



Plant Archives

Journal home page: www.plantarchives.org

DOI Url: <https://doi.org/10.51470/PLANTARCHIVES.2021.v21.no1.204>

SEASONAL DISTRIBUTION OF PHYTOPLANKTON DIVERSITY AND PRIMARY PRODUCTION IN A FRESH WATER POND AT KUMBAKONAM, TAMIL NADU, INDIA

Durairaju, R. and Sivakami, R.*

Arignar Anna Government Arts College, Musiri - 621 211, Tamil Nadu, India

(Affiliated to Bharathidasan University- Tiruchirappalli - 620 024)

*Email: drsiva17@gmail.com

(Date of Receiving-15-01-2021; Date of Acceptance-04-04-2021)

ABSTRACT

Measurement of primary production is essential for assessing the level of fish production and potential of exploitable fisheries. Hence the present study was focused on assessing the seasonal distribution of phytoplankton, its diversity and primary productivity in a fresh water system located in Kumbakonam, Tamil Nadu. The present study could record 4 groups of phytoplankton, Cyanophyceae, Chlorophyceae, Bacillariophyceae and Euglenophyceae. Among these Cyanophyceae was found to be the most dominant group in terms of planktonic count in all the three seasons followed by Chlorophyceae. Further, Cyanophyceae and Chlorophyceae preferred presummer to record their highest counts while Bacillariophyceae and Euglenophyceae preferred post summer season. The Shannon's index of species diversity, diversity index of family and species richness indices were highest during the presummer and lowest during the rainy season. Similarly among the 3 seasons, GPP was highest during the presummer season.

Keywords: Seasonal Distribution, Phytoplankton Diversity, Species Diversity Indices and Productivity

INTRODUCTION

Water is one of the most important natural resources that are required for the life and health of organisms. The unique peculiarity of an aquatic ecosystem is its inhabitation by planktonic organisms (Umavathy *et al.*, 2007). Phytoplankton being autotrophs initiate the aquatic food chain and the secondary producers (Zoo plankton) and tertiary producers depend on them directly or indirectly for food (Thirunavukkarasu *et al.*, 2013). Phytoplanktonic study is a very useful tool for the assessment of water quality (Jiji Joseph, 2017) and natural regions which are characterized by typical species or species groups (Sampathkumar and Ananthan, 2007). On addition phytoplankton plays a significant role in the global biogeo-chemical cycling of many nutrients (Thirunavukkarasu, *et al.*, 2013). Phytoplankton has also been recently used as an indicator to observe and understand changes in the ecosystem because it seems to be strongly influenced by climatic factors (Li *et al.*, 2000; Soni and Thomas, 2014). Further, phytoplankton plays a major role in aquaculture as they serve as food for the larval stages of crustaceans, fish and all stages of bivalves (Volkman *et al.*, 1989).

One of the best ways to feed the increasing Indian population is to use fish which is a source of cheap animal protein. Hence the measurement of primary production becomes essential for assessing the level of fish production and potential exploitable fisheries as suggested by Thirunavukkarasu *et al.*, (2013). Hence the present study was focused on assessing the seasonal population density,

diversity and primary production in a fresh water pond at Kumbakonam, Tamil Nadu.

MATERIAL AND METHODS

Study Area

The aquatic system chosen for the present investigation is a pond situated in Kumbakonam District, Tamil Nadu, India and referred to as fish culture pond. The pond is located at Swamimalai at an elevation of about 80MSL and it is mostly perennial.

It has a water spread area of about 0.5 ha and a depth of about 4m when full. As the pond is close to the Cauvery River; it is currently used for agriculture, cattle bathing and building constructions. On the other hand, environmental degradation is felt intensely in this area (Jayaram, 2000; Kalavathy *et al.*, 2011). Thus this system is prone to more pollution.

Physico – Chemical Variables

Water samples were drawn from surface and bottom and stored in separate polyethylene bottles for later analyses in the laboratory. While some physico-chemical variables like estimation of dissolved oxygen (DO), hydrogen ion concentration (pH), free carbon dioxide (free CO₂), phenolphthalein alkalinity (PPA) and methyl orange alkalinity (MOA) were analysed in the field itself, all other variables were analysed in the laboratory, Duplicate sample of all variables were taken and the average values taken.

The atmospheric, surface and bottom water temperatures were measured using a centigrade mercury thermometer calibrated to 100°C. Atmospheric temperature was measured in shade, while surface water temperature was analysed by taking the surface water in a container and then measuring it. The water level of the pond was measured using a graduated rope provided with a weight at one end. The measurement was done on every sampling day at a particular spot. While the transparency of the water column was measured using a Secchi's disc, total dissolved solids (TDS) was estimated by evaporating the water samples in a porcelain dish (Saxena, 1987); dissolved oxygen (DO) was estimated using unmodified Winkler's method (Ellis *et al.*, 1984). While free carbondioxide (free CO₂) and alkalinity (phenolphthalein and methyl orange) were determined according to Saxena (1987), pH was measured in the field itself with a digital pH pen (Hanna) and electrical conductivity was measured using a water analysis kit (Elico). Nutrients like phosphate, silicate, ammonia-nitrogen, nitrite-nitrogen, sulphate, calcium and magnesium were estimated according to APHA (1995). Nitrate-nitrogen (NO₃-N) was estimated after Mackereth (1961) and chloride after Strickland and Parsons (1972). While oxidizable organic matter, nitrogenous organic matter and suspended solids were done following APHA (1995), Trivedy and Goel (1986) and Taylor (1949), biological oxygen demand (BOD) was estimated following the procedure of Sawyer and Bradney (1946) and chemical oxygen demand (COD) as per Moore *et al.*, (1949).

Phytoplankton Analysis

Surface water samples were collected with the help of a satin net (pore diameter 4.5µ) fitted to an aluminum frame between 7:00 and 8.00 am for a period of one year (2019). Collection was done on a monthly basis. The Sample were immediately transferred to glass containers for later microscopic analyses. Lugol's solution was also added as a preservative. Care was also taken to observe some fresh samples. The counting of algae was done using a Sedgwick-Rafter Counting cell (Saxena, 1987). Samples were isolated and identified by standard methods (Pearsall *et al.*, 1946; Desikachary, 1959; Starmach, 1966; Pennak, 1978; Rippka *et al.*, 1979; Prescott *et al.*, 1982; Adoni and Vaishy, 1985; Trivedy *et al.*, 1987; Sridharan 1989) In addition, diversity indices and productivity were also calculated following Trivedy *et al.*, (1987). Finally, the results obtained in the present study were statistically treated for a meaningful discussion.

RESULTS AND DISCUSSION

Table 1 shows the various physicochemical factors that were analysed along with the ranges during the three seasons of the year. Table 2 records the various phytoplankton that were recorded during the three seasons.

As evident from the Table, Phytoplankton that occurred in the system belonged to Cyanophyceae Chlorophyceae, Bacillariophyceae and Euglenophyceae.

On the whole, a total of 21 species were noticed for the three seasons. A comparison of the 3 seasons (May-Aug.) record 20 species each while the rainy season (Sep. – Dec.) recorded only 16 species.

A groupwise comparison reveal that cyanophyceae was recorded by 5 species during presummer and rainy season and only 4 species during the post summer season as *Synecocystis aquatilis* was absent during this period. Among the cyanophycean species, *Microcystisaeruginosa* was found to dominate all the three periods.

The group Chlorophyceae was represented by 9 species were found to occur during the presummer period, the post summer period was recorded by 8 species as *Volvox aureus* was absent. However, the rainy season was recorded by only 7 species as *Chlorella vulgaris* and *Chlamydomonas simplex* were absent. During the presummer seasons in terms of count *C. vulgaris* was found to dominate why in post summer *C. simplex* dominated and in rainy season *Eudorina elegans* dominated.

Bacillariophycene was represented by 4 species during the period of study. All the four species were recorded during the post summer season while only 3 were found during the presummer season as *Pinnularia major* was absent. The rainy season recorded only 2 species as *Nitzhia sigmoidea* and *P. major* were absent during this period. Among this group, *Cyclotella compta* was found to dominate during presummer and rainy season while in post summer, *Synedra ulna* was found to dominate in terms of count.

Euglenophyceae was represented by 3 species during the presummer and post summer periods while the rainy season recorded only 2 species as *Trachelomonas hispida* was absent. *Euglena viridis* was found to dominate during presummer and post summer seasons while *Phacus suecica* dominated rainy season in terms of count.

Cyanophycean class count was found to vary between 200 (rainy) to 470 i/l (presummer) while Chlorophycean count ranged between 160 (presummer) and 390 i/l (presummer) and Bacillariophycean count from 30 (rainy) to 100 i/l (post summer). Euglenophycean count on the other hand ranged between 30 (rainy) to 200 i/l (postsummer). Thus in terms of group count Cyanophyceae and Chlorophyceae preferred presummer while Bacillariophyceae and Euglenophyceae preferred post summer to record their highest class counts. Phytoplankton total count reveals that the count ranged between 420 to 980 i/l. While the highest count was recorded during the presummer season the lowest was recorded in the rainy season. (Table 1-2) The comparison of the percentage composition in

Table 1: Physico – Chemical Analysis of water of a pond at Kumbakonam

S.No	Physico-chemical Analysis	Unit	Ranges	Average
1.	Atmospheric Temperature	°C	28 - 37	32.5
2.	Water Temperature	°C	26.0 - 34.0	30
3.	pH	unit	7.0 - 8.8	7.9
4.	Dissolved Oxygen	mg/l	4.7 - 7.9	6.3
5.	Carbondi Oxide	mg/l	0.5 - 1.4	0.95
6.	Alkalinity	mg/l	180 - 290.6	253.3
7.	Total Dissolved Solids	mg/l	160 - 270	90
8.	Phosphate-P	mg/l	1.2 - 2.4	1.8
9.	Nitrate-N	mg/l	0.96 - 1.8	1.38
10.	Ammonia-N	mg/l	0.11 - 0.26	0.37
11.	Calcium	mg/l	40.6 - 72.0	56.3
12.	Magnesium	mg/l	16.7 - 24.5	41.2
13.	Chloride	mg/l	26.5 - 42.5	69
14.	BOD	mg/l	13.5 - 29.5	21.5
15.	COD	mg/l	25.5 - 38.6	32.05
16.	Water level	cm	140 - 420	280
17.	Transparency	cm	48 - 69	58.5

terms of class count for the three periods reveals that Cyanophyceae dominated in all the three seasons followed by Chlorophyceae. However, the percentage composition was the same for Bacillariophyceae and Euglenophyceae for presummer season while in the post summer Euglenophyceae dominated over Bacillariophyceae.

In the present study, 4 groups of phytoplankton belonging to Cyanophyceae, Chlorophyceae, Bacillariophyceae and Euglenophyceae were recorded. A perusal of literature reveals that many workers (Devi and Singara, 2007; Rout and Borah, 2009; Mahor and Singh, 2010; Jiji Joseph, 2017) have also reported the presence of the above groups in the various aquatic systems analysed.

Among the various groups, Cyanophyceae was the most dominant member in all the three seasons. Literature reveals that similar observations were also noticed by others in other systems of India (Santhosh *et al.*, 2007; Hudder, 1995, Hujare, 2008; Khanna and Indu, 2009; Jeyabaye, 2010; Arumugam, 2017). Ganapati *et al.*, (1953) suggested that Cyanophyceae dominates while temperature, pH, alkalinity, silicate and phosphate levels are high. Neelam (2009) reported that the dominant native of Cyanophycean members are the characteristic features of eutrophic environments as they have high concentrations of nutrients especially phosphate and nitrate.

In the present study *M. aeruginosa* was the most dominant Cyanophyceae. Literature reveals that this species has been reported to be the commonest algal to occur in almost all tropical ponds including many fresh water systems of Tamil Nadu (Ganapati, 1955, Sreenivasan, 1968; Nandan and Patel, 1983; Sivakami, 1996; Sirajunisa, 2014).

Literature reveals that chlorophytes prefer different seasons in different aquatic systems. However, Melack *et al.*, (1982) reported that phosphate influenced their growth more than nitrate. While Pearsall (1932) suggested that they occurred when nitrate and phosphate were moderate. However, Kavitha *et al.*, (2005) suggested that low temperature during monsoon coupled with average pH and average nutrient content favors the growth of Chlorophyceae.

In the present study, Bacillariophyceae was recorded in high numbers during the post summer season. Similar observation were also recorded by Kastoori bai (1991); Kundangar and Zutshi (1985) and Hegde and Bharti (1985). According to Munawar (1970) regular supply of nutrients encouraged the growth of diatoms while Ansari (2015) reported that the presence of phosphate, nitrate, silicate and total hardness promoted their growth. However, Wetzel (1983) reported that of all the aspects of chemical determination of succession and productivity, the negative relationship between diatoms and silicate contraction is among the most apparent.

In the present study, Euglenophyceae recorded maximum count during the post summer season. Similar report was suggested by Jiji Joseph (2017). Jasprica *et al.*, (2006) reported that higher temperature and nitrate content increased the growth of Euglenophyceae while Ansari *et al.*, (2015) suggested that high temperature chloride; TDS and BOD play an important role in their growth.

The various indices of phytoplankton diversity are presented in Table 3. While the Shannon's index of species diversity was found to range while between 0.94 (rainy)

Table 2: Seasonal changes of Phytoplankton in a Freshwater pond at Kumbakonam

S.No	Phytoplankton	Rainy Season Sep.-Dec.	Pre Summer Jan.-Apr.	Post Summer May-April
	Cyanophyceae			
1.	<i>Anabena circinalis</i>	10	40	20
2.	<i>Microcystis aeruginosa</i>	140	310	230
3.	<i>Chroococcus giganteus</i>	10	30	60
4.	<i>Spirulina major</i>	30	70	40
5.	<i>Synechocystis aquatilis</i>	10	20	-
	Class Count	200	470	350
	Percentage Occurrence	47.61	47.96	41.18
	Chlorophyceae			
1.	<i>Chlorella vulgaris</i>	-	120	20
2.	<i>Pediastrum duplex</i>	20	30	10
3.	<i>Scenedesmus obliquus</i>	20	40	50
4.	<i>Ankistrodesmus falcatus</i>	30	20	10
5.	<i>Ulothrix zonata</i>	10	10	10
6.	<i>Chlamydomonas simplex</i>	-	40	60
7.	<i>Zygnema stellinum</i>	20	30	10
8.	<i>Eudorina elegans</i>	40	70	40
9.	<i>Volvox aureus</i>	20	30	-
	Class Count	160	390	210
	Percentage Occurrence	38.09	39.79	24.71
	Bacillariophyceae			
1.	<i>Nitzschia sigmoidea</i>	-	10	30
2.	<i>Synedra ulna</i>	10	20	30
3.	<i>Cyclotella comta</i>	20	30	10
4.	<i>Pinnularia major</i>	-	0	20
	Class Count	30	60	90
	Percentage occurrence	7.14	6.12	10.59
	Euglenophyceae			
1.	<i>Euglena viridis</i>	10	20	130
2.	<i>Phacus suecica</i>	20	30	40
3.	<i>Trachelomonas hispida</i>	-	10	30
	Class count	30	60	200
	Percentage Occurrence	7.14	6.12	23.53
	Total Count	420	980	850

Table 3: Seasonal changes of Phytoplankton diversity in a Freshwater pond at Kumbakonam

S.No	Details	Shannan Weiner's Diversity Index of species	Shannan Weiner's Diversity Index of Family	Species Richness Index (d)
1.	Rainy/ winter season	0.94	1.14	1.32
2.	Presummer season	1.44	1.49	1.84
3.	Postsummer season	1.26	1.20	1.60

Table 4: Seasonal changes of Phytoplankton productivity in a Freshwater pond at Kumbakonam

S.No	Details	GPP gO ₂ /m ² /day	NPP gO ₂ /m ² /day	CR gO ₂ /m ² /day
1.	Rainy/ winter season	35.4	22.5	12.9
2.	Presummer season	42.2	34.6	7.6
3.	Postsummer season	38.6	29.9	9.0

and 1.44 (summer), the diversity index of family ranged between 1.14 (rainy) and 1.49 (post summer) and species richness index from 1.32 (rainy) to 1.89 (post summer). According to Odum (1971) higher diversity means larger food chains and greater possibilities for negative feedback control which in turn will increase stability.

The details of productivity are presented in Table 4. The GPP was found to range between 35.4 (rainy) to 42.2g O₂/M²/day (presummer) while NPP ranged between 22.5 (rainy) and 34.76g O₂/M²/day (presummer) and CR from 7.62 - 12.09g O₂/M²/day.

In the present study minimal GPP was recorded in the rainy season. This is in conformity with the report of Sreenivasan (1964) who suggested that productivity decreased during the cooler months in addition to deterioration of phytoplankton. Goldman and Wetzel (1963) attributed temperature as an important factor in determining seasonal productivity while Sultan *et al.*, (2003) reported that temperature, Solar radiation and available nutrients are important limiting factors contributing to seasonal variation in any aquatic system. Kund Hansen (1997) opined that productivity is influenced by availability of nutrients while Lin *et al.*, (1997) suggested that nitrogen and phosphorus are mostly responsible for algal growth while Umavathi *et al.*, (2007) reported that in a producture system, respiration accounts for a large proportion of GPP.

REFERENCES

- Adoni, A.D., Vaishya, A.K. 1985. Phytoplankton productivity: Seasonal, diel and vertical periodicity in a central Indian reservoir. *Bull. Bot. Soc. Sagar.*, 32: 219-228.
- Ansari, 2015. Limnological Studies with reference to Phytoplankton Diversity in Ponds of Semi arid zone of Western Uttar Pradesh. *Biological Forum – An International Journal* 9(2): 129-147.
- APHA (1995). Standard methods for the examination of water and wastewater. 21st edition. American Public Health Association, Washington, USA, p. 2240.
- Arumugam, S, (2017) Assemblages of phytoplankton diversity in different zonation of Muthupet mangroves. *Reg Stud Mar Sci* 3:234–241
- Desikachary, T.V. 1959. Cyanophyta: Monographs in algae. Indian Council of Agricultural Research, New Delhi.
- Devi Anita, U., and Singara Charya, M.A., 2007. Phytoplankton in lower Manair dam and Kakatica canal, Karimnager. *Nat. Environ. Pollu. Tech.* 6(4): 643-648.
- Ellis, M. M., Westyfall, B. A. and Ellis, M. D. (1984). Determination of water quality. *Fish and Wild Life Services*. US Department, Interior Res. Dep., 9: 122
- Ganapati, S.V. 1955. The ecology of a temple tank containing permanent bloom of *Microcystis aeruginosa* (Kutz). *J. Bom. Nat. Hist. Soc.* 42(1) : 65-77.
- Ganapati, S.V., Chacko, P.I., Srinivasan, R. 1953. Hydrobiological conditions of the Gangadhareswarer Temple Tank, Madras. *J. Asiatic. Soc. Sci.*, 19: 149-158.
- Hegde, G.R., Bharti, S.G. 1985. Comparative phytoplankton ecology of freshwater ponds and lakes of Dharwad, Karnataka State, India.
- Hudder, B.D., 1995. Hydrobiological studies in lentic freshwater bodies of Hubli Ph.D. Thesis. Submitted to Karnataka University, Dharwad, pp. 267.
- Hujare Milind, S., 2008. Seasonal variation of phytoplankton in the freshwater tank of Talsande, Maharashtra. *Nat. Envir. And Pollu. Tech.* 7(1): 147- 150.
- Jasprica N., Hafner D., Cari M., Rimac A. 2006. A preliminary investigation of phytoplankton of karstic pools (Dugi otok island, Croatia). *Acta Bot. Croat.* 65 (2), 181–190.
- Jayabhaye, U.M., 2010. Studies on Phytoplankton diversity in Sawana Dam, Maharashtra, India. *Samiksha Aur Mulyankan.*, 11(11-12): 40-42.
- Jayaram, K. C. (2000). Kaveri Riverine System: An Environmental Study. Madras Science Foundation, Chennai. pp. 131-144.
- Jiji Joseph. 2017. Diversity and distribution of phytoplankton in an artificial pond. *Int. J. Adv. Res. Biol. Sci.* 4(5): 114-122.
- Kalavathy, S., Rakesh Sharma, T. and Sureshkumar, P. (2011), Water Quality Index of River Cauvery in Tiruchirappalli district, Tamil Nadu, *Arch. Environ. Sci.*, 5: 55-61.
- Kastooribai, R.S. 1991. A comparative study of two tropical lentic systems in the context of aquaculture. Ph. D.

Pp. 120.

Thesis, University of Madras, India.

The composition of the phytoplankton in relation to dissolved substances. *J. Ecol.*, 20: 241-262.

- Kavitha, A., Regini Balasingh, G.S. and Raj, A. D.S., 2005. Ecology of two temple ponds of Kanyakumari district. *Ind. Hydrobiol.*, 8 (1): 61-65.
- Khanna, V. and Indu. Y., 2009. Planktonic diversity in the holy lake of Pushkar Ajmar. *Nat. Envir. Pollu. Tech.*, 8(2): 339-342.
- Knud-Hansen CF (1997). Experimental design and analysis in aquaculture. In Egan HS, Boyd CE (eds) Dynamics of pond aquaculture: CRS Press Boca - Raton New York, pp. 325-375.
- Kundangar, M.R.D. and Zutshi, D.P. 1985. Environmental features and plankton communities of two Himalayan rural lakes. *Proc. Nat. Symp. Pure and Applied Limnology.* (ed) Adoni, *Bull.Bot.Soc.Sagar.* 32: 40-47.
- Li, M. A., Gargett and Denman, K., 2000. What determines seasonal and inter-annual variability of phytoplankton and zooplankton in strongly estuarine systems? Application to the semi- enclosed estuary of Strait of Georgia and Juan de Fuca Strait. *Estuarine, Coastal and Shelf Science*, 50: 467-488.
- Lin CK, Teichert-Coddington DR, Green BW, Veverica KL (1997). Fertilization regimes. In Egan HS, Boyd CE (eds) Dynamics of Pond Aquaculture: CRC Press, Boca Raton - New York, p. 73-107.
- Mackereth, F. J. H. (1961). Water analysis for limnologists. Freshwater Biological Association, Ambleside, West Morland.
- Mahor R.K. and Singh B., 2010. Diversity and seasonal fluctuation of phytoplankton in fresh water reservoir Igra Gwalior (M.P). *Int. Res. J.*, 1(10): 51-52.
- Melack, J. M., P. Kilham, and T. R. Fisher. 1982. Responses of phytoplankton to experimental fertilization with ammonium and phosphate in an African soda lake. *Oecologia* 52: 321-326.
- Moore, W. A., Croner, R. C. and Ruchhoft, C. C. (1949). Dichromate reflux method for determination of oxygen consumed. *Anal. Chem.*, 21: 953.
- Munawar, M. 1970. Limnological studies on freshwater ponds of Hyderabad-11. *Hydrobiologia*. 36: 105-128.
- Nandan, S. N. and Patel, R. J. (1983). Phytoplankton and physiochemical parameters used as indicators of eutrophication. *J. Plant Nature*, 2: 17-22.
- Neelam, V., B.Avinash, and Dwivedi, S.N., 2009. Planktonic biodiversity of Bhoj wetland, Bhopal, India. *J.Appl. Sci. Environ. Manage.*, 13(4): 103- 111.
- Odum, E. P., 1971. Fundamentals of ecology. Saunders, Philadelphia. pp. 546.
- Pearsall, W.H. 1932. Phytoplankton in the English lakes. II. The composition of the phytoplankton in relation to dissolved substances. *J. Ecol.*, 20: 241-262.
- Pearsall, W.H., Gartiner, A.C., Greenshields, F. 1946. Freshwater biology and water supply in Britain. *Freshwater Biol. Assoc. Br. Empire Publ.*, 11: 1-90.
- Pennak, R.W. 1978. Freshwater invertebrates of the United States, 2nd edn. John Wiley and Sons, New York. 803 Pp.
- Prescott, G.W., Bicudo, C.E.M., Vinyard, W.C. 1982. A synopsis of North American Desmids. 2: University of Nebraska Press, Lincoln. 700 Pp.
- Rippka, R.J., Deruelles, J.B., Waterberry, M., Herdman, M., Stainer, R.Y. 1979. Generic assignments, Strain Histories and Properties, Pure Cultures of Cyanobacteria. *J. Gen. Microbiol.*, 111: 1-61.
- Rout J. and Borah, D., 2009. Algal diversity in Chatla wetland in Cachar District (Southern Assam). *Assam Uni.J. of Sci and Technology: Biol., Sci.*, 4(1): 46 – 55.
- Sampathkumar, P. and Ananthan, G. (2007). Phytoplankton. In: Coastal Biodiversity in Mangrove Ecosystems, UNU-INWEH-UNESCO- International Training Course Manual. 205-212.
- Santhosh, S.V., S. Shoba, Girijalakshmi and Thara, J.C., 2007. Phytoplankton abundance of Paravarkappil backwater of Kerala. *Seaweed. Res. Util.*, 29 (112): 155-163.
- Sawer, C. N. and Bradney, L. (1946). Modernization of BOD test for determining the efficiency of the sewage treatment process. *Sewage Works J. USA*, 18: 113.
- Saxena, D. (1987). *Soil water and waste water analysis*. New Delhi Publication. p. 283.
- Sirajunisa, V. (2014). Limnochemical and Biological Diversity of Aathivayal Lake, Kottaiappattinam, Pudukkottai District, with Special Reference to Bioremediation of Heavy Metals. Ph.D. Thesis, Bharathidasan University, Tiruchirappalli, Tamil Nadu.
- Sivakami, R. 1996. Limnological profile of two contrasting lentic systems and their aquaculture Potential. Ph.D. Thesis, Bharathidasan University, Tiruchirappalli, India.
- Soni, H.B. and Thomas, S., 2014. Associative dependence among plankton and macrophytes as pollution markers at tropical lentic environment, Gujarat, India. *International Journal of Environment*. 3 (2): 175-191.
- Sreenivasan, A . 1964. Limnological features of an primary production in a polluted Moat at Vellore, Madras State . *Environmental Health* 6 : 237-245 .
- Sreenivasan, A. 1968. Limnology of tropical impoundments. IV. Studies of two hard water reservoirs in Madras State. *Arch. F. Hydrobiol.*, 65: 205-222.
- Pearsall, W.H. 1932. Phytoplankton in the English lakes. II.

- Sridharan, V.T. 1989. Phytoplankton and algae studies. Techniques of plankton methodology. Prepared for Training workshop on Integrated Environmental Research programme on Kaveri River. Pp. 1-15.
- Starmach, K. 1966. Cyanophyta - Since Galucophyta - Gluokofity. Flora Slodowokan Polsi, Tom 2. Warszawa, 800 Pp.
- Strickland, J. D. H. and Parsons, T. R. (1972). A practical handbook of sea water analysis. *Bull. Fish. Res.*, 167: 310.
- Sultan, S., Chouhan, M. and Sharma, V.I. (2003): Physico-chemical status and Primary productivity of Pahunj reservoir, Uttar Pradesh. *Journal of the Inland Fisheries Society of India* 35: 73-80.
- Taylor, E. (1949). The examination of waters and water supplies (Thresh, Beal and Suckling). 6th ed. J & A Churchill Ltd., London, p. 340.
- Thirunavukkarasu, K., Soundarapandian, P., Varadharajan, D. and Gunalan, B. 2013. Zooplankton Composition and Community Structure of Kottakudi and Nari Backwaters, South East of Tamil Nadu. *J. Environ. Anal. Toxicol.* 4: 200(10) : 4172/2161-0525.1000200.
- Trivedy, R. K. and Goel, P. K. (1986). Chemical and Biological methods for water pollution studies. *J. Ecophysiol.*, 37: 77-86.
- Trivedy, R.K., Goel, P.K., Trisal, C.K. 1987. Practical methods in ecology and environmental science. Environmental Publication, Karad, India. Pp. 380.
- Umavathi, S., Longakumar, K. and Subhashini. D. 2007. Studies on the nutrient content of Sular pond in Coimbatore, Tamil Nadu, *Journal of Ecology and Environmental Conservation*, 13(5), pp. 501-504.
- Volkman, J.K., Jeffrey, S.W., Nichols, P.D., Rogers, G.I. and Garland, C. D. 1989). Fatty Acid and Lipid Composition of 10 Species of Microalgae Used in Mariculture. *J. Exp. Mar. Biol. Ecol.* 128:219-240.
- Wetzel, R.G. 1983. Limnology, 2nd edn. Saunders College Publishing Co., New York. 767 Pp.