

IDENTIFICATION OF THE CHEMICAL PROFILE OF *RHIZOPHORA MUCRONATA* MANGROVE GREEN LEAVES FROM THE EASTERN COAST OFASAHAN, NORTH SUMATRA, INDONESIA.

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Abstract

Mangrove plants play an economically and ecologically important role and give various benefits to the people living on the eastern coast of North Sumatra, Indonesia. The purpose of the present study was to determine the nutritional value and tannin content of *Rhizophora mucronata* mangrove leaves. Proximate analysis of the green mangrove leaves revealed that they contained fat $(2.62\pm0.61\% \text{ DM})$, crude protein $(6.79\pm0.17\% \text{ DM})$, ash $(9.32\pm0.61\% \text{ DM})$, moisture $(19.95\pm0.70\% \text{ DM})$, and carbohydrate $(61.32\pm1.77\% \text{ DM})$. The tannin content of the green leaves was $2.42\pm1.39 \text{ mg/g DM}$.

Key words : Eastern Coast of Asahan, mangrove leaves, proximate composition, tanin.

Introduction

Rhizophora is believed to play a more important role compared to all the other mangrove genera in the entire eastern coast North Sumatra. *Rhizophora mucronata* (Rhyizophoraceae) is also known as the red mangrove or the Asiatic mangrove. Mangroves are a productive (Komiyama *et al.*, 2003) and dynamic ecosystem of carbon absorption (Ray and Jana, 2017). Moreover, mangroves play an important role in the protection of the coastline, in improving the water quality around the coastal environment and in supporting the coastal and marine food chains (Lee *et al.*, 2014, Naidoo, 2017, Ariyanto, 2019), higly productive ecosystem (Ariyanto *et al.*, 2018a, Ariyanto *et al.*, 2019) climate change mitigation Gilman *et al.*, 2008 and organism habitat (Ariyanto *et al.*, 2018b).

Mangroves are capable of adapting to environmental conditions, including changes in salinity (Hoppe-Speer *et al.*, 2011). Kathiresan *et al.*, (2018) revealed that the *R. mucronata* mangrove has a rough leaf characteristic with a thick wax layer which can protect against excessive ***Author for correspondence :** E-mail: dafitariyanto676@gmail.com

radiation from sunlight and can prevent transpiration, helping preserve water in the leaf's tissues. Salinity can affect plants in various ways such as causing ion toxicity, nutritional disorders, physiological dehydration, oxidative stress, metabolic process modifications, membrane incapacity and reduction of cell division (Parida and Das, 2005).

Litter, especially leaf litter, is exported from the mangrove ecosystem by tidal water both in fresh and unfresh conditions in the form of large and fine pieces of leaves which can enrich the intertidal sediment with nutrients. The compound often found in *R. mucronata* is tannin (Hardoko *et al.*, 2016, Aljaghthmi *et al.*, 2018, Ariyanto *et al.*, 2018c, Kathiresan *et al.*, 2018). Kraus *et al.*, (2003) stated that this compound is found in the leaves, roots, wood, bark and fruits of this plant. Shelaret *et al.*, (2012) revealed that this plant produces various nutritional chemical compounds which play a role in protecting against and preventing various diseases. The differences in the chemical composition of the material could be due to the influence of the environment or the

habitat and mangrove species (Nurjanah *et al.*, 2014). Chemical compounds are able to react positive effects in physiologically (Azmir *et al.* 2013). The chemical compounds found in mangrove leaves can act as antioxidant, (Reddy *et al.* 2016; Chakraborty dan Raola, 2017; Kaur *et al.* 2019), antidiabetic (Pandey *et al.* 2014; Nour *et al.* 2016; Adhikari *et al.* 2018), and anti-inflammatory agents (Chakraborty and Raola, 2017; Ray *et al.* 2017). The purpose of the present study was to determine the nutritional contents and chemical composition of *R. mucronata* mangrove leaves.

Materials and Methods

Time and Location

The study was conducted February to May 2019. Mangrove leaf samples were collected from Asahan Regency, North Sumatra. The data analysis was conducted at the SIG Laboratory, Bogor.

Sample Preparation

The leaves collected were taken to the laboratory and then classified based on their color. The leaves were rinsed using distilled water, ground using a 1mm mesh, and had a chemical analysis conducted on them.

Proximate Composition

Rhizophora mucronata leaves were analyzed for their moisture, protein, crude fat and ash contents and their extractive value using the method explained by AOAC (2005).

Tannin

An amount of 0.5g of *R. mucronata* leaf powder was boiled for 30 minutes in a 250 ml conical flask filled water until it reached a volume of 75 ml. 1ml of the product was then diluted using water until it reached a volume of 75 ml, corrected using 5 ml, Folin-Denis reagent and 10ml sodium carbonate solution. The Folin-Denis reagent was made in a dark-colored bottle by dissolving 100g of sodium tungstate and 20g phosphomolybdic acid in 750ml of distilled water, adding 50ml of phosphoric acid, making it a 1L solution by adding water after 2 hours. The intensity of the color of the mixture was assessed at 700 nm after 30 minutes. The tannin content was stated as mg tannic acid equivalent.

Statistical analysis

The statistical analysis was conducted using descriptive statistics. The sample average and deviation standard were calculated triplicate.

Results and Discussion

Table 1 shows the difference between proximate

Table 1: The difference between proximate contents of greenleaves collected on the Coast of Asahan, NorthSumatra, Indonesia.

Parameters	Green leaves
Energy from fat (Kacl/100g)	23.58 ± 5.48
Ash Content (%)	9.32±0.61
Protein Content (%)	6.79 ±0.17
Total Fat (%)	2.62±0.61
Moisture Content (%)	19.95±0.70
Carbohydrate (%)	61.32±1.77
Total Energy (Kcal/100g)	296.01±4.96
Tanin (mg / g)	2.42±1.39

contents of green leaves. Protein Content and Carbohydrate have 61.32 ± 1.77 (%) and 6.79 ± 0.17 (%). Protein Content and Carbohydrate have the relationship with the potency for rumimant of fish. The high protein has the impact for growth in fish.

Kaur *et al.*, (2019), reported that the *R. mucronata* leaves protein, fat, fiber, ash and moisture content were $11.32\pm0.35\%, 0.29\pm0.20\%, 0.78\pm0.65\%, 1.81\pm0.13\%$ and 34.91 ± 0.41 . The normal function of the digestive system depends on the presence of adequate crude fiber. Fiber aids in maintaining health and reducing cholesterol content (Bello *et al.*, 2008) and preventing diabetes (Lajide *et al.*, 2008). The moisture content is an activity index which is used as a measurement for stability and susceptibility to microbial contamination (Davey, 1989). High moisture content indicates a higher water-soluble enzyme and co-enzyme activity which is needed for metabolism. High water content in plants makes them susceptible to microbial growth and thus decomposition (Iheanacho *et al.*, 2009).

Dasgupta *et al.*, (2018) reported that the carbohydrate content of *R. mucronata Lamk* leaves is 70% mg/g. The carbohydrate content indicates the composition of various saccharides and represents the carbohydrate fraction digestible by enzymes and can be absorbed through metabolism (FAO, 2003). Carbohydrates can be a source of energy. High carbohydrate content can be considered as a potential energy source. Yadav *et al.*, (2014) added that the plant's carbohydrate content is beneficial in the immune system if it is used as a food supplement. The proportion of ash content reflects the mineral content of the food material.

The researach of tanin shows 2.42 ± 1.39 mg/g. Khattab *et al.*, (2012) reported that the tannin content of *R. mucronata* leaves ranged between 2.26 and 3.86mg/g. Kathiresan *et al.*, (2018) *also reported that Rhizophora mucronata* had a higher tannin content than *Avicennia marina* at 0.86±0.16 mg/g. Hardoko*et al.*, (2016) reported that the total tannin (mg/100g) was 110,000.00 \pm 11,422.01. Balakrishnana *et al.*, (2016) found that the tannin content of *R. mucronata* leaves ranged between 1.231 and 5.452 (mg g-1). (Cruz *et al.*, 2015) stated that the tannin content of *R. mangle* was 4.5 \pm 0.2 (mg g-1) of mangrove leaves.

Tannin is an anti-nutritional compound (cyanide, phytate, and tannin). Tannin is a plant polyphenol which has the ability to form a complex with metal ions and macromolecules such as protein and polysaccharides (Dei *et al.*, 2007, Bruyne *et al.*, 1999). Tannin is also claimed to have a negative effect on protein digestibility (Sathe and Salunkhe, 1984). Leaves undergo changes in tannin concentration during leaf development and maturation (Kandil *et al.*, 2004, Lin *et al.*, 2006).

The factors that affect the tannin content include age, reproductive development and levels of regulating components such as hormones. Mangrove leaves are photosensitive and contain a high concentration of tannin which can transform quickly (Hernes *et al.*, 2000). Various studies regarding tannin include the concentration of various vegetation types, growth stages and environmental conditions 9 (Lin *et al.*, 2006, Maie *et al.*, 2008).

The relatively high tannin content in immature leaves is unsurprising as tannin protects leaves against herbivores (Roberetson and Duke, 1987). Balakrishnana *et al.*, (2016) added that younger leaves are more susceptible to the effect of herbivores compared to older leaves. Meanwhile, a high tannin content in immature leaves is expected to provide structural support for photosynthesis and protection (Coley and Barone, 1996). When leaves reach maturity and begin to age, the tannin content declined as seen in table 1. The total content declined gradually during the orange phase, followed by a rapid decline in the brown and black phases (Lin *et al.*, 2007).

Conclusion

A high carbohydrate content indicates a rich potential energy source. The tannin content was relatively high in immature leaves.

Acknowledgements

The authors are thankful to Ministry of Research and Technology of Higher Education, Indonesia for providing fellowship for reasearch "Hibah Pekerti, 2019" and to IPB University for the excellent facilites.

References

Association of Official Analytical Chemist (AOAC) (2005). Official Method of Analysis of the Association of Official Analytical Chemist 18th Edition. Arlington: The Association of Official Analytical Chemist, Inc.

- Adhikari, A., M. Ray, T.K. Sur, S. Biswas, R.K. Roy, A.K. Hazra and A.K. Das (2018). Anti-Diabetic Activityof *Rhizophora mucronata* Leaves in Streptozotocin-Nicotinamide Induced Animal Model. *The Journal of Middle East and North Africa Sciences.*, 4(8): 1-7.
- Aljaghthmi, O., H. Heba and I.A. Zeid (2018). Bioactive Compounds Extracted from Mangrove Plants (Avicennia marina and Rhizophora mucronata): an Overview, Pathophysiology, 1-21. https://doi.org/10.1016/ j.pathophys.2018.09.002
- Ariyanto, D., D.G. Bengen, T. Prartono and Y. Wardiatno (2018a). Productivity and CNP availability in *Rhizophora apiculata* Blume and *Avicennia marina* (Forssk.) Vierh. at Banggi Coast, Central Java-Indonesia. *AES Bioflux.*, **10(3)**:137-146.
- Ariyanto, D., D.G. Bengen, T. Prartono and Y. Wardiatno (2018b). The association of *Cassidula nucleus* (Gmelin 1791) and *Cassidula angulifera* (Petit 1841) with mangrove in Banggi Coast, Central Java, Indonesia. *AACL Bioflux.*, **11(2)**: 348-361.
- Ariyanto, D., D.G. Bengen, T. Prartono and Y. Wardiatno (2018c). The relationship between content of particular metabolites of fallen mangrove leaves and the rate at which the leaves decompose over time. *Biodiversitas.* **19:** 700-705.
- Ariyanto, D. (2019). Food Preference on *Telescopium* telescopium (Mollusca : Gastropoda) Based on Food Sources in Mangrove Ecosystem. *Plant Archives*, **19(1)**: 913-916.
- Ariyanto, D., D. G. Bengen, T. Prartono, and Y. Wardiatno (2019). The Physicochemical Factors and Litter Dynamics (*Rhizophora mucronata* Lam.and *Rhizophora* stylosa Griff) of Replanted Mangroves, Rembang, Central Java, Indonesia. *Environ. Nat. Resour. J.* 17(4): 11-19.
- Azmir, J., I.S.M. Zaidul, M.M. Rahman, K.M. Sharif, A. Mohamed, F. Sahena, M.H.A. Jahurul, K. Ghafoor, N.A.N. Norulaini and A.K.M. Omar (2013).Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering.*, **117** (4): 426-436. https://doi.org/10.1016/j.jfoodeng.2013.01.014.
- Bello, M.O, O.S. Falade, S.R.A. Adewusi and N.O. Olawore (2008). Studies on the chemical compositions and antinutrients of some lesser known Nigeria fruits. *African Journal of Biotechnology.*, 7(21): 3972-9.
- Bruyne, D.T., L. Pieters, H. Deelstra and A. Ulietinck (1999). Condensed vegetable tannins: biodiversity in structure and biological activities. *Biochemisyty of System Ecology.*, 27: 445-59.
- Chakraborty, K. and V.K. Raola (2017). Two rare antioxidant and anti-inflammatory oleanenes from loop root Asiatic mangrove *Rhizophora mucronata*. *Phytochemistry.*, **135**: 160-168. oi:10.1016/j.phytochem.2016.12.013.
- Coley, P.D. and J.A. Barone (1996). Herbivory and plant defenses in tropical forests. Annual Review of Ecology Evolution and Systematics 27: 305-335. https://doi.org/ 10.1146/annurev.ecolsys.27.1.305.
- Cruz, S.M., N. Marroquín, N.E. Alvarez, D.E. Chang and A. Cáceres (2015). Evaluation of Mangrove (*Rhizophora*

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mangle L.) products as coloring, antimicrobial and antioxidant agents. *International J. of Phytocosmetics and Natural Ingredients.*, **2:**12. https://doi.org/10.15171/ jjpni.2015.12.

- Dasgupta, M., A. Ghosh and S. Mukherjee (2018). Influence of Biochemical Properties of Mangrove Leaves on Quality of Rhizosphere soils. *Environ. Sci. Ind. J.*, 14(1):165.
- Davey, K.R. (1989). A predictive model for combined temperature and water activity on microbial growth during the growth phase. *J. Appl. Microbiol.*, **65**(5): 483-8.
- Dei, H.K., S.P. Rose and A.M. Mackenzie (2007). Shea nut (*Vitellaria paradoxa*) meal as a feed ingredient for poultry. *World's Poultry Science Journal.*, **63(4):** 611-24.
- FAO (2003). Food energy: methods of analysis and conversion factors. Report of a Technical Nutrition Paper, 77.
- Gilman, E.L., J. Ellison, N.C. Duke and C. Field (2008). Threats to mangroves from climate change and adaptation options: A review. *Aquatic Botany.*, 89: 237-250. https://doi.org/ 10.1016/j.aquabot.2007.12.009.
- Hagerman, A.E. and A.E. Butler (1978). Protein precipitation method for quantitative determination of tannins *Journal* of Agriculture and Food Chemistry., **26:** 809-812.
- Hardoko, Sasmito B.B. and Y.E. Puspitasari (2016). Antidiabetic and antioxidant activities of tannin extract of Rhizophora mucronata leaves. J. of Chemical and Pharmaceutical Research., 8(3): 143-148.
- Hernes, P.J. and J.I. Hedges (2000). Determination of condensed tannin monomers in environmental samples by capillary gas chromatography of acid depolymerization extracts. *Analytical Chemistry.*, **72:** 5115-5124. https://doi.org/ 10.1021/ac991301y.
- Hoppe-Speer, S.C.L., J.B. Adams, A. Rajkaran and D. Bailey (2011). The response of the red mangrove *Rhizophora mucronata* Lam. to salinity and inundation in South Africa. *Aquatic Botany.*, **95(2):** 71-76. doi:10.1016/j.aquabot. 2011.03.006.
- Iheanacho, K., A.C. Ubebani (2009). Nutritional composition of some leafy vegetable consumed in ImoState, Nigeria. J. Appl. Aci. Environ. Manag., **13(3):** 35-8.
- Kandil, F.E., M.H. Grace, D.S. Seigler and J.M. Cheeseman (2004). Polyphenolics in *Rhizophora mangle* L. leaves and their changes during leaf development and senescence. *Trees.*, 18: 518-528. https://doi.org/10.1007/s00468-004-0337-8.
- Kathiresan, K., K. Saravanakumar, N. Asmathunisha, R. Anburaj and V. Gomathi (2018). Biochemical markers for carbon sequestration in two mangrove species (Avicennia marina and Rhizophora mucronata). Beni-Suef University Journal of Basic and Applied Sciences., 7(4): 733-739. doi:10.1016/j.bjbas.2018.10.003.
- Kaur, S, S.A.M. Yacoob, A. Venktraman, Y. Nagarajan, S. Vasudevan and B. Punniyamoorthy (2019). Proximate composition and *in vitro* antioxidant properties of *Rhizophora mucronata* plant part extract. *Asian Journal of Green Chemistry.*,3(3): 345-352. DOI: 10.22034/ ajgc.2018.143172.1091.

- Khattab, R.A., A. Gaballa, S.M. Zakaria, A.A. El-Sayed Ali, S.I. Sallam and T. Temraz (2012). Phytochemical Analysis of Avicennia marina and Rhizophora mucronata by GC-MS CATRINA., 7(1): 115-120.
- Komiyama, A., J.E. Ong and S. Poungparn (2008). Allometry, biomass and productivity of mangrove forests: a review. *Aquatic Botany.*, 89: 128-137. https://doi.org/10.1016/ j.aquabot.2007.12.006.
- Kraus, T.E.C., Z. Yu, C.M. Preston, R.A. Dahlgren and R.J. Zasoski (2003). Linking chemical reactivity and protein precipitation to structural characteristics of foliar tannins. *Journal of Chemical Ecology.*, **29**: 703-730. https://doi. org/10.1023/A:1022876804925.
- Lajide, L., M.O. Oseke and O.O. Olaoye (2008). Vitamin C, fiber, lignin and mineral contents of some edible legume seedlings. *J. of Food and Technology.*, **6(6)**: 237-41.
- Lee, S.Y., J.H. Primavera, F. Dahdouh Guebas, K. McKee, J.O. Bosire, S. Cannicci, K. Diele, F. Fromard, N. Koedam and C. Marchand (2014). Ecological role and services of tropical mangrove ecosystems: a reassessment. *Global Ecology* and Biogeography., 23: 726-743. http://dx.doi.org/10.1111/ geb.12155.
- Lin, Y.M., J.W. Liu, X. Ping, P. Lin, Z.H. Ding and L.S.H. Sternberg (2007). Tannins and nitrogen dynamics in mangrove leaves at different age and decay stages (Jiulong River Estuary, China). *Hydro. biologia.*, **583**: 285-295. https://doi.org/10.1007/s10750-006-0568-3.
- Lin, Y.M., J.W. Liu, P. Xiang, P. Lin, G.F. Ye and L.D.S.L. Sternberg (2006). Tannin dynamics of propagules and leaves of *Kandelia candel* and *Bruguiera gymnorrhiza* in the Jiulong River Estuary, Fujian, China. *Biogeochemistry.*, **78:** 343-359. https://doi.org/10.1007/s10533-005-4427-5.
- Maie, N., O. Pisani and R. Jaffe (2008). Mangrove tannins in aquatic ecosystems: Their fate and possible influence on dissolved organic carbon and nitrogen cycling. *Limnology* and Oceanography., 53(1): 160-171. https://doi.org/10.43 19/lo.2008.53.1.0160.
- Mangrio, A.M., M. Rafiq, S.H.A. Naqvi, S.A. Junejo, S.M. Mangrio and N.A. Rind (2016). Evaluation of Phytochemical Constituents And Antibacterial Potential Of Avicennia Marina And Rhizophora Mucronata From Indus Delta Of Pakistan. *Pakistan Journal of Biotechnology.*, 13(4): 259-265.
- Naidoo, G. (2017). The mangroves of South Africa: an ecophysiological review. *South African of Journal Botany.*, **107:** 101-113. http://dx.doi.org/10.1016/j.sajb.2016.04.014.
- Nour, A., J. Nitthiya and S. Omer Manal (2016). The Potential of *Rhizophora mucronata* in Extracting the Chemical Composition and Biological Activities as Mangrove Plants: A Review. *Australian Journal of Basic and Applied Sciences.*, **10(4)**: 114-139.
- Nurjanah, A. Abdullah and S. Sudirman (2014). Antioxidant activity and active components water spinach (*Ipomoea aquatica* Forsk.). Journal of Innovation and Entrepreneurship., **3(1):** 68-75.

- Pandey, A.K., P.P. Gupta and V.K. Lai (2014). Hypoglycemic effect of *Rhizophora mucronata* in streptozotocin induced diabetic rats. *Journal of Complementary Integrative Medicine.*, **11(3)**: 179-183. doi:10.1515/jcim-2012-0057.
- Parida, A.K. and A.B. Das (2005). Salt tolerance and salinity effects on plants: A review. *Ecotoxicology and Environmental Safety.*, 60(3): 324-49. https://doi.org/ 10.1016/j.ecoenv.2004.06.010.
- Ray, M., A. Adhikari, T.K. Sur, S.E. Besra, S. Biswas and A.K. Das (2017). Evaluation Of Anti-Inflammatory Potential Of Ethanolic Extract Of The Leaves Of *Rhizophora* mucronata, A Sunderban Mangrove. International Journal of Research and Development in Pharmacy and Life Sciences., 6(1): 2506-2516.
- Ray, R. and T.K. Jana (2017). Carbon sequestration by mangrove forest: one approach for managing carbon dioxide emission from coal-based power plant. *Atmospheric Environment.*, **171:** 149-154. https://doi.org/10.1016/j.atmosenv.2017. 10.019.

- Reddy, A.R.K. and J.R. Grace (2016). *In vitro* evaluation of antioxidant activity of methanolic extracts of selected mangrove plants. *Med. aromat. plants.*, 5: 2167-0412. doi:10.4172/2167-0412.1000250.
- Robertson, A.I. and N.C. Duke (1987). Insect herbivory on mangrove leaves in North Queensland. *Aust. J. Ecol.*, **12**: 1-7. https://doi.org/10.1111/j.1442-9993.1987.tb00921.x.
- Sathe, S.K. and D.K. Salunkhe (1984). Technology of removal of unwanted components of dry bean. *Crit. Rev. Food Sci. Nutr.*, **21**:263-86.
- Yadav, M., S. Chatterji, S.K. Gupta and G. Watal (2014). Preliminaryphytochemical screening of six medicinal plants used in traditional medicine. International *Journal of Pharmacy and Pharmaceutical Sciences.*, 6(5): 539-542.