co 671 (4 cm), Co 08020 (4 cm), Co 10033 (3 cm) when compared to 270th DAP with Co 09004 (17 cm), Co 14011 (18 cm) and Co 671 (22 cm), Co 08020 (18 cm), Co 10033 (19 cm). Whereas, no growth has been observed in susceptible genotypes Co 86032, Co 07015, Co 92020, and Co 98008 at 120th DAP, when compared to 270th DAP in Co 86032(19), Co 07015 (14), Co 92020(15), and Co 98008(18). The number of internodes in water deficit stress affected canes was relatively lesser than in cane after rewatering. Similarly, (Hemaprabha et al., 2012; Misra et al., 2020) showed that there is decrease in the number of internodes in drought affected canes.

At 120th DAP, total plant height was high in water tolerant cultivars Co 09004 (190.5 cm), Co 14011 (186 cm), Co 671 (203 cm) and Co 10033 (223 cm) when compared to 270th DAP in Co 671 (379 cm), Co 10033 (356 cm), Co 09004 (370 cm) and Co 14011 (372 cm). Reduction was seen in susceptible cultivars at 120 DAP Co 06015 (144.5 cm), Co 07015 (153 cm), Co 92020 (155 cm) and Co 86032 (145 cm) in comparison with 270th DAP Co 06015 (309 cm), Co 07015 (344 cm), Co 92020 (318 cm) and Co 86032 (373 cm), Stalk length in tolerant cultivars at 120th DAP in Co 09004 (18.5 cm), Co 14011 (20.5 cm), Co 671 (21 cm) and Co 10033 (17cm) when compared to 270th DAP in Co 09004 (210 cm), Co 14011 (199 cm), Co 671 (231 cm) and Co 10033 (223 cm). There was no growth seen as such at 120 DAPS in susceptible genotypes Co 98008, Co 07015, Co 86032, and Co 92020 when compared to 270th DAP in genotypes Co 98008(199), Co 07015(182), Co 86032(199), and Co 92020(163). The difference in the stalk length after rewatering was highest than the decrease in the stalk length at water deficit stress. Cane height was affected by water deficit stress as illustrated by (Inman-Bamber & Smith, 2005; Misra et al., 2020; Silva et al., 2008). At 120 DAP, cane diameter at 120th DAP in tolerant cultivars such as Co 09004 (2.3 cm), Co 14011 (2.3 cm), Co 12007 (2.4 cm) and Co 10033 (2.4) when compared at 270th DAP in Co 09004 (3.1 cm), Co 14011 (3.0 cm), Co 12007 (3.0 cm) and Co 10033(3.2 cm). Cultivar Co 06015 showed less variation of cane diameter (2.1 and 2.7 cm) at 120th and 270th DAP respectively, whereas, slight difference has been observed in cane diameter of cultivar Co 07015 at 120th and 270th (0 and 2.9) DAP, which shows that it is very susceptible to water stress conditions. Under water deficit stress the canes are relatively thinner than the cane after rewatering. Similar finding made (Lal et al., 1968; Misra et al., 2020; Silva et al., 2008) demonstrated the effect of drought on stalk diameter and revealed that the cultivars which has thinner stalk might flourish well than that of the thicker stalk. Further, (Gomati & Chandran, 2009) made similar finding related to sugarcane waterlogging condition. Those genotypes which were resistant to water stress showed greater number of tillers, stalk length, cane diameter, number of internodes and total plant height (Table 2 and 3).

According to the report (Silverio *et al.*, 2017) the RB073040, RB867515, RB72454, RB855536, and RB073028 variety showed higher increase in stem length with 45.83 cm, 45.50 cm, 44.66 cm, 42.50 cm, and 40.66 cm respectively. Whereas the RB073028 variety was remarkable when compared to the remaining variety with 45.50 cm stalk length. However, the variety RB073036 showed shortest stalk length of 33.33 cm. further at 60 kPa tension significant difference in stem length was noted.

Similar findings were made (Ecco *et al.*, 2014) under water deficit stress note that RB867515 had highest increase of 13.7 cm value whereas RB855536 with 14.8 cm had lowest value hence concluded that longer stress period had slower recovery rate. A study made (Endres *et al.*, 2018) on different phenophases of water deficit stress led to mean reduction in plant height of 44.5% where variety RB855113 recorded highest reduction with (54.0%) and SP79-1011 recorded lowest reduction of 34.8%.

Minimum, Maximum, Average, and Standard Deviation (SD) Analysis

A greater number of coefficients of variation were found for the traits such as TPH at T4, and TPH at T2 with 713.08, 373.29 values respectively. The average CVs varied between 0.02 and 713.08%. The standard deviation varied between 0.15% and 26.70% (Table 4). Thus, the highest variability observed for the values of CV was TPH at T4 with standard deviation of 26.70% and the trait with lowest variability was cane diameter at T4 representing the standard deviation value of 0.15%. Hence based on the maximum and minimum values, it is possible to observe within the variables indicating the influence of different factors on its measurement. Similar observation was made (Couto *et al.*, 2013) with three variables taken into consideration having an average CV of 6.46 and 13.77%. And standard deviation varied between 3.97% and 6.07%.

Pearson Correlation Coefficient

Total plant height (Table 5) was significantly positively correlated with number of tillers ($r = 0.548^{**}$, 0.540^{**} , 0.562^{**} , 0.549^{**}. Stalk length was significantly positively correlated with number of tillers ($r = 0.682^{**}$, 0.600^{**} , 0.600^{**} , 0.807^{**} , 0.617^{**}), and total plant height at 90 and 120 DAP (r = 0.611**, 0.568**, 0.642**, 0.599**). Number of internodes was positively correlated with stalk length (r=0.876**, 0.811***, 0.749**, 0.971**, 0.629**, 0.780**). A study by Gomati et al. (2014) reported that increase in internodal length under water logging conditions will have a better adaptation to stress. Cane diameter significantly positively correlated with number of tillers (r=0.548**, 0.576**) which indicates that the genotypes are resistant towards water deficit stress at 90 and 120 DAP. A similar observation was made by Kumar and Kumar (2014) who identified that stalk diameter had a positive association with number of internodes, cane weight, and cane yield. The positive correlation between cane diameter and stalk length (r=0.776**, 0.627**, 0.686**, 0.764**) were seen at 90 and 120 DAP. Cane diameter correlated with number of internodes (r=0.856**, 0.684**, 0.690***, 0.793***, 0.540***). A study (Smiullah et al., 2013) recorded through correlation studies that cane height was

positively significantly associated with cane diameter (0.345).

Principal Component Analysis

Principal component analysis (PCA) was performed for twenty treatments consisting of five traits at different developmental stages were simplified into four principal components whose cumulative variance contributes to 85.82%. PC1 and PC2 contribute 71.95% of the variance, and PC1 and PC3 exhibited a variance of 65.96%. The first principal component had a variation of 57.79% and cumulative variance of 57.79% whereas the variation for second principal component was 14.16% with cumulative variance 71.96% followed by the third principal component with a variation of 8.17% with cumulative variance 80.13% and nevertheless the fourth principal component with variance 5.68% having a cumulative variance of 85.82%. Figure 1 describes the Scree plot for twenty-three sugarcane cultivars on principal component 1-20.

It was observed from PC1 that cane diameter at T_1 and T_2 were found to be the most potent variables contributing 0.901 and 0.883% respectively. Whereas, total plant height at T_3 (0.936), and number of tillers at T4 (0.904) were best explained by PC2. Similarly, total plant height at T_1 (0.880) and cane diameter at T4 (0.700) were best explained by PC3. Further, number of tillers at T_1 (0.828) and T2 (0.832) were effective variables for PC4. Thus, the PCA analysis reduced data dimensionality by showing correlation between the traits and PCA analysis helps to characterize potential sugarcane cultivars for further development of water stress resistance in breeding programme (Table 6).

Agglomerative Hierarchical Clustering (AHC) Analysis

Figure 2 represents the dendrogram for twenty three sugarcane cultivars. AHC analysis categorized sugarcane varieties in to four main clusters as cluster I, cluster II, cluster III and cluster IV. Cluster I consisted of ten cultivars, i.e., Co 90003, Co 92002, Co 92013, Co 13006, Co 12007, Co 94005, Co 98008, Co 98017, Co 07015, and Co 0303. Whereas cluster II consists of nine genotypes, viz., Co 671, Co 08020, Co 14011, Co 09004, Co 85019, Co 86032, Co 95020, Co 05001, and Co 13003. Cluster III consists of one genotype namely Co 10033 and cluster IV comprises three genotypes, Co 06015, Co 92020, and Co 93009. (Tawadare et al., 2019) found that stalk length was significantly correlated with internode length followed by plant height which was in positive correlation with internode length. Ongala et al. (2016) studied the genetic diversity of sugarcane clones using 19 traits, in which they observed the first three principal components showing 80.8% of total variation. Zhou et al. (2015) reported 111 accessions through cluster analysis based on the nine qualitative traits of principal component where they considered the first four components with a cumulative variation of 74.42% and the cluster divided 111 accessions of sugarcane into high sugar and low sugar group. Another study by Raza et al. (2017) based on principal component analysis concluded that stalk height, internode length, and leaf area index had a positive correlation and hence these characters can serve as a desirable tool for improving the sugarcane yield.

CONCLUSION

As the formative phase is very sensitive, water stress caused during this phase affects the crop. Thus, through morphological parameters, one can easily identify the cultivars which are tolerant to water deficit conditions as these traits are potentially reliable. The PCA identifies four principal components contributing 57.79% of the total variation which provides necessary information with multiple correlated variables. In addition, AHC also classifies twentythree cultivars into four clusters. Based on this, the elite nine varieties such as Co 671, Co 08020, Co 14011, Co 09004, Co 85019, Co 86032, Co 13003, Co 05001 and Co 95020 have been identified as high-water stress tolerant genotypes. Hence, these sugarcane cultivars can further be considered for potential sugarcane breeding programmes under water deficit conditions for genetic improvement of sugarcane crop.

Table 1 : Morphological traits and measurement techniques used for the 23 sugarcane cultivars.

Morphological traits	Measurement technique
Number of Tiller	Total number of tillers in a row
Total Plant Height (cm)	Total Length of plant from tip of the leaf until bottom of bud
Stalk Length (cm)	Length of stalk from top to bottom
Number of Internodes	Number of internodes per stalk
Cane Diameter (cm)	Diameter of stem from top, middle, and bottom

Table 2 : Morphological	responses a	studied	(number	of tillers,	total	plant	height	and s	stalk	length)	for	23	sugarcane	cultivars
under water stress condition	ons.													

Canadama	1	Number	of Tillers	*]	Fotal Plant H	eight (cm)*	Stalk Length (cm)*					
Genotype	T1	T2	Т3	T4	T1	T2	Т3	T4	T1	T2	Т3	T4	
Co 09004	28±1.00	25±0.58	42±2.52	88±2.52	183±2.65	190.5±4.54	254±2.65	370±11.54	9±0.58	18.5±1.15	44±3.61	210±5.29	
Co 14011	29±0.58	24±1.00	43±3.21	86±6.24	171.5±1.61	186±6.03	256±4.73	372±12.86	9.5±1.04	20.5±2.05	45±3.51	199±7.00	
Co 0303	23±0.58	23±1.15	39±3.46	71±1.73	151±2.52	152.5±4.44	232±5.20	357±3.51	ND	9±1.00	39±1.44	180±1.53	
Co 13003	28±0.58	28±1.73	42±1.53	85±2.52	161±1.73	163.5±2.36	256±6.00	368±6.24	8±1.00	14±1.80	43±4.36	198±6.66	
Co 98017	23±0.58	20 ± 2.00	38±2.65	69±4.93	148.5±2.29	151±4.73	227±7.64	330±5.13	4.5±0.50	9±1.32	36±3.51	181±3.06	
Co 95020	30±0.58	23±0.58	40±3.21	85±1.73	152±2.08	154±5.51	260±5.13	373±9.54	2±0.06	14±1.00	43±3.51	202±5.57	
Co 93009	31±0.58	26±1.15	38±1.73	72±1.53	170±3.51	171±5.51	226±4.93	266±3.75	4.6±0.29	12.5±0.90	37±2.52	158±3.21	
Co 92013	28±1.15	27±1.15	39±1.73	71±2.65	172.5±2.78	173±5.03	226±6.08	334±13.58	5±0.58	9±1.00	40±4.73	188±5.20	
Co 12007	22±058	16±1.53	38±3.06	72±2.08	165.5±1.32	168±5.69	228±5.20	359±4.65	9.8±1.40	11.5±0.75	39±2.89	187±4.04	
Co 07015	16±0.58	16±1.00	35±3.06	67±6.24	150±2.52	153±10.44	225±4.36	344±6.11	ND	ND	36±3.51	182±3.61	
Co 08020	22±1.00	20±0.58	39±2.65	83±6.03	180±1.73	181±8.39	254±4.36	375±3.06	10.5 ± 1.32	20±1.32	43±4.93	208±8.08	
Co 85019	25±0.58	25±1.53	41±2.00	86±3.21	185±2.29	190±7.55	257±5.29	375±5.03	9.5±1.26	11.5±0.75	41±1.15	202±7.09	
Co 86032	12±0.58	12±0.58	40±4.36	84±5.86	142±1.53	145±2.08	256±4.58	373±11.68	ND	ND	39±4.16	199±3.21	
Co 90003	23±0.58	21±2.25	38±2.89	72±1.00	174±3.06	176±4.86	236±5.69	346±4.00	6±0.76	9±0.43	38±3.06	191±4.58	
Co 13006	16±1.53	15±1.53	35±3.06	68±1.15	165±4.04	170±4.58	231±6.24	337±12.90	5.8±0.42	9±0.50	37±4.93	188±6.03	
Co 92002	25±1.73	22±2.31	36±2.08	67±4.04	171.5±3.33	173±6.66	229±5.86	341±4.44	2.5±0.29	8±0.30	37±6.03	192±3.61	
Co 06015	13±1.00	12±1.53	34±3.61	63±2.52	141±2.65	144.5±6.71	207±4.73	309±4.51	3±0.12	5.3±0.56	33±3.06	145±6.43	
Co 92020	16±1.00	14±1.53	37±1.53	67±3.06	150±3.00	155±6.00	219±10.41	318±7.05	ND	ND	34±4.27	163±9.64	
Co 94005	20±1.53	18±3.06	38±2.08	71±3.21	164±2.89	166.5±12.98	228±6.51	348±8.14	ND	10±0.65	36±5.69	170±8.72	
Co 98008	23±0.58	22±2.08	38±3.79	70±2.31	183±1.26	188±3.12	233±7.55	351±9.29	ND	ND	35±2.31	199±14.64	
Co 05001	26±2.52	22±1.00	40±5.03	82±5.51	164±3.06	166±2.52	252±3.46	371±11.56	4±0.58	8.5±0.90	43±4.16	210±9.07	
Co 10033	28±2.08	24±3.06	37±2.00	73±4.51	220±5.69	223±9.39	228±7.64	356±8.54	8.5±0.76	17±1.32	44±4.04	223±10.54	
Co 671	27±2.52	25±2.65	44±2.31	89±3.79	193±8.19	203±7.77	259±4.73	379±6.21	11±1.73	21±0.87	48±3.21	231±7.37	

Note: 'ND' indicates not determined

^{*}90th (T₁), 120th (T₂), 150th (T₃), 270th (T₄) DAP

a i	N	umber of In	ternodes (ci	Cane Diameter (cm)*								
Genotype	T1	T2	Т3	T4	T1	T2	T3	T4				
Co 09004	2±0.00	3±0.58	7±0.58	17±1.00	2.2±0.15	2.3±0.06	2.6±0.06	3.1±0.10				
Co 14011	2±0.58	4±0.58	7±1.00	18±1.53	2.2±0.06	2.3±0.06	2.5±0.06	3±0.06				
Co 0303	ND	2±0.58	6±0.00	16±1.53	ND	2.1±0.06	2.3±0.06	2.8±0.06				
Co 13003	2±0.58	3±0.58	7±1.00	17±1.00	2±0.12	2.2±0.12	2.5±0.06	2.9±0.06				
Co 98017	1±0.00	2±0.00	6±0.58	14±1.15	2.2±0.06	2.3±0.06	2.6±0.10	3±0.10				
Co 95020	1±0.00	2±0.00	7±0.58	18±1.00	2.1±0.06	2.2±0.06	2.5±0.06	3±0.06				
Co 93009	1±0.00	2±0.00	6±0.58	16±1.53	2±0.06	2.1±0.06	2.4±0.12	2.9±0.10				
Co 92013	2±0.00	2±0.00	7±1.00	18±1.53	2.1±0.06	2.3±0.06	2.6±0.06	3±0.12				
Co 12007	1±0.00	2±0.00	6±0.58	17±1.00	2.2±0.06	2.4±0.06	2.5±0.06	3±0.00				
Co 07015	ND	ND	5±0.58	14±0.58	ND	ND	2.2±0.10	2.9±0.10				
Co 08020	2±0.00	4±0.58	7±1.15	18±1.53	2.2±0.06	2.2±0.00	2.4±0.06	2.8±0.06				
Co 85019	2±0.00	2±0.00	6±0.58	18±1.53	2.1±0.15	2.2±0.06	2.5±0.10	3.1±0.10				
Co 86032	ND	ND	6±1.00	19±1.53	ND	ND	2.3±0.15	3±0.15				
Co 90003	2±0.00	2±0.00	6±1.00	17±1.53	2.1±0.06	2.1±0.06	2.3±0.06	2.9±0.10				
Co 13006	2±0.00	2±0.00	7±1.15	16±1.53	2.1±0.10	2.2±0.06	2.4±0.06	2.9±0.15				
Co 92002	1±0.00	2±0.00	7±0.58	18±1.15	2.2±0.06	2.2±0.06	2.5±0.15	3.2±0.10				
Co 06015	1±0.00	1±0.00	5±0.58	14±1.53	2±0.06	2.1±0.06	2.3±0.10	2.7±0.06				
Co 92020	ND	ND	6±1.00	15±1.53	ND	ND	2.1±0.06	2.8±0.10				
Co 94005	ND	2±0.00	6±1.15	16±1.15	ND	2.1±0.06	2.3±0.12	2.8±0.15				
Co 98008	ND	ND	6±0.58	18±1.53	ND	ND	2.2±0.10	2.9±0.15				
Co 05001	1±0.00	2±0.00	7±1.00	18±1.53	2.2±0.06	2.3±0.06	2.6±0.06	3±0.15				
Co 10033	2±0.00	3±0.58	7±0.58	19±1.53	2.2±0.10	2.4±0.10	2.7±0.10	3.2±0.10				
Co 671	2±0.00	4±0.58	7±1.00	22±1.53	2.2±0.12	2.2±0.06	2.6±0.10	3.3±0.06				

Table 3 : Morphological responses studied (number of internodes and cane diameter) for 23 sugarcane cultivars under water

Note: 'ND' indicates Not Determined *90th (T₁), 120th (T₂), 150th (T₃), 270th (T₄) DAP

 Table 4 : Descriptive analysis for 23 sugarcane cultivars under water stress conditions.

Traits [*]	Range	Minimum	Maximum	Mean	Standard Error	Standard Deviation	Variance						
			Numbe	er of Tillers									
T1	19.00	12.00	31.00	23.22	1.14	5.48	30.00						
T2	16.00	12.00	28.00	20.87	0.99	4.74	22.48						
T3	10.00	34.00	44.00	38.74	0.53	2.56	6.57						
T4	26.00	63.00	89.00	75.70	1.73	8.30	68.86						
Total Plant Height													
T1	79.00	141.00	220.00	167.72	3.82	18.30	334.97						
T2	78.50	144.50	223.00	171.46	4.03	19.32	373.29						
T3	53.00	207.00	260.00	238.22	3.25	15.61	243.63						
T4	113.00	266.00	379.00	350.09	5.57	26.70	713.08						
			Stall	k Length									
T1	11.00	0.00	11.00	4.92	0.82	3.93	15.42						
T2	21.00	0.00	21.00	10.32	1.35	6.49	42.10						
T3	15.00	33.00	48.00	39.57	0.83	3.96	15.71						
T4	86.00	145.00	231.00	191.57	4.19	20.09	403.53						
			Number	of Internodes									
T1	2.00	0.00	2.00	1.17	0.17	0.83	0.70						
T2	4.00	0.00	4.00	2.00	0.25	1.21	1.45						
T3	2.00	5.00	7.00	6.39	0.14	0.66	0.43						
T4	8.00	14.00	22.00	17.09	0.39	1.86	3.45						
			Cane	Diameter									
T1	2.20	0.00	2.20	1.58	0.20	0.96	0.92						
T2	2.40	0.00	2.40	1.83	0.18	0.87	0.75						
T3	0.60	2.10	2.70	2.43	0.03	0.16	0.02						
T4	0.60	2.70	3.30	2.97	0.03	0.15	0.02						

 $^{90^{\text{tr}}}(\text{T}_1), 120^{\text{tr}}(\text{T}_2), 150^{\text{tr}}(\text{T}_3), 270^{\text{tr}}(\text{T}_4) \text{ DAF}$

Traits [#]		Ni	umber	of Til	lers	To	tal Pla	ant He	ight		Stalk	Lengt	h	Nur	nber o	of Inter	rnodes	С	ane Di	amete	r
		T1	T2	T3	T4	T1	T2	T3	T4	T1	T2	T3	T4	T1	T2	T3	T4	T1	T2	T3	T4
Number of Tillers	T1 T2 T3 T4	1 .931** .578** .466*	* 1 * .611** .471*	* 1 .905**	* 1																
Total Plant Height	T1 T2 T3 T4	.548** .540** .415* .142	.562 .549 .434 .184	* .284 * .353 .844 .614	.291 .347 * .961** * .716**	1 .987** .229 .258	* 1 .278 .297	1 .776 ^{**}	* 1												
Stalk Length	T1 T2 T3 T4	.475* .682** .635** .445*	.441* .600* .611 .464*	.501 [*] .600 ^{**} .807 ^{**} .619 ^{**}	.546 .617 .858 .708	.611** .568** .544** .655**	.642 ^{**} .599 ^{**} .583 ^{**} .672 ^{**}	.444 .520 .802 .744	.351 .362 * .690 * .796	1 .820 [*] .696 [*] .535 [*]	* 1 * .807* * .522*	* 1 .836*	* 1								
Number of Internodes	T1 T2 T3 T4	.509* .640** .620** .441*	.523 .596 .572 .445	.384 * .574** * .523* .645**	.448 [*] .559 ^{**} .549 ^{**} .663 ^{**}	.578 ^{***} .540 ^{***} .503 ^{**} .621 ^{***}	.593 .567 .510 .638	.388 .490 .573 .661	.246 .374 * .447* * .598**	.876 [*] .811 [*] .503 [*] .451 [*]	* .749* * .971* .655* .477*	* .629* * .780* * .732* .753*	* .485* * .518* * .682** * .816**	1 .768 [*] .617 [*] .401	* 1 * .689* .467*	** 1 * .642*	** 1				
Cane Diameter	T1 T2 T3 T4	.548** .576** .670** .497*	.430 .491 .594 .458	.230 .244 * .447* .447*	.309 .228 .458* .447*	.390 .330 .513* .603**	.386 .310 .505* * .623*	.252 .162 .381 *.425*	.073 .098 .319 .397	.776 [*] .627 [*] .662 [*] .462 [*]	* .686 * .764 * .696 .417	* .495* * .459* * .697* .615*	.324 .186 * .590** * .733**	.856 [*] .684 [*] .683 [*] .424 [*]	.690 .793 .692 .386	.540 .495 .669 .526	** .271 * .173 ** .471* ** .697**	1 .799* .780* .443*	* 1 * .740*' .261	* 1 .716*	* 1

Table 5 : Pearson's Correlation Coefficient among different morphological characters studied for twenty-three sugarcane cultivars under water stress conditions.

[#]90th (T₁), 120th (T₂), 150th (T₃), 270th (T₄) DAP

Table 6 : Principal component analysis of 23 sugarcane cultivars for morphological traits against water deficit stress.

Troita	Component										
11/2/15	1	2	3	4							
Number of Tillers											
T1	0.400	0.192	0.279	0.828							
T2	0.309	0.223	0.300	0.832							
Т3	0.183	0.816	0.086	0.420							
T4	0.246	0.904	0.101	0.203							
Total Plant Height											
T1	0.311	0.061	0.880	0.205							
T2	0.315	0.123	0.867	0.185							
Т3	0.161	0.936	0.087	0.165							
T4	0.070	0.858	0.236	-0.168							
Stalk Length											
T1	0.804	0.300	0.346	-0.009							
T2	0.773	0.372	0.216	0.297							
Т3	0.453	0.734	0.350	0.259							
T4	0.183	0.690	0.645	0.052							
Number of Internodes											
T1	0.839	0.187	0.321	0.073							
T2	0.797	0.354	0.196	0.267							
Т3	0.458	0.446	0.362	0.317							
T4	0.102	0.618	0.629	0.148							
Cane Diameter											
T1	0.901	0.032	0.169	0.121							
T2	0.883	-0.005	0.002	0.280							
Т3	0.679	0.237	0.362	0.306							
T4	0.208	0.325	0.700	0.186							
Total	11.559	2.832	1.635	1.137							
% of Variance	57.799	14.162	8.175	5.686							
Cumulative %	57.799	71.962	80.137	85.824							

 $^{r}90^{m}(T_{1}), 120^{m}(T_{2}), 150^{m}(T_{3}), 270^{m}(T_{4}) \text{ DAP}$



Fig. 1 : Scree plot describing the Eigen value of 23 sugarcane cultivars exposed to water deficit stress.



Fig. 2 : Dendrogram of cluster analysis for 23 sugarcane cultivars exposed to water deficit stress.

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Conflict of Interest

Authors have no conflict of interest.

REFERENCES

- Bordonal, R.O.; Carvalho, J.L.N.; Lal, R.; Figueredo, E.B.; Oliveira, B.G. and Scala, N.L. (2018). Sustainability of sugarcane production in Brazil - A review. Agronomy for Sustainable Development 38(2): 13.
- Cia, M.C.; Guimaraes, A.C.R.; Medici, L.O.; Chabregas, S.M. and Azevedo, R.A. (2012). Antioxidant response to water deficit by drought tolerant and sensitive sugarcane varieties. *Annals of Applied Biology*, 161(3): 313-324.
- Chidambaram, K. and Sivasubramaniam, K. (2017). Morphological Characterization and Identification of Morphological Markers for Selected Sugarcane (Saccharum spp.) Cultivars. International Journal of Current Microbiology and Applied Sciences, 6(12): 509-518
- Couto, M.F.; Peternelli, L.A.; Barbosa, M.H.P. (2013). Classification of the coefficients of variation for sugarcane crops. *Cienc Rural*, 43(6): 957-961
- Devi, K.; Gomati, R.; Kumar, R.A.; Manimekalai, R. and Selvi, A. (2018). Field tolerance and recovery potential of sugarcane varieties subjected to drought. *Indian Journal of Plant Physiology*, 23(2): 271-282.
- Dinh, T.H.; Watanabe, K.; Takaragawa, H.; Nakabaru, M. and Kawamitsu, Y. (2017). Photosynthetic response and nitrogen use efficiency of sugarcane under drought stress conditions with different nitrogen application levels. *Plant Production Science*, 20(4): 412-422.
- Ecco, M.; Santiago, E.F. and Lima, P.R. (2014). Biometric responses in young sugarcane plants subjected to water stress and aluminum. *Comunicata Scientiae*, 5(1): 59-67.
- Endres, L.; Santos, C.M.; Souza, G.V.; Menossi, M. and Santos, J.C.M. (2018). Morphological changes recorded in different phenophases of sugarcane plants subjected to water stress in tropical field conditions. *Australian Journal of Crop Science*, 12(7): 1041-1050.
- Ferreira, T.H.S.; Tsunada, M.S.; Bassi, D.; Araujo, P.; Mattiello, L.; Guidelli, G.V.; Righetto, G.L.; Goncalves, V.R.; Lakshmanan, P. and Menossi, M. (2017). Sugarcane water stress tolerance mechanism and its implication on developing biotechnology solution. *Frontier in Plant Science*, 8: 1077.
- FAO (2005) Major food and agricultural commodities and major producers. FAO, Rome.
- FNP (2009) AGRIANUAL Anurio da Agricultura Brasileria, Sâo Paulo, 497.
- Gascho, G.J. and Shih, S.F. (1983). Sugarcane. In: Teare ID, Peet MM (Ed). Crop-water relations. New York: Wiley-*Interscience*, 445-479.
- Gomathi, R. and Chandran, K. (2009). Effect of water logging on growth and yield of sugarcane clones. Sugarcane Breeding Institute (SBI-ICAR). *Quarterly News Letter*, 29(4): 1–2.

- Gomati, R.; Rao, P.N.G.; Chandran, K. and Athiappan, S. (2014). Adaptive responses of Sugarcane to waterlogging stress: an overview. *Sugar Technology*, 17(4). doi: 10.1007/s12355-014-0319-0.
- Hemaprabha, G.; Nagarajan, R. and Alarmelu, S. (2006). Parental potential of sugarcane clones for drought resistance breeding. *Sugar Technology*, 8(1): 59-62.
- Hemaprabha, G.; Swapna, S.; Lavanya, D.L.; Sajitha, B. and Venkataramana, S. (2011). Evaluation of drought tolerance potential of Elite genotypes and progenies of sugarcane (*Saccharum* sp. hybrids). *Sugar Technology*, 15(1): 9-16.
- Hussain, A.; Khan, Z.I.; Ghafoor, M.Y.; Ashraf, M.; Parveen, R. and Rashid, M.H. (2004). Sugarcane, sugar metabolism and some abiotic stresses. *International Journal of Agriculture and Biology*, 6: 732-742.
- ICRISAT (2009). Training Manual on Sustainable Sugarcane Initiative: Improving Sugarcane Cultivation in India, An Initiative of ICRISAT- WWF Project, ICRISAT, Patancheru, Andhra Pradesh, India.
- Inman-Bamber, N.G. and Smith, D.M. (2005). Water relations in sugarcane and response to water deficits. *Field Crops Research*, 92: 185–202.
- In book (2019). Recent Advances in Sugarcane Cultivation for Increased Productivity (eds. T. RajulaShanthy and Bakshi Ram (pp.1-14) Publisher: ICAR Sugarcane Breeding Institute, Coimbatore.
- Kumar, S. and Kumar, D. (2014). Correlation and path coefficient analysis in sugarcane germplasm under subtropics. *African Journal of Agricultural Research*, 9: 148-153.
- Lal, K.N.; Mehrotra, D.N. and Tandon, J.N. (1968). Growth behaviour, root extension and juice characters of sugarcane in relation to nutrient deficiency and drought resistance. *Indian Journal of Agricultural Science*, 38(5): 790–804.
- Misra, V.; Solomon, S.; Mall, A.K.; Prajapati, C.P.; Hashem,
 A.; Abd-Allah, E.F.; Ansari, M.I. (2020).
 Morphological assessment of water stressed sugarcane:
 A comparison of waterlogged and drought affected
 crop. Saudi Journal of Biological Sciences, 27(5):
 1228-1236
- Ongala, J.; Mwanga, D. and Nuani, F. (2016). On the use of principal component analysis in sugarcane clone selection. *Journal of the Indian Society of Agricultural Statistics*, 70(1): 33-39.
- Pedrozo, C.A.; John, J.; Marcio, H.P.B.; Jorge, A.D.S.; Jong-Won, P. and Nora, S.G. (2015). Differential morphological, physiological, and molecular responses to water deficit stress in sugarcane. *Journal of Plant Breeding and Crop Science*, 7(7): 225-231.
- Ramesh, P. (2000). Effect of different levels of drought during the formative phase on growth parameters and its relationships with dry matter accumulation in sugarcane. *Journal of Agronomy Crop Science*, 185(2): 83-89.
- Raza, I.; Farooq, M.A.; Masood, M.A.; Abid, S.; Anwar, M.Z.; Hassan, M. and Mustafa, R. (2017). Exploring relationship among quantitative traits of sugarcane varieties using principal component analysis. *Science*, *Technology and Development*, 36(3): 142-146.
- Silverio, J.M.; Silva, T.J.A.; Silva, E.M.B.; Iaia, A.M.; Duarte, T.F. and Pires, R.C.M. (2017). Drought

tolerance of the sugar cane varieties during the initial development. *Australian Journal of Crop Science*, 11(6): 711-715.

- Silva, M.D.A.; Silva, J.A.G.D.; Enciso, J.; Sharma, V. and Jifon, J. (2008). Yield components as indicators of drought tolerance of sugarcane. *Scientia Agricola*, 65(6): 620–627.
- Smiullah, F.; Usman, A.K. and Ijazl, A. (2013). Genetic variability of different morphological and yield contributing traits in different accession of Saccharum officinarum L. Universal Journal of Plant Science, 1: 43-48.
- Srivastava, A.K. and Rai, M.K. (2012). Sugarcane production: impact of climate change and its mitigation. *Biodiversitas*, 13(4): 214-227.
- Tawadare, R.; Thangadurai, D.; Kandagave, R.B.; Mundaragi, A. and Sangeetha, J. (2019). Phenotypic characterization and genetic diversity of sugarcane varieties cultivated in Northern Karnataka of India based on principal component and cluster analyses. *Brazilian Archives of Biology and Technology*, 62: e19180376. https://doi.org/10.1590/1678-4324-2019180376
- Zhou, H.; Yang, R.Z. and Li, Y.R. (2015). Principal component and cluster analyses for quantitative traits in GT sugarcane germplasm (*Saccharum* spp. hybrids). *International Journal Agricultural Innovation and Research*, 3(6): 1686-1690.