



## ANTIFUNGAL ACTIVITY OF *TAGETES ERECTA* EXTRACT AND *TRICHODERMA HARZIANUM* ON THE PATHOGENIC FUNGUS *FUSARIUM VERTICILLOIDES*

Abdulzahra Jabar Ali<sup>1</sup>, Ali Faraj Jubair<sup>2</sup> and Mushtak Talib Mohammadali<sup>3</sup>

<sup>1</sup>Ministry of Higher Education and Scientific Research-Official, Research and development Department, Iraq

<sup>2</sup>College of Agriculture, AlMuthanna University, Iraq

<sup>3</sup>College of Agriculture, Kerbala University, Iraq

### Abstract

Plant extracts of *Tagetes erecta* and culture filtrate of *Trichoderma harzianum* was used to check their antifungal efficacy against *Fusarium verticilloides* mycelial growth. The results of this study revealed that the plant extracts and culture filtrate of *T. harzianum* significantly inhibited the mycelial growth of the pathogen. The effect was increased with the increasing of the concentration of extract and the period of treatment. The percentages of inhibition ranged between 20.93% – 65.65% in the case of plant extract of *T. erecta* and ranged between 13.43% - 50.81% in the case of culture filtrate of *T. harzianum*.

**Keywords:** *Fusarium verticilloides*, Biocontrol, *Tagetes erecta*, *Trichoderma harzianum*.

### Introduction

Plant extracts are generally assumed more acceptable and less hazardous than synthetic products and can be used as alternative antifungal treatment (Jobling, 2000; Guerrero-Rodríguez *et al.*, 2007). Aqueous plant extracts from garlic, creosote bush, and clove inhibited the growth of *Fusarium oxysporum* f. sp. *lycopersici*, *Rhizoctonia solani*, and *Verticillium dahliae* (López-Benítez *et al.*, 2005). *Tagetes erecta* L. is an annual ornamental plant. It has been shown that extracts of the marigold flower have antibacterial activity. In addition, in our previous study we demonstrated that the extracts prepared from the marigold root had inhibitory effects on a variety of common plant pathogens (Chen *et al.*, 2003). A study by Céspedes *et al.* (2006) reported that chloroform/methanol extracts of *Tagetes lucida* inhibited 89% of the colony radial growth of *F. moniliforme*. *Trichoderma* spp. are effective biocontrol agents against different pathogens and some isolates are also known for their ability to induce systemic resistance in plants (Harman *et al.*, 2004). The *Trichoderma harzianum* expresses inhibition against a broad spectrum of soil-borne pathogens including *F. oxysporum* f. sp. *cubense*, *F. oxysporum* f. sp. *niveum*, *F. oxysporum* f. sp. *melonis*, *F. oxysporum* f. sp. *cucumerinum* and *Rhizoctonia solani* (Huang *et al.*, 2011). Study was conducted under greenhouse conditions revealed that the application of *T. harzianum* and chitosan (1 g/l) as root dipping treatment combined with chitosan (0.5 g/l) as foliar spray has reduced *Fusarium* crown and root rot caused by *Fusarium oxysporum* f. sp. *radicis-lycopersici* incidence and severity by 66.6 and 47.6%, respectively (El-Mohamedy *et al.*, 2014). The mechanisms of action of *Trichoderma* spp. include competition for space and nutrients, antibiosis, antagonism, inhibition of pathogen enzymes and plant growth enhancement (Abd-El-Khair *et al.*, 2010; Howell, 2003). The objective of the present study is to assess the effect of *Tagetes erecta* extract and *Trichoderma harzianum* filtrate on the pathogenic fungus *Fusarium verticilloides*.

### Material and Methods

**Location of study:** The study was conducted in the laboratory of Agriculture College – university of AlMuthana. For the management of *F. verticilloides*, *Trichoderma*

*harzianum* isolate and plant extracts of *Tagetes erecta* were used under in vitro conditions.

**Source of *Trichoderma harzianum* isolate:** Isolate of *T. harzianum* was obtained from the Ministry of science and Technology - Directorate of Agriculture Research - Department of Biotechnology.

**Source of *Tagetes erecta* plants:** *Tagetes erecta* plants were obtained from the native nurseries. The plant material (leaves, flowers and roots) were used in this study.

**Effect of plant extracts on mycelial growth of *F. verticilloides*:** The antifungal ability of methanol plant extracts was assessed by poisoned food technique described by (Nene and Thapliyal, 1993). Fresh samples of plant material of *Tagetes* plants were washed with tap water followed by distilled water. The samples were disinfected by using sodium hypochlorite (5%). The samples were then dried in an oven at 70 °C for 2 days. After drying samples were ground to make powder and 10g of powder was dissolved in 100 ml of methanol. The samples were placed for 48 hrs in methanol which was then filtered by passing them through double layer filter papers. The filtrate is consider a stock solution(S) and different concentrations were prepared which were (S , S/2 , S/4).The PDA medium was poisoned by adding 10 ml of each concentration into 100 ml of medium. The medium was poured into sterilized Petri dishes and after solidification; these were inoculated with 5 mm block of pathogenic culture in the center of plates. The plates were incubated at 25 °C and each treatment was replicated three times. However, control was retained by mixing the medium only with 10 ml of distilled water. After 24 hrs the growth of the pathogen in each Petri plate was detected and colony diameter was measured (Vincent, 1947).

**Effect of Culture Filtrate of *T. harzianum* on *F. verticilloides*:** *Trichoderma harzianum* were cultured in conical flask containing Potato Dextrose Broth (PDB) for 20 days on rotary shaker at 25°C. The culture filtrate of *Trichoderma harzianum* was harvested after 20 days of incubation. To collect the filtrate, the liquid cultures were filtered through 2 layers of Whatman No.1 filter paper to remove hyphal fragments and finally filtered using a 0.22 um-sized membrane filter. Thus the samples were ready for further use. The filtrate of *Trichoderma* was mixed with PDA

separately to have different concentrations 12.5%, 25% and 50%. Supplemented, PDA mixtures were poured in sterilized petriplates. Then, 10 days old culture of 5 mm agar disc of *F. verticilloides* was placed in the center of the petriplates. The experiment was replicated 3 times and mycelial growth was measured after 7 days of incubation. For control, mycelium growth was counted in fresh PDA media without fungal filtrate. Observation of percent inhibition of mycelium growth (PIMG) was recorded by using the formula calculated by the formula: Inhibition % =  $(r_1 - r_2 / r_1) \times 100$  Where  $r_1$  was the radial growth of pathogen in control,  $r_2$  was the radial growth of pathogen in treatment (Ghildiyal and Pandey, 2008).

## Results and Discussion

### Effect of plant extracts on mycelial growth of *F. verticilloides*: Plant extracts of *Tagetes erecta* were used to

**Table 1 :** Antifungal activity of *Tagetes erecta* extract against *F. verticilloides*

Treatments	Concentrations	Mycelial growth of <i>F. verticilloides</i> (mm) after					
		3 days	%Inhibition	5 days	%Inhibition	7 days	%Inhibition
<i>T. erecta</i> extract	s	11.6	55.03	20.8	58.89	28.3	65.65
	s/2	14.2	44.96	26.3	48.02	38.1	53.76
	s/4	20.4	20.93	34.2	32.41	46.1	44.05
Control	0	25.8	-	50.6	-	82.4	-
LSD <sub>0.01</sub>		0.49	0.072	0.62	0.042	0.71	0.128

Each value is a mean of three replicates

The effect of the extract of *T. erecta* on the growth of the mycelium of *F. verticilloides* may be due to the presence of active compounds that have antifungal effects like thiophenes which affect the growth of the pathogen and several studies were confirmed the effect of this compound against pathogens (Mares *et al.*, 2002; Romagnoli *et al.*, 1998). Another study shows that the essential oil from leaves and thiophene rich extracts from marigold roots have significantly good antifungal activity against a number of soil borne and foliar plant pathogens (Saha *et al.*, 2012). Similar findings correlate with the findings of Martínez *et al.* (2014) who reported that marigold is effective against the *F. oxysporum*, because it restricts the disease symptoms 88.5% caused by the pathogen. The compounds viz., sesquiterpenes, saponins and flavonoids have certain anti-fungal properties, which is cent percent in marigold. Martínez (2012) reported that flavonoids are possessing antifungal activity against pathogens such as *Penicillium* sp. and *Rhizopus* sp. Phenolic compounds are active against the pathogen cell membranes, resulting in leakage of cytoplasmic. Similar study was conducted by Wavare *et al.* (2017) to evaluate the effectiveness of extracts of Marigold sp. (*Tagetes erecta* L.) against *Fusarium oxysporum* f. sp. *ciceri* and revealed that the floral water extract proved highly effective to reduce incidence of Fusarium wilt (69.31%) under greenhouse conditions. Kumar *et al.* (2019) investigated the antifungal activities of essential oil of *T. minuta* aerial parts against two plant pathogenic fungi *Aspergillus niger* and *Fusarium solani*

and showed that the essential oil exhibited 100% inhibition of *A. niger* at 1% concentration level while 100% inhibition of *F. solani* was observed at 0.12% concentration level. Recent study about antifungal activities of the flowers' essential oil of *Tagetes minuta*, (Z)-tagetone and thiotagetone against *Candida lipolytica*, *Candida parapsilosis*, *Trichosporon asahii* revealed that the essential oil exhibited high activities with a minimum inhibitory concentration (MIC) of 46.75 µg/mL for *C. lipolytica*, 54.63 µg/mL for *C. parapsilosis* and 28.33 µg/ mL for *T. asahii*, while the compound (Z)- tagetone presented MIC values of 57.29 µM, 72.92 µM and 57.29 µM, respectively (de Oliveira *et al.*, 2019).

and showed that the essential oil exhibited 100% inhibition of *A. niger* at 1% concentration level while 100% inhibition of *F. solani* was observed at 0.12% concentration level. Recent study about antifungal activities of the flowers' essential oil of *Tagetes minuta*, (Z)-tagetone and thiotagetone against *Candida lipolytica*, *Candida parapsilosis*, *Trichosporon asahii* revealed that the essential oil exhibited high activities with a minimum inhibitory concentration (MIC) of 46.75 µg/mL for *C. lipolytica*, 54.63 µg/mL for *C. parapsilosis* and 28.33 µg/ mL for *T. asahii*, while the compound (Z)- tagetone presented MIC values of 57.29 µM, 72.92 µM and 57.29 µM, respectively (de Oliveira *et al.*, 2019).

**Effect of *T. harzianum* culture filtrate on *F. verticilloides* growth:** The results of this study had been presented in table (2). The results had been revealed that the culture filtrate of *T. harzianum* has great potential to inhibit the growth of *F. verticilloides*. The effect of culture filtrate was increased with the increasing of the concentration of culture filtrate. The radial growth of mycelium were 36.2 mm, 44.3 mm and 51.6 mm a the concentrations 50%, 25% and 12.5% respectively after 7days of treatment as compared with the control 73.6mm. The lowest percentage of inhibition of mycelial growth was recorded at the concentration 12.5% after 3days which was 13.43% and the highest percentage of inhibition of mycelial growth was recorded at the concentration 50% after 7days which was 50.81%.

**Table 2 :** Activity of *T. harzianum* filtrate on *F. verticilloides* growth

Treatments	Concentrations	Radial mycelial growth of <i>F. verticilloides</i> (mm) after					
		3 days	%Inhibition	5 days	%Inhibition	7 days	%Inhibition
<i>T. harzianum</i> culture filtrate	50%	11.2	44.27	26.4	45.67	36.2	50.81
	25%	15.6	22.38	30.6	37.03	44.3	39.80
	12.5%	17.4	13.43	36.3	25.30	51.6	29.89
Control	0%	20.1	-	48.6	-	73.6	-
LSD <sub>0.01</sub>		0.68	0.072	2.19	0.176	3.54	2.93

The effect of culture filtrate of *T. harzianum* against *F. verticilloides* may be due to the presence of certain compounds such as N-phenylethylenediamine, phenol, phthalic acid, diallylamine and propanal in the filtrate of *Trichoderma* (Anita et al., 2012). The result of this study is in agreement with many workers who have also reported the inhibitory effect of culture filtrate of *Trichoderma* spp. upon several plant pathogens. Mishra et al. (2011) reported that more than 50% growth inhibition was found at 10% cell free culture filtrate of *T. viride* against pathogens like *R. solani*, *S. rolfisii*, *M. phaseolina* and *C. capsici* while at 20% concentration 100 % mycelial growth inhibition was observed which suggest the inhibitory action of cell free culture filtrate of *Trichoderma* as found in the present experiment. Bokhari and Parveen, (2012) found that culture filtrates of *T. harzianum* caused reduction in the growth of *Fusarium solani* by 21.3. The results of this study are also confirmatory with previous findings made by different scientists. *Trichoderma* species have characteristic to grow rapidly and have ability to suppress the pathogen by competing them for food and habitat (Devi et al., 2012) and also by inhibit pathogen through mycoparasitism (Khurood and Jite, 2012). Study was conducted by Ferrigo et al. (2014) to evaluate the effect of *T. harzianum* against the plant pathogenic fungus *F. verticillioideis* and they showed that seed bioprimering with *T. harzianum* can be a promising and environmentally friendly way to control *F. verticillioideis* kernel colonization and fumonisin accumulation. The result of this study is in agreement with the study of Gawade et al. 2012 who found the isolate of *Trichoderma* have good antagonistic effect on the mycelial growth of *F. moniliformae* and they showed that the effect was increased with the increasing of the concentration of culture filtrate. *Allium Sativum*, *Eucllyptus globulus* and *Lantana camara* and two bio-control agents (*Trichoderma harzianum* and *Trichoderma viridi*) were evaluated to check their antifungal activity against *Fusarium oxysporum* f. sp. *lycopersici*. It was observed that all the plant extracts showed significant results. *S. aromaticum* was the most effective in control of *F. oxysporum* f. sp. *Lycopersici* followed by *A. Sativum*, *E. globulus* and *L. camara*. Both the bio-control agents inhibited the growth of fungus *T. harzianum* showed 42.60% growth inhibition and *T. viride* exhibited 36.69% growth inhibition. Application of plant extracts and bio-control agents are cost effective, easily available and ecofriendly for the management of fusarium wilt disease (Khan, et al. 2017). *Trichoderma virens* and *T. viride* significantly increased the amount/activity of secreted antifungal metabolites in response to volatile compounds (VCs) produced by 13 strains of *Fusarium oxysporum*, a soilborne fungus that infects diverse plants. This response suggests that both *Trichoderma* spp. recognize the presence of *F. oxysporum* by sensing pathogen VCs and prepare for attacking pathogens (Li, et al. 2018).

## References

- Abd-El-Khair, H.; Khalifa, R.K.M. and Haggag, K.H.E. (2010). Effect of *Trichoderma* species on damping-off diseases incidence, some plant enzymes activity and nutritional status of bean plants. *J. Am. Sci.* 6: 122-132.
- Anita, S.; Ponmurugan, P. and Ganesh Babu, R. (2012). Significance of secondary metabolites and enzymes secreted by *Trichoderma atroviride* isolates for the biological control of Phomopsis canker disease. *African Journal of Biotechnology.* 11(45): 10350-10357.
- Bokhari, N.A. and Perveen, K. (2012). Antagonistic action of *Trichoderma harzianum* and *Trichoderma viride* against *Fusarium solani* causing root rot of tomato. *African Journal of Microbiology Research.* 6(44): 7193-7197.
- Céspedes, C.L.; Ávila, J.G.; Martínez, A.; Serrato, B.; CalderónMugica, J.C. and Salgado-Garciglia, R. (2006). Antifungal and antibacterial activities of Mexican Tarragon (*Tagetes lucida*). *Journal of Agriculture and Food Chemistry* 54: 3521-3527.
- Chen, H.B.; Wang, J.S.; Zhang, Z.G. and Wang, J.M. (2003). Effects of extract from root of *Tagetes patula* on watermelon resistance against *Fusarium* wilt disease. *Acta Phytopathol. Sin.* 5: 439-443.
- De-Oliveira, D.H.; Abib, P.B.; Giacomini, R.X.; Lenardão, E.J.; Schiedeck, G.; Wilhelm, E.A.; Luchese, C.; Savegnago, L. and Jacob, R.G. (2019). Antioxidant and antifungal activities of the flowers' essential oil of *Tagetes minuta*, (Z)-tagetone and thiotagetone, *Journal of Essential Oil Research.* 31(2): 160-169.
- Devi, S.S.; Sreenivasulu, Y.; Saritha, S.; Kumar, M.R.; Kumar, K.P. and Sudhakar, P. (2012). Molecular diversity of native *Trichoderma* isolates against *Fusarium oxysporum* f. sp. *lycopersici* (Sacc.). A casual agent of *Fusarium* wilt in tomato (*Lycopersicon esculentum* Mill.). *Archives of Phytopathology and Plant Protection*, 45: 686-698.
- El-Mohamedy, R.S.R.; Abdel-Kareem, F.; Jabnoun-Khiareddine, H. and Daami-Remadi, M. (2014). Chitosan and *Trichoderma harzianum* as fungicide alternatives for controlling *Fusarium* crown and root rot of tomato. *Tunisian Journal of Plant Protection*, 9: 31-43.
- Ferrigo, D.; Raiola, A.; Rasera, R. and Causin, R. (2014). *Trichoderma harzianum* seed treatment controls *Fusarium verticillioideis* colonization and fumonisin contamination in maize under field conditions. *Crop Protection*, 65 : 51-56.
- Gawade, D.B.; Pawar, B.H.; Gawande, S.J. and Vasekar, V.C. (2012). Antagonistic Effect of *Trichoderma* Against *Fusarium moniliformae* the Causal of Sugarcane Wilt. *American-Eurasian J. Agric. & Environ. Sci.*, 12(9): 1236-1241.
- Ghildiyal, A. and Pandey, A. (2008). Isolation of cold tolerant strains of *Trichoderma* sp. from glacial sites of Indian Himalayan region. *Res. J. Microbiol.* 3: 559-564.
- Guerrero-Rodríguez, E.; Solís-Gaona, S.; Hernández-Castillo, F.D.; Flore-Olivas, A.; Sandoval-López, V.; Jasso-Cantú, D. (2007). Actividad biológica in vitro de extractos de *Fluorensia cernua* D.C. en patógenos de postcosecha: *Alternaria alternata* (Fr.: Fr.) Keissl., *Colletotrichum gloeosporioides* (Penz.) Penz. y Sacc. y *Penicillium digitatum* (Pers.: Fr.) Sacc. *Revista Mexicana de Fitopatología* 25: 48-53.
- Harman, G.E.; Howell, C.R.; Viterbo, A.; Chet, I. and Lorito, M. (2004). *Trichoderma* species opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2: 43-56.
- Howell, C.R. (2003). Mechanism employed by *Trichoderma* species in the biological control of plant diseases: The

- history and evaluation of current concepts. *Plant Dis.* 87: 1-10.
- Huang, X.Q.; Chen, L.H.; Ran, W.; She, Q.R. and Yang, X.M. (2011). *Trichoderma* sp. strain SQR-T37 and its bio-organic fertilize could control *Rhizoctonia solani* damping-off disease in cucumber seedlings mainly by the mycoparasitism. *Appl Microbiol Biotechnol*, 91: 741–755.
- Jobling, J. (2000). Essential oils: A new idea for postharvest disease control. *Good Fruit and Vegetables Magazine*, Sydney Postharvest Laboratory Information Sheet. 3p.
- Khan, A.A.; Iqbal, Z.; Khan, W.A.; Khan, A.R. and Khan, AA. (2017). Antifungal potential of plant extracts and *Trichoderma* spp. against fusarium wilt of tomato caused by *Fusarium oxysporum* F. Sp. *Lycopersici*. *Plant Protection*, 01(01): 01-05.
- Khirood, D. and Jite, P.K. (2012). In-vitro efficacy of *Trichoderma viride* against *Sclerotium rolfsii* and *Macrophomina phaseolina*. *Notulae Scientia Biologicae*, 4: 39-44.
- Kumar, R.; Pandey, A. and Varshney, V.K. (2019). Antifungal Activity of the Essential Oil of *Tagetes minuta* Against Some Fungi of Forestry Importance. *Journal of Biologically Active Products from Nature*, 9(1): 67-72.
- Li, N.; Alfiky, A.; Wang, W.; Islam, M.; Nourollahi, K.; Liu, X. and Kang, S. (2018) . Volatile Compound-Mediated Recognition and Inhibition Between *Trichoderma Biocontrol* Agents and *Fusarium oxysporum*. *Front. Microbiol.* 9: 2614.
- López-Benítez, A.; López-Betancourt, S.R.; Vázquez-Badilio, M.E.; Rodríguez-Herrera, S.A.; Mendoza-Elos, M. and Padrón-Corral, E. (2005). Inhibition of mycelial growth of *Fusarium oxysporum* Schlechtend. f. sp. *lycopersici* (Sacc.) Snyder and Hansen, *Rhizoctonia solani* Kühn, and *Verticillium dahliae* Kleb. by aqueous plant extracts. *Revista Mexicana Fitopatología* 23: 183-190.
- Mares, D.; Tosi, B.; Romagnoli, C. and Poli, F. (2002). Antifungal Activity of *Tagetes patula* Extracts. *Pharmaceutical Biology*, 40(5): 400–404.
- Martínez, G.E.; Reyes, B.S. and Sanjuanillo, D. (2014). Effect of antagonists and plant extracts in the control of Protea wilt (*F. oxysporum*). *Am. J. Plant Sci.* 5: 3203-3212.
- Martínez, J.A. (2012). Natural Fungicides Obtained from Plants, Fungicides for Plant and Animal Diseases. In: Dhanasekaran D, Ed., *Fungicides for Plant Animal Diseases*.
- Mishra, B.K.; Mishra, R.K.; Mishra, R.C.; Tiwari, A.K.; Yadav, R.S. and Dikshit, A. (2011). Biocontrol efficacy of *Trichoderma viride* isolates against fungal plant pathogens causing disease in *Vigna radiata* L. *Archives of Applied Science Research*. 3(2): 361-369.
- Nene, Y.L. and Thapliyal, P.N. (1993). Fungicides in plant disease control, 3rd edition ed. International Science Publisher.
- Romagnoli, C.; Mares, D.; Sacchetti, G. and Bruni, A. (1998). The photodynamic effect of 5-(4-hydroxy-1-butiny)-2,2  $\zeta$  -bithienyl on dermatophytes. *Mycol Res.*, 102: 1519–1524.
- Saha, S.; Walia, S.; Kundu, A.; Kumar, B. and Joshi, D. (2012). Antifungal Acetylinic Thiophenes from *Tagetes minuta*: Potential Biopesticide. *Journal of Applied Botany and Food Quality*, 85: 207 – 211.
- Vincent, J.M. (1947). Distortion of fungal hyphae in the presence of certain inhibitors. *Nature* 159: 850-850.
- Wavare, S.H.; Gade, R.M. and Shitole, A.V. (2017). Antifungal efficacy of floral extracts, biocontrol agents and fungicides against *Fusarium oxysporum* f. sp. *Cicero*. *Indian Phytopath.* 70(2): 191-199.