



## REDUCING NEPHROTOXICITY INDUCED BY $\text{TiO}_2$ NANOPARTICLES BY SOME MEDICINAL PLANTS

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### Abstract

Titanium dioxide ( $\text{TiO}_2$ ) nanoparticles (NPs) are used as a white pigment in the production of paints, plastics, foods, paper, cosmetics and toothpaste. Although the wide ranges of uses, there is not enough information on the effect of  $\text{TiO}_2$ NPs on animal and human health. Our study evaluated the ameliorative effects of Turmeric and/or Graviola extract against the titanium dioxide nanoparticles ( $\text{TiO}_2$ NPs) on rats for 1 month. The study was performed in 80 Adult male Albino rats and divided into eight groups. Rats were administered  $\text{TiO}_2$  NPs by 600 mg/Kg body weight (BW) orally, Turmeric by 400 mg/kg BW orally and Graviola by 400 mg/kg BW orally. Administration of  $\text{TiO}_2$ NPs induced a highly significant increase in serum Creatinine levels, BUN levels and MDA in addition,  $\text{TiO}_2$ NPs rats exhibited anemia, thrombocytopenia, leukocytosis and significantly decrease in catalase. Also  $\text{TiO}_2$ NPs induced histological alterations in the kidney. But groups that were administrated with Turmeric and /or Graviola showed improvement when compared with control group. Our results indicate that Turmeric and/or Graviola effectively protect against  $\text{TiO}_2$  nephrotoxicity.

**Key words :**  $\text{TiO}_2$ NPs, Graviola, Turmeric, Catalase, MDA and nephrotoxicity.

### Introduction

Nanotechnology is a main important new technology in the field of molecular science that can be used in medical, agriculture, processing, military, cosmetics and manufacturing fields (Kisin ER, Murray AR *et al.*, 2007; Robertson TA, Sanchez WY, Roberts MS., 2010). The main basis of the nanotechnology has resulted in a lowering of particle sizes, which enhances cellular uptake efficiencies and gives novel physical characters that are potentially useful in biomedical research (Tholouli *et al.*, 2008; Stark, 2011; Hemmerich P.H. and von Mikecz A.H., 2013).

The huge number of products containing nanoparticles (NPs) and their wider applications results in increased released to the environment; NPs vary in their physical, chemical and toxicological properties with regards to the same material on a large scale.  $\text{TiO}_2$ NPs have two main

forms of crystal structures, termed rutile and anatase. Both are toxic but the anatase NPs may result much more toxic than rutile NPs and these particles alternately associated with oxidizing mechanisms of an organism, which able to generate ROS (Gurr *et al.*, 2005) including oxidative stress, lipid peroxidation and DNA damage (Vamanu *et al.*, 2008; Falck *et al.*, 2009; Wang JX *et al.*, 2009; Ibrahim M.A., khalaf M.K. *et al.*, 2015).

In recent years,  $\text{TiO}_2$ NPs are particularly used in paints, plastics, papers, ink, cosmetics and skin care products especially in the case of 1-100 nm size (Nemmar *et al.*, 2008; Brunet *et al.*, 2009) Moreover, it has unique characteristics such as small size, large surface area per unit mass and high activity that NPs can rapidly enter the human body and then imposes high health risk on human Excellency (Oberdorster *et al.*, 2005; Warheit *et al.*, 2007).  $\text{TiO}_2$  NPs containing products are easily pass through the human body by many means with different

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ways and may interrupt the body metabolism. Until now, most of the toxicological studies of  $\text{TiO}_2$  NPs in mammalian models have concentrated on the toxicity by inhalation or dermal exposure. In toxicological researches of exposure to NPs, the alterations of some enzyme levels either by increasing or decreasing which directly reflects the damage in specific cells and organs (Moss OR, VA., 2006; Shi H., et al., 2013).

Antioxidants in food play a major role in the prevent and hindering of oxidative stress-related diseases/disorders (Pérez-Jiménez, J. arranz et al., 2008). Turmeric (*Curcuma longa*), the bright yellow of the spice rainbow is a powerful medicinal plant that has long been used in the Chinese and Indian medicine, (Warrier PK, 1994) as an anti-inflammatory factor and to treat a lot of disorders including jaundice, hepatitis, (Deshpande et al., 1991) flatulence, cystic fibrosis, menstrual distress, & rheumatoid arthritis. (Paranjape et al., 2001) It also reduces cholesterol level, thus prevents cardiovascular disorders & has antioxidant effect against liver, kidney diseases, and cancer (Bafna and Mishra, 2006; khajehdehi P., 2012; Khorsandi L., Mansouri E., et al., 2016). Many studies detect the protective and the therapeutic effects of Cur in many forms of kidney injury, renal oxidative stress, lipid peroxidation (Kim BH et al., 2016)

Graviola (*Annona muricata*) is a natural plant used in treatment of some diseases and disorders. Researches proved that the leaves extract has antihypertensive properties in rats. Moreover other properties and uses of *A. muricata* leaves mentioned by traditional folk medicine such as anti-cancer agent, hypoglycemic, antibacterial, anti-fungal and anti-mutagenic (Adeyemi et al., 2008; Moghadamtousi S.Z., Fadaeinab M., Nikazd S. et al., 2015).

## Materials and Methods

### Experimental animals

Eighty healthy adult male Swiss albino rats, their weights  $170\pm20.0$  g, 6-8 weeks old, brought from the experimental animal house of the National Research Centre (NRC), Cairo University, Egypt. The animals were kept under typical environmental conditions on 12 hours light/dark cycle under a constant temperature of  $(25\pm1)$  °C, and a relative humidity, free access to food and water was permitted all the time.

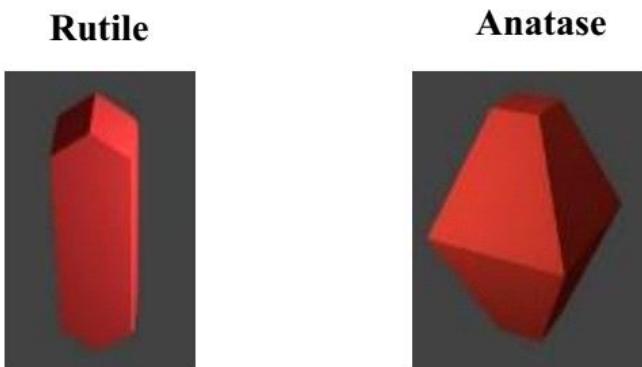
### Experimental design

All animals were randomized and divided into eight equal groups.

### Characterization of Titanium Dioxide Nanoparticles

$\text{TiO}_2$ NPs are typically available in the anatase or

rutile crystal forms. Factors such as particle size, crystal form and aggregation potential affect their bioactivity. (Warheit D, et al., 2007) Different crystal forms of the nanoparticle have previously been shown to elicit different toxicological responses. Fig.1 is a cartoon representation of rutile and anatase crystal forms of  $\text{TiO}_2$  nanoparticles.



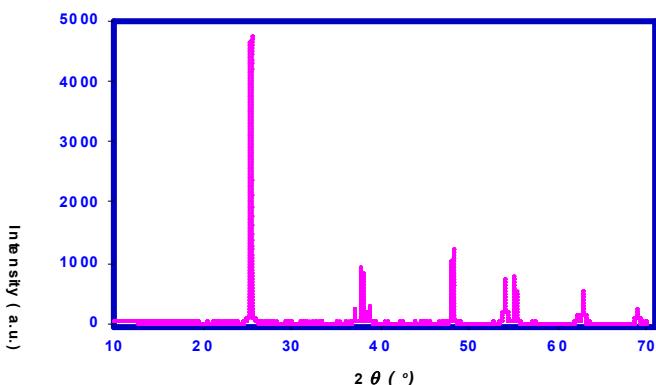
**Fig.1:** Schematic Representation of Rutile and Anatase Crystal shapes of Titanium Dioxide Nanoparticles.

### Preparation of $\text{TiO}_2$ nanoparticle suspensions

We chose a commercially available form of  $\text{TiO}_2$ NPs (anatase) for the study to detect its toxicological results on rats.  $\text{TiO}_2$ NPs brought from Loba Chemie, Mumbai city, India. The NPS were UV sterilized and stock suspensions were kept in sterile Phosphate Buffer Saline (PBS) at pH 7.4, shaken for 2 minutes, sonicated for 30 min and stored in dark at 4°C until use, and shaking well before gavaging (Joo NY, et al., 2013).

### X-Ray Diffraction (XRD)

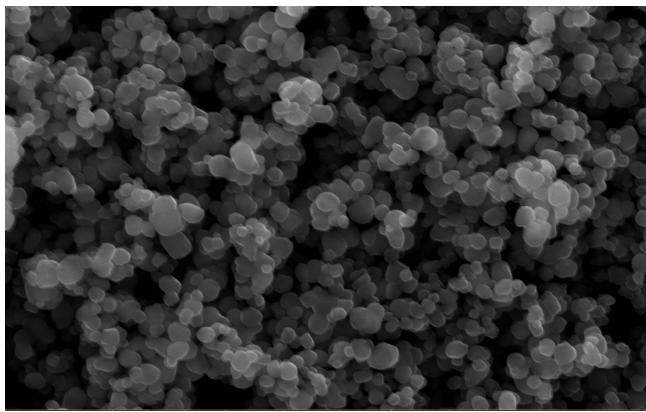
XRD pattern of the investigated  $\text{TiO}_2$  sample is illustrated in Fig. 2, the sample of  $\text{TiO}_2$  revealed anatase form with excellent crystallinity. Also, it crystallized in the well-known tetragonal symmetry with 4 molecules per unit cell. The statistics were assessed and indexed with the ICDD (International Centre for Diffraction Data) card no 21-1272.



**Fig. 2:** XRD patterns of Spherical  $\text{TiO}_2$  nanoparticles.

## Field Emission Scanning Electron Microscopy (FESEM)

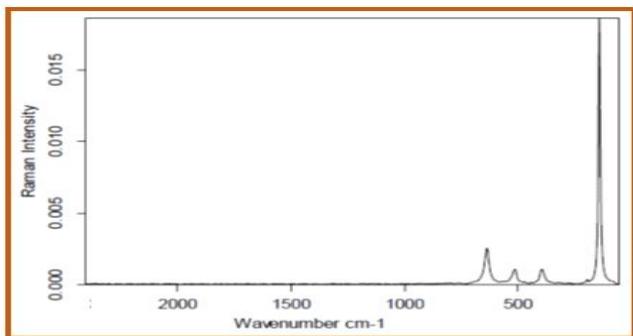
The particle size was calculated by Scherer's formula and it was found to be 69 nm. The FESEM Fig. of  $\text{TiO}_2$  nanoparticles is illustrated in Fig. 3, the  $\text{TiO}_2$  particles appeared to be with homogenous distribution with a small degree of agglomeration.



**Fig. 3:** FESEM image of  $\text{TiO}_2$  nanoparticles.

## FT-Raman Spectroscopy

The phase of  $\text{TiO}_2$  NPs was confirmed from FT-Raman analysis and the spectra are shown in Fig. 4, where active peaks near 144, 305, 510, 624  $\text{cm}^{-1}$  are prominent for the anatase phase  $\text{TiO}_2$  NPs because of Eg, B1g, A1g and Eg vibrational changes respectively. Neither signal characteristics of brookite nor rutile phases of  $\text{TiO}_2$  having Raman shifts in the range of 249- 826  $\text{cm}^{-1}$  respectively, appear in the spectra.



**Fig. 4:** FT-Raman spectra of  $\text{TiO}_2$  nanoparticles.

## 3. Chemicals (Reagent)

### Titanium dioxide nano particles ( $\text{TiO}_2$ )

$\text{TiO}_2$  NPs (lot no. # SL53251301, CAS:13463-67-7, Mumbai, India) was obtained from LOBA Chemie, assay (cerimetric) min. 99.0%, Molecular weight 79.90 g/mol

### Turmeric (Curcumin)

Rapid release gelatinous Capsules, (400 mg/Capsule) was obtained from Vitamin World, Inc. Ronkonkoma, NY 11779 U.S.A. and kept at temperature not exceeding

30°C until using and away from moisture. Each Capsule Contains: 400 mg turmeric (rhizome), preservative-free gelatin, Vegetable Cellulose, Silica and vegetable magnesium Stearate. Aliquots 400 mg/kg Turmeric were dissolved in distilled water and administered orally for 30 consecutive days to groups of Turmeric, Turmeric + Graviola,  $\text{TiO}_2$  + Turmeric + Graviola rats

### Graviola (*Annona muricata*)

Rapid release gelatinous veggie Capsules, (750 mg/Capsule) was obtained from Maximum International Maximize, U.S.A. and kept at temperature not exceeding 30°C until using and away from moisture. Each Capsule Contains: 750 mg Graviola (leaf), preservative-free gelatin, Vegetable Cellulose, dicalcium phosphate, Silicon dioxide and vegetable magnesium Stearate. Aliquots 400 mg/kg Graviola were dissolved in distilled water and administered orally for 30 consecutive days to groups of Graviola, Turmeric+ Graviola,  $\text{TiO}_2$  + Turmeric+ Graviola rats. Animal weights were measured and evaluated daily.

## Reagents and laboratory wares

All reagents used in this study were analytical of the purest grades. All glasses and plastic wares were cleaned with detergent and acid, and washed with distilled water.

## Dissection of animals

At the end of the study, the animals were fasted overnight and maintained under light ether anesthesia, immolated by cervical dislocation, and samples were collected from each animal into two tubes both with and without anticoagulants. And the Kidneys were dissected out.

## Blood and tissue samples collection

Blood was drawn from all animals in each group and centrifuged at 3000 rpm for 10 minutes. Plasma and Serum samples were stored at 0°C until biochemical analysis in the same day. Kidney was removed, cleared from adhering connective tissue; the kidney was fixed in 10% formalin for histopathological examination. And the second kidney tissue was used for the analysis of antioxidant and oxidative stress parameters.

## Preparation of kidney homogenate

Specimens from kidney tissue were weighted and homogenized for 10 sec. then being mixed with 1:9 cold phosphate buffer (PH 7.0) (Maldonado *et al.*, 2003), in an ice-containing medium. The homogenates were centrifuged at 19,000 rpm at 4°C for 30 min and the supernatants obtained were transferred into eppendorf tubes, and preserved at -80°C until using. The supernatant was used to measure the MDA level (Satoh K, 1978).

## Biochemical, Antioxidant and Lipid peroxidation Analysis

The determination of creatinine was determined according to (Bartels H, 1971). And (Fabiny DL and Ertingshausen G, 1971) Using reagent kits obtained from BioSystems Chemical Company (Spain). Creatinine in the sample reacts with picrate in alkaline medium forming a colored complex. The complex formation rate is measured in a short period to avoid interferences.

Levels of BUN in Serum were determined in accordance with the method provided by the Diamond Diagnostics kits. (Kaplan A.Urea. Kaplan a *et al.*, 1984, Tabacco A *et al.*, 1979 and Fawcett J K *et al.*, 1960).

Levels of CAT were assayed by the method of Aebi and Fossati (Aebi, H., 1984 and fossati, P., *et al.*, 1980). Briefly, Collect blood using an anticoagulant such as heparin Centrifuge the blood at 4000 r.p.m for 15 min. at 4°C. Pipette off the top yellow plasma layer without disturbing the white buffy layer. Store plasma on ice until assaying or freeze at – 80°C. The plasma stored will be stable for at least one month.

Levels of MDA were determined by Satoh's method (Satoh K, 1978). During dissection; perfuse the chosen tissue with a PBS Solution to eliminate any red blood cells and debris. Homogenate the tissue in 5 : 10 ml cold buffer per gram tissue, homogenate of kidney was made by 0.1 mol/L phosphate buffer then centrifuged at 4°C, at 4000 rpm for 15 minutes, then take the supernatant for assaying and store on ice. In case of not processing on the same day, freeze the sample at – 80°C and the sample will be stable for at least one month.

## Histopathological Examination

For histological studies, animals were sacrificed and Kidneys were dissected out and fixed in 10% neutral formalin then stained by using Hematoxylin, Eosin.

## Statistical Analysis

The data were examined by using one-way analysis of variance (ANOVA) followed by LSD computers to contrast various groups with each other and with the control. Outcomes and findings were stated as mean ± SD. The level of significance was stated as P>0.05 for insignificantly different, while P<0.05 was significantly different, while P<0.01 and P<0.001 were highly and very highly significantly different, respectively.

## Results

### Biochemistry

#### Kidney functions

The animals given Titanium alone exhibited highly

significant elevation in both serum Creatinine and BUN levels in comparison with the control. Also, the Rats in Titanium treated groups BUN and Creatinine showed a significantly reduced when compared with Ti group and slightly increased when compared with control group. As shown in (Table 1).

**Table 1:** Kidney functions Concentrations in different experimental animals.

Groups	Creatinine mg/dL	Blood Urea mg/dL
Control	0.77±0.06	30.98±2.64
Turmeric	0.73±0.06	28.68±4.04
Graviola	0.72±0.04	30.66±3.20
Turmeric + Graviola	0.73±0.04	22.92±2.09
Titanium	0.97±0.07*	58.00±8.83**
Ti + Turmeric	0.89±0.08	37.66±9.65*
Ti+ Graviola	0.80±0.07	39.26±5.21*
Ti+ Turmeric+ Graviola	0.84±0.08	38.40±5.15*

The data were expressed as Mean±SD & \* = P<0.05 and \*\* = P<0.001.

## The antioxidants and oxidative stress analysis

In the table 2, the catalase (CAT) level was highly significant decreasing in Ti group But the CAT was significant increase in Turmeric and Graviola groups. Also the CAT level was highly significant increasing In group of (Turmeric + Graviola), while there were no significant differences in other groups when compared with control one.

The MDA was highly significant Increasing in Ti group while the MDA was significant increasing in Ti+Turmeric and Ti+Graviola groups. But In case of titanium+ Turmeric + Graviola group, there were marked improvement approximately near to the results of the control group.

## Hematology

In the table 3, the hemoglobin was significant

**Table 2:** The antioxidants and oxidative stress in different experimental animals.

Groups	Catalase U/L	MDA nmol/ml
Control	218.30±21.97	1.49±0.25
Turmeric	334.04±42.05*	1.55±0.42
Graviola	320.16±0.34*	1.78±0.51
Turmeric + Graviola	496.41±16.02**	1.24±0.19
Titanium	87.12±18.11 **	5.63±0.84**
Ti + Turmeric	194.46±22.63	2.57±0.57*
Ti+ Graviola	209.12±14.60	2.52±0.75*
Ti+Turmeric+Graviola	291.73±56.12*	1.98±0.30*

The data were expressed as Mean±SD & \* = P<0.05 and \*\* = P<0.001.

decreasing in Titanium group and in case of treatment with Turmeric or Graviola or both of them; there is no differences between the other groups when compared with control one. In case of the WBCs count, the highly significant increasing was in Titanium group and a significant increase in the titanium treated group when compared with control group. The highly significant decreasing in platelets count was recorded in Titanium group, but in titanium treated groups there is no change when compared with the control group.

**Table 3:** The hematological parameters changes in different experimental animals.

Groups	HBg/dl	TLC103 /cm	PLT103 /cm
Control	13.62±0.8	8.4±1.34	479.6±29.53
Turmeric	13.96±1.28	8.3±1.65	441.6±38.35
Graviola	13.58±1.07	9.14±1.35	469.4±42.22
Turmeric + Graviola	14.2±1.13	8.32±0.79	529.8±31.11*
Titanium	11.88±0.66*	18.08±3.22**	353.2±40.61**
Ti + Turmeric	13.8±0.96	11.94±2.84*	413.2±66.43
Ti+ Graviola	13.7±0.68	10.82±3.07	458.0±47.99
Ti + Turmeric + Graviola	14.54±0.87	12.18±2.66*	500.2±71.01

The data were expressed as Mean±SD & \* = P<0.05 and \*\* = P<0.001.

### Histopathological observations.

#### Normal kidney control rats

There were no histological observations differences between experimental control and normal control. The kidney of normal rat is a bean-shaped organ, divided morphologically into an outer cortex and inner medulla. The functional unit of the kidney is the nephron. Each nephron is formed of Malpighian corpuscles and renal tubules; the latter consist of three major segments: a proximal convoluted tubule, loop of Henle and a distal convoluted tubule. The Malpighian corpuscles are spherical in shape and each consists of a tuft of blood capillaries known as glomerulus and a double-walled epithelial capsule known as Bowman's capsule Fig. 5a. The urinary space of the Malpighian corpuscle continues into the lumen of the proximal tubule. The proximal convoluted tubules are lined with simple cuboidal or columnar epithelial cells. The proximal convoluted tubules are lined with simple cuboidal or columnar epithelial cells with acidophilic cytoplasm and exhibit brush borders at their apices. The nuclei of these cells are spherical and centrally located. The distal convoluted tubules are differentiated from the proximal convoluted tubules by the absence of brush borders, acquiring wider lumen and lined with simple cuboidal epithelia. The nuclei of these

cells are spherical and centrally located Fig. 5a.

#### TiO<sub>2</sub> NPs group observations

Microscopic examination of kidney sections taken from rats of TiO<sub>2</sub> NPs group showed different signs of injury. Abnormal structures of glomeruli, such as being lobed with shrinkage of some glomeruli forming widen and irregular urinary space was also observed. Complete disappearance of some glomeruli and appearance of glomerular debris with ruptured Bowman's capsules which lead to formation of focal area of necrosis were seen. Also kidney exhibited severe and complete epithelial cell damage and desquamation (flattening epithelium) comparing to the control group and mononuclear leucocytes infiltration in the interstitial tissue of the renal tubules and in the glomerular tufts. Tubular necrosis, dilated tubules and fibrotic tissues were shown causing distribution in the architecture of the kidney. The epithelial cells were vacuolated and destructed near their brush borders. However the degenerated epithelial cells were sloughed as fragment and expelled into lumen of the renal tubules. Therefore; necrotic debris were observed in the tubule lumens as hyaline casts Fig. 5b and Fig. 5b\*.

#### TiO<sub>2</sub> NPs + Turmeric group

TiO<sub>2</sub> NPs + Turmeric group showed that the Turmeric recovered the most abnormal injuries caused by Titanium NPs except some congestion but glomeruli, urinary space and convoluted tubules appeared normal Fig. 5c.

#### TiO<sub>2</sub> NPs + Graviola group

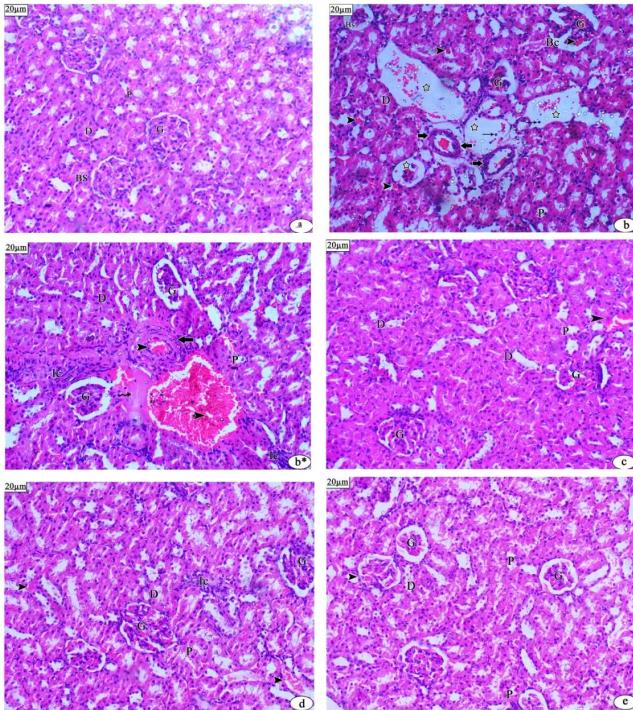
Microscopic examination of Kidney sections taken from rats treated with of TiO<sub>2</sub> NPs + Graviola group showed that the Graviola recovered some abnormal injuries caused by Titanium NPs except some congestion and Abnormal structures of glomeruli, such as being lobed with shrinkage of some glomeruli forming widen and irregular urinary space was also observed. Also kidney exhibited slightly epithelial cell damage and desquamation comparing to the control group with mononuclear leucocytes infiltration in the interstitial tissue of the renal tubules Fig. 5d

#### TiO<sub>2</sub> NPs +Turmeric + Graviola group

Microscopic examination of kidney sections taken from rats treated with of TiO<sub>2</sub> NPs + Turmeric + Graviola group showed that the Turmeric and Graviola recovered almost the most abnormal injuries caused by Titanium NPs and an approximate regain of normal appearance of the renal corpuscle and most of the renal tubules approximately Fig. 5e.

### Explanation of the figures

**Fig. 5a:** Kidney section of control: showing The



**Fig. 5:** Kidney sections of different animals groups.

Malpighian corpuscles are spherical in shape and each consist of a tuft of blood capillaries, the glomerulus (G), and a double walled epithelial capsule, the Bowman's capsule, The urinary space (BS), proximal (P) tubule and the distal convoluted tubules (D). H&E X 200

**Fig. 5b and 5b\*:** Kidney section of  $\text{TiO}_2$  NPs Rats: showing most glomeruli (G) were shranked, lobed and necrotic, some tubular cells show pyknotic nuclei and vacuolation, several congestion and interstitial hemorrhage (arrow head  $\blacktriangleright$ ), deposition of hyaline like material in some proximal tubules (zigzag arrow), mononuclear cell infiltration (IC), fibrosis (Thick arrow), exfoliation of some lining epithelial cells (arrow $\rightarrow$ ), of several necrotic areas of glomeruli (star \*), widening of bowman's space (BS). H&EX 200

**Fig. 5c:** Kidney section of  $\text{TiO}_2$  NPs Rats treated with Turmeric: showing some blood congestion ( $\blacktriangleright$ ), the glomeruli (G) appeared slightly swollen and the urinary space of the capsules got narrow was observed. H&E X 200

**Fig. 5d:** Kidney section of  $\text{TiO}_2$  NPs Rats treated with Graviola: showing some blood congestion ( $\blacktriangleright$ ), abnormal structures of some glomeruli (G), such as being lobed with shrinkage of some glomeruli forming widen and irregular urinary space, little mononuclear cell infiltration between degenerated tubules and renal corpuscle (IC), was also observed. H&E X 200.

**Fig. 5e:** Kidney section of  $\text{TiO}_2$  NPs Rats treated

with both of Turmeric and Graviola: showing an approximate regain of normal appearance. The renal corpuscle and most of the renal tubules approximately regained their normal appearance. H&E X 200.

## Discussion

Nanotechnology is the latest technology which applied highly to gather basic sciences, agriculture, food resources, biotechnology and medicine (Weir A et al., 2012). Nanoparticles pollution is considered as a new problem recently (Liu S, Yang Z., 2013). Stability of dioxide metallic nanoparticles is very high in environment and food chain which causes continuation of their toxicity (Shi JW, ZhangF, ZhaoYL, Chai ZF., 2006). Because nanoparticles have high surface action, they have a perfect capability to pass through organs. Nano- $\text{TiO}_2$  can be transported into several organs in the body through the blood and lymphatic circulatory systems. (Wang JX et al., 2007).

Nanoparticles raise toxicity by interfering in membrane structure, oxidative stress, binding protein or DNA, leading to production of active oxygen (ROS) and cell death (Cohen-Naftaly M. et al., 2011). In current study, toxicity of  $\text{TiO}_2$  NPs was investigated due to their recurrent application in industries (Weir A et al., 2012).

Medicinal plants are traditionally used to treat human diseases. Turmeric (*Curcuma longa*) is one of the most commonly used herbal medicines. Curcumin (Cur) is the major and essential component of the Turmeric (Chattopadhyay I et al., 2004). Many studies reveal the protective and the therapeutic effects of Cur in different forms of kidney injury, renal oxidative stress and lipid peroxidation (Kim BH et al., 2016). Turmeric has several benefits such as antioxidant, anti-inflammatory, anti-hyperlipidemic and anticancer protective effects (Chattopadhyay I et al., 2004).

Graviola (*A. muricata*) is a natural plant that has been used as a remedy for treatment of a variety of sickness. Studies have shown that the leaves of graviola have many properties documented by traditional uses include anti-cancerous, hypoglycemic, anti-bacterial, anti-fungal, anti-mutagenic among others (Adeyemi DO et al., 2008).

The purpose of our study was to investigate the potential renal protective activity of Turmeric and/or Graviola against  $\text{TiO}_2$  NPs induced nephrotoxicity.

Kidneys are particularly susceptible to xenobiotics and renal excretion, it considered as an expected and possible elimination route for NPs in living organisms (Burns AA et al., 2009). As the measure of plasma concentrations of creatinine and urea is usually a marker of kidney function and its other conditions (Ahamed et

al. 2010), so in the present study, in order to state any comment about nephrotoxicity, we investigated the above factors and also renal tissue structure.

According to the our results we found that oral administration of 400 mg/kg BW of Turmeric and/or Graviola for 30 consecutive days attenuate the nephrotoxicity caused by TiO<sub>2</sub> NPs. by decreasing kidney Functions (Urea, and creatinine ).

Our results revealed that serum creatinine and urea levels showed marked improvement in the treated titanium groups with turmeric and/or Graviola and lack of effect of TiO<sub>2</sub> nanoparticles on creatinine and urea excretion by kidney. Regulation of urea by kidneys is a vital part of the rat's body metabolism. In addition, to the role of urea as a carrier of waste nitrogen, it plays some interactions in the system of nephrons (Zhang *et al.*, 2012). Serum urea level showed improvement in the treated titanium groups with turmeric and/or Graviola when compared with the control group. But in TiO<sub>2</sub> NPs group a marked increase in serum urea and creatinine levels were recorded. In this regard, it can be observed the studies that nanoparticles of TiO<sub>2</sub> have raised the levels of urea (Tang *et al.*, 2010, Guo *et al.*, 2009) and creatinine (Tang *et al.*, 2010, Zhao *et al.*, 2010).

Our results in accordance with the results of (Morgan A *et al.*, 2017) he showed that administration of TiO<sub>2</sub>NPs to rats lead to obvious elevation of renal parameters, depletion of renal antioxidant enzymes with noticeable elevation in MDA concentration. Our findings come in accordance with results reported by (Escárcega-González CE *et al.*, 2016 and Farkhooni FM *et al.*, 2016). Those increases might be due to over accumulation of TiO<sub>2</sub>NPs in kidney.

In contrast, there are certain studies stated that nanoparticles of TiO<sub>2</sub> have decreased the level of urea (Liu *et al.*, 2009, Zhao *et al.*, 2010) and creatinine (Wang *et al.*, 2009).

Our results in agreement with (Kim BH *et al.*, 2016) who assessed the ameliorative effect of Curcumin (Turmeric) on renal Oxidative stress and lipid peroxidation in Type 2 Diabetic Nephropathy rats. He said that the Curcumin exhibits renoprotective characters by preventing renal lipid accumulation and oxidative stress.

Our results showed significant decreasing in Hemoglobin concentration and platelets count of TiO<sub>2</sub> NPs group, but in case of treatment with Turmeric and/or Graviola; there was no differences when compared with control one. Our results revealed a highly significant increase In the WBCs count of TiO<sub>2</sub>NPs group, and a significant increase in titanium treated groups (Ti+ Turmeric& Titanium+Graviola & Ti+ Turmeric+Graviola)

when compared with control group.

Similar observations were obtained by (Usunobun Usunomena, 2014). He studied the protective pretreatment effects of *Annona muricata* ethanolic leaf extract (400 mg / kg body weight) for 7 days versus Dimethylnitrosamine. Results showed Increase in hematological parameters compared to DMN alone group.

Our results in agreement with USunobun Usunomena except WBCs decreased because he used DMN for induction the hepatotoxicity Where DMN reduced the immunity but in our study highly significant increasing was in Titanium group and a significant increase in the group (Ti+ Turmeric& Titanium +Graviola & Ti+ Turmeric+Graviola) when compared with control group.

According to the obtained results we found that oral administration of 600 mg/kg BW of TiO<sub>2</sub> NPs(69 nm) for 30 consecutive days result in severe oxidative stress, indicated by significant elevation of MDA the indicator for lipid peroxidation (LPO) and a significant reduction for CAT concentration.

(Schanen BC *et al.*, 2009) stated that as a result of TiO<sub>2</sub>NPs photosensitivity, nano-TiO<sub>2</sub> can turn out to be a substance that produces reactive oxygen species (ROS) in the body, subsequent in free radical metabolic imbalances and an increases in ROS. Extreme ROS generate toxicity, leading to in the production of biofilms compounds that stimulate lipid peroxidation injury. When ROS is injurious, organisms use a lot of enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GSH-Px) to overcome ROS. O<sub>2</sub><sup>-</sup> is transformed to H<sub>2</sub>O<sub>2</sub> and O<sub>2</sub> by SOD, while CAT and GSH-Px can eradicate H<sub>2</sub>O<sub>2</sub> by converting it to H<sub>2</sub>O and O<sub>2</sub> (Buege J A and Aust SD 1978; Beauchamp C and Fridovich I 1971). Extreme production of ROS disrupts the balance of the liver oxidant / antioxidant system, leading to lipid peroxidation and cell apoptosis (Liu HT *et al.*, 2010).

In the present investigation, the decreased CAT concentration and the elevated MDA levels designated that the production of oxidative stress upon TiO<sub>2</sub>NPs toxicity. (Natarajan *et al.* 2015) reported that, TiO<sub>2</sub>NPs could stimulate oxidative stress and decreased the total antioxidant levels and therefore mediate LPO. (Jeon JM *et al.*, 2013) considered that part of the ROS generation might be owing to the catalytic natures of TiO<sub>2</sub>NPs. The stimulation of oxidative stress in renal tissue up on TiO<sub>2</sub>NPs intoxication had been reported by many studies (Gu SX *et al.*, 2011; Zhao JF *et al.*, 2010).

Our study revealed that titanium treated groups (Titanium+ Turmeric & Titanium+ Graviola& Titanium+ Turmeric + Graviola) showed marked improvement

occurred in CAT enzyme and lipid peroxidation. The depletion of oxidative enzymes after  $\text{TiO}_2$  NPs exposure was observed by (Salim A et al., 2013). Though, the activity of the CAT enzyme was increased when treated with turmeric.

(Ciftci et al., 2011) stated Turmeric possibly will prevent TCDD (2,3,7,8-tetra chlorodibenzo-P-dioxin) induced reactive oxygen species (ROS) due to its antioxidant character. Antioxidants existing in turmeric overcome free radicals thus fix the action of antioxidant enzymes. This means that the composites with antioxidant natures existing in turmeric protected antioxidant enzymes from exhaustion or synergized the action of these enzymes (Suhit et al., 2010). Along with the obtained results we found that oral administration of  $\text{TiO}_2$  NPs (69 nm) induced obvious nephrotoxicity marked by histopathological changes in kidney where  $\text{TiO}_2$  NPs causing several harms to rat kidney, showing several glomeruli (G) were shrunk, lobed and necrotic, some tubular cells show pyknotic nuclei and vacuolation, several congestion and interstitial hemorrhage, deposition of hyaline like material in some proximal tubules, mononuclear cell infiltration (IC), fibrosis, exfoliation of some lining epithelial cells, of several necrotic areas of glomeruli and widening of Bowman's space were observed.

The great penetrative powers of nano- $\text{TiO}_2$  permit it to pass through several organs of the body. Upon accumulations in the internal organs, nano- $\text{TiO}_2$  creates toxicity, triggering organ damage.  $\text{Ti}^{4+}$  deposition increased in each organ, involving the brain, and particularly the liver, kidney and spleen. Administered nano- $\text{TiO}_2$  predominantly deposited in the livers of rats, with an accumulation of 69% afterward administration for 5 min and 80% after administration for 15 min (Huggins CB and Froehlich JP 1966). Furthermore, a pathological assessment by Chen et al., demonstrated that nano- $\text{TiO}_2$  is accumulated in the liver, where it produces liver cell apoptosis, necrosis, and liver fibrosis, and in the kidney, where it triggers glomerular swelling. Nano- $\text{TiO}_2$  has also been shown to create toxicity in the liver and kidneys (Chen JY et al., 2009).

Accumulating studies showed that the exposure to  $\text{TiO}_2$  NPs able to bring about many renal pathological alterations in the way of inflammation of the glomeruli, cell necrosis, degenerative changes and fibrosis (Farkhouni FM et al., 2016; Huang K et al., 2015) which come in accordance with the current data.

In the histopathological studies, it was clearly found that the typical architecture of kidney tissue was observed in control, turmeric and Graviola groups. But In  $\text{TiO}_2$  NPs Group most glomeruli (G) were shrunk, lobed and necrotic, some tubular cells show pyknotic nuclei and

vacuolation, several congestion and interstitial hemorrhage, deposition of hyaline like material in some proximal tubules, mononuclear cell infiltration, fibrosis, exfoliation of some lining epithelial cells and widening of Bowman's space was observed.

Concerning the effect of nanoparticles of titanium dioxide in the kidney, (Wang et al., 2007) supposed that nanoparticles of  $\text{TiO}_2$  have been stored in the cells of kidney and began the pathological changes and nephron-like toxicity in the form of inflammation of the glomeruli of the kidney. And also particles of 25 nm  $\text{TiO}_2$  NPs can considerably increase the urea level of serum compared with the control group (Wang et al., 2007). On the other hand, the investigations show that gold nanoparticles increase the quantity of urea, but urea level return to normal after certain time. This is due to the primary shock of kidney that step by step overcome and the renal function returned to normal (Zhang et al., 2012).

According to (Borm et al., 2006) stated that nanoparticles when pass into the blood, they may be eliminated by different mechanisms depending on the way of absorption and properties of their surface. The most mutual way to remove the nanoparticles is throughout the kidneys. This process contains filtration and purification of blood in the glomeruli of the kidney nephron (Borm et al., 2006).

Recent studies show Oxidative stress can stimulate cell death by several signaling ways. The anti-inflammatory action of turmeric was described by (Hanai and Sugimoto, 2009). Turmeric's anti-inflammatory actions may be due to its capability to suppress both biosynthesis of inflammatory prostaglandins from arachidonic acid, and neutrophil function during inflammatory states (Tayyem et al., 2006; Hanai and Sugimoto, 2009).

In the current study, titanium groups treated with 400mg/kg body weight of the Turmeric extract showed no significant histological changes. From the results, it is evident that Turmeric extract has significant anti-hepatotoxic, anti-nephrotoxic activity against  $\text{TiO}_2$  NPs induced hepatotoxicity, nephrotoxicity in albino rats.

Moderate protection was observed with 400mg/kg BW dose of *A. muricata* leaf extract. There was no significant necrosis and normal structures of glomeruli (G), except some glomeruli being lobed and shrunk leading to forming widen and irregular urinary space. That mean Graviola (*A. muricata*) showed mild to moderate improvement in toxicity. Treatment with *A. muricata* leaf extract restored the renal architecture and protected the kidney tissue by preventing the toxic chemical reaction, oxidative stress, lipid peroxidation changes in the kidney tissues, (Koul IB, Kapil A, 1994; Handa SS, Sharma A 1990).

## References

- Adeyemi, D.O., O.A. Komolafe, O.S. Adewole, E.M. Obuotor and T.K. denowo (2008). Anti hyperglycemic activities of *Annona muricata* (Linn). *African Journal of Traditional, Complementary and Alternative Medicines*, **25:6(1)**: 62-9.
- Ahamed, M., M.S. AlSalhi and M.K.J. Siddigui (2010). Silver Nanoparticle Applications an Human Health. *Clinica. Chimica. Acta.*, **411** : 1841-1848.
- Bafna, A.R. and S.H. Mishra (2006). Protective effect of bioactive fraction of turmeric extract against cyclophosphamide induced suppression of humoral immunity in mice. *J. of Ethnopharmacology*, **104**: 426-442.
- Bartels, H. and M. Böhmer (1971). Eine mikromethode zur kreatininbestimmung. *Clin. Chim. Acta.*, **32**:81-85.
- Beauchamp, C. and I. Fridovich (1971). Superoxide dismutase: improved assays and assay applicable to acrylamide gels. *Anal. Biochem.*, **44**: 276-86
- Borm, P., D. Robbins, S. Haubold, T. Kuhlbusch and *et al.*,(2006). The Potential Risks of Nanomaterials: a Review Carried Out for ECETOC. *Part. Fibre Toxicol.*, **3**: 1-35.
- Brunet, L., D.Y. Lyon, E.M. Hotze, P.J. Alvarez and M.R. Wiesner (2009). Comparative photoactivity and antibacterial properties of C-60 fullerenes and titanium dioxide nanoparticles. *Environ. Sci. Technol.*, **43**: 4355-4360.
- Buege, J.A. and S.D. Aust (1978). Microsomal lipid peroxidation. *Methods Enzymol.*, **52**: 302-10
- Burns, A.A., J. Vider, H. Ow, E. Herz, O. Penate-Medina, M. Baumgart, S.M. Larson, U. Wiesner and M. Bradbury (2009). Fluorescent silica nanoparticles with efficient urinary excretion for nanomedicine. *Nano Lett.*, **9(1)**: 442-448.
- Chattopadhyay, I., K. Biswas, U. Bandyopadhyay and R.K. Banerjee (2004). Turmeric and curcumin: Biological actions and medicinal applications. *Cur. Sci.*, **87**: 44-53.
- Chen, J., X. Dong, J. Zhao and G. Tang (2009). In-vivo acute toxicity of titanium dioxide nanoparticles to mice after intraperitoneal injection. *J. Applied Toxicol.*, **29**: 330-337. DOI: 10.1002/jat.1414
- Ciftci, O., I. Ozdemir, S. Tanyildizi, S. Yildiz and H. Oguzturk (2011). Antioxidative effects of curcumin,  $\alpha$ -myrcene and 1,8-cineole against 2,3,7,8-tetrachlorodibenzo-p-dioxin-induced oxidative stress in rats liver. *Toxicol. Ind. Health*, **27**: 447-53.
- Cohen-Naftaly, M. and S.L. Friedman (2011). *Therap. Adv. Gastroenterol.*, **4(6)**: 391-417.
- Deshpande, U.R., S.G. Gadre, A.S. Raste, D. Pillai, S.V. Bhide and A.M. Samuel (1998). Protective effect of turmeric (*Curcuma longa* L) extract on carbon tetrachloride – induced liver damage in rats. *Indian J. Exp. Biol.*, **36**:573-577.
- Escárcega-González, C.E., I.G. Reynoso-Andeola, F. Jaramillo-Juárez, H. Martínez-Ruvalcaba and F.A. Posadas del Rio (2016). The ginkgo biloba extract reverses the renal effects of titanium dioxide nanoparticles in adult male rats, *Biochem. Res. Int.*
- Fabiny, D.I. and G. Erttingshausen (1971). Automated reaction-rate method for determination of serum creatinine with CentriflChem. *Clin. Chem.*, **17**: 696-700.
- Falck, G.C., H.K. Lindberg, S. Suhonen, M. Vippola, E. Vanhala and J. Catalan *et al.*, (2009). Genotoxic effects of nanosized and fine TiO<sub>2</sub>. *Human Exp. Toxicol.*, **28**: 339-352.
- Farkhoooni, F.M., A. Noori and A. Mohammadi (2016). Effects of titanium dioxide nanoparticles toxicity on the kidney of male rats. *International journal of life sciences*, **10(1)**: 65-69
- Fawcett, J.K. and J.E. Scott (1960). A rapid and precise method for the determination of urea. *J. Clin. Path.*, **13**: 156-9.
- Gui, S.X., Z.L. Zhang, L. Zheng, Q.Q. Sun, X.Z. Sang, X.R. Liu, G.D. Gao, Y.L. Cui, Z. Cheng, J. Cheng, M. Tang and F.S. Hong (2011). The molecular mechanism of kidney injury of mice caused by exposure to titanium dioxide nanoparticles. *J. Hazard. Mater.*, **195**: 365-370.
- Gurr, J.R., A.S.S. Wang, C.H. Chen and K.Y. Jan (2005). Ultrafine titanium dioxide particles in the absence of photoactivation can induce oxidative damage to human bronchial epithelial cells. *Toxicology*, **213**: 66-73.
- Hanai, H. and K. Sugimoto (2009). Curcumin has bright prospects for the treatment of inflammatory bowel disease. *Curr. Pharm.*, **15**: 2087-94.
- Handa, S.S. and A. Sharma (1990). Hepatoprotective activity of andrographolide against galactosamine & paracetamol intoxication in rats. *Indian J. Med. Res.*, **92**: 284-92
- Hemmerich, P.H. and A.H. Mikecz (2013). Defining the subcellular interface of nanoparticles by live-cell imaging. *PLoS One* 8 (4)e62018.
- Huang, K., C. Wu, K. Huang, W. Lin, C. Chen, S. Guan, C. Chiang and S. Liu (2015). Titanium nanoparticle inhalation induces renal fibrosis in mice via an oxidative stress upregulated transforming growth factor- $\beta$  pathway. *Chem. Res. Toxicol.*, **28(3)**: 354-356.
- Huggins, C.B. and J.P. Froehlich (1966). High concentration of injected titanium dioxide in abdominal lymph nodes. *J. Exp. Med.*, **124**: 1099-106.
- Ibrahim, M.A., A.A. Khalaf, M.K. Galal, H.A.H.A. Ogaly and A.H.M. Hassan (2015). Ameliorative influence of green tea extract on copper nanoparticle-induced hepatotoxicity in rats. *Nanoscale Res. Lett.*, **10(1)**: 1-9.
- Jeon, J.M., W.J. Kim and M.Y. Lee (2013). Studies on liver damage induced by nanosized-titanium dioxide in mouse. *J. Environ. Biol.*, **34**: 283-287.
- Joo, N.Y., J. Lee, S.J. Kim, H.M. Park, W.S. Yun, M. Yoon, N. W. J. J.O.N. Song and nanotechnology (2013). Preparation of an aqueous suspension of stabilized TiO<sub>2</sub> nanoparticles in primary particle form, **13(9)**: 6153-6159.
- Khajehdehi, P. (2012). Turmeric: Reemerging of a neglected Asian traditional remedy. *J. Nephropathology*, **1**:17-22
- Khorsandi, L., E. Mansouri, M. Orazizadeh and Z. Jozi (2016). Curcumin Attenuates Hepatotoxicity Induced by Zinc OxideNanoparticles in Rats. *Balkan. Med. J.*, **33**: 252-7.
- Kim, B.H., E.S. Lee, R. Choi and J. Nawaboot *et al.*, (2016). Protective effects of Curcumin on renal oxidative stress and lipid metabolism in a rat model of type 2 diabetic Nephropathy. *Yonsei. Med. J.*, **3**: 664-673.
- Kisin, E.R., A.R. Murray, M.J. Keane, X.C. Shi, D. Schwegler-

- Berry and O. Gorelik *et al.*, (2007). Single-walled carbon nanotubes: geno-and cytotoxic effects in lung fibroblast V79 cells. *J. Toxicol. Environ. Health A*, **70**: 2071-2079.
- Koller, A. and A. Kaplan *et al.*, (1984). Total serum protein. Clin Chem The C.V. Mosby Co. St Louis. Toronto. Princeton; 1316-1324 and 418.
- Koul, I.B. and A. Kapil (1994). Effect of diterpenes from *Andrographis paniculata* on antioxidant defense system and lipid peroxidation. *Indian J. Pharmacol.*, **26**: 296-300.
- Liu, H.T., L.L. Ma, J. Liu, J.F. Zhao, J.Y. Yan and F.S. Hong (2010). Toxicity of nano-anatase TiO<sub>2</sub> to mice: liver injury, oxidative stress. *Toxicol. Environ. Chem.*, **92**: 175-86.
- Liu, H.T., L.L. Ma, J.F. Zhao, J. Liu, J.Y. Yan, J. Ruan and F.S. Hong (2009). Biochemical toxicity of nano-anatase TiO<sub>2</sub> particles in mice. *Biol. Trace. Elem. Res.*, **129**: 170-80.
- Liu, S. and Z. Yang (2013). *Methods Mol. Biol.*, **1028**: 135-45.
- Maldonado, P.D., D. Barrera, I. Rivero, R. Mata, O.N. Medina Campos, R. Hernández-Pando and J. Pedraza-Chaverri (2003). Antioxidant S-allylcysteine prevents gentamicin-induced oxidative stress and renal damage. *Free Radic. Biol. Med.*, **35**: 317-324.
- Moghadamtousi, S.Z., M. Fadaeinab and S. Nikazd *et al.*, (2015). *Annona muricata* : A review of its traditional Uses, Isolated Acetogenins and biological activities. *Int. J. Mol. Sci.*, **16**: 15625-15658.
- Morgan, A., M.K. Galal, H.A. Ogaly and M.A. Ibrahim *et al.*, (2017). *Biomedicine & pharmacotherapy*, **93**: 779-787.
- Moss, O.R. and V.A. Wong (2006). When nanoparticles get in the way: impact of projected area on *in vivo* and *in vitro* macrophage function. *Inhal. Toxicol.*, **18(10)**: 711-716.
- Natarajan, V., C.L. Wilson, S.L. Hayward and S. Kidambi (2015). Titanium dioxide nanoparticles trigger loss of function and perturbation of mitochondrial dynamics in primary hepatocytes, *PLoS One* 10(8).
- Nemmar, A., K. Melghit and B.H. Ali (2008). The acute proinflammatory and prothrombic effects of pulmonary exposure to rutile TiO<sub>2</sub> nanorods in rats. *Exp. Biol. Med.*, **233**: 610-619.
- Oberdorster, G., E. Oberdorster and J. Oberdorster Nanotoxicology (2005). An emerging discipline evolving from studies of ultrafine particles. *Environ. Health Perspect.*, **113(7)**: 823-839.
- Paranjape, P. (2001). Indian medicinal plants. In: A guide to herbal medicine. Pratisthan: Delhi, 148-149.
- Pérez-Jiménez, J., S. Arranz, M. Tabernero, M.E. Díaz-Rubio, J. Serrano and I. Goñi *et al.*, (2008). Updated methodology to determine antioxidant capacity in plantfoods, oils and beverages: Extraction, measurement and expression of results. *Food Research International*, **41(3)**: 274-285.
- Robertson, T.A., W.Y. Sanchez and M.S. Roberts (2010). Are commercially available nanoparticles safe when applied to the skin? *J. Biomed. Nanotechnol.*, **6**: 452-468.
- Salim, A., A. Zohair and abou-arab (2013). Protective effect of turmeric on 2,3,7,8-Tetra Chlorodibenzo-P-Dioxin (TCCD) induced oxidative stress and hepatotoxicity in rats. *J. Applied sci.*, **9(3)**: 1790-1797.
- Satoh, K. (1978). Serum lipid peroxide in cerebrovascular disorders determined by new colorimetric method. *Clin. Chem. Acta.*, **90**: 37-34.
- Schanen, B.C., A.S. Karakoti, S. Seal, D.R. Drake, W.L. Warren and W.T. Self (2009). Exposure to titanium dioxide nanomaterials provokes inflammation of an *in vitro* human immune construct ACS Nano 3 2523-32.
- Shi, H., R. Magaye, V. Castranova and J. Zhao (2013). Titanium dioxide nanoparticles: a review of current toxicological data. *Part Fibre Toxicol.*, **10**: 15.
- Shi, J.W., F. Zhang, Y.L. Zhao and Z.F. Chai (2006). *Toxicology Letters*, **161(2)**: 115-23.
- Stark, W.J. (2011). Nanoparticles in biological systems. *Angew. Chem. Int. Ed.*, **50**: 1242-1258.
- Suhit, G., K. Meghana, B. Ramesh and P. Anant (2010). Activity of water soluble turmeric extract using hydrophilic excipients. *Food Sci. Technol.*, **43(1)**: 59-66.
- Tabacco, A. *et al.*, (1979). *Clin. Chem.*, **25**: 336-337.
- Tang, M., T. Zhang, Y. Xue and *et al.*, (2010). Dose Dependent *In vivo* Metabolic Characteristics of Titanium Dioxide Nanoparticle. *J. Nanosci. Nanotechnol.*, **10**: 8575-8583.
- Tayyem, R.E, D.D. Heath, W.K. Al-Delaimy and C.L. Rock (2006). Curcumin content of turmeric and currypowders. *Nutrition and Cancer*, **55**: 126-131.
- Tholouli, E., E. Sweeney, E. Barrow, V. Clay, J.A. Hoyland and R.J. Byers (2008). Quantum dots light up pathology. *J. Pathol.*, **216**: 275-285.
- Usunobun, U. (2014). Protective effects of *Annona muricata* ethanolic leaf extract against Dimethylnitrosamine (DMN)-Induced Hepatotoxicity, IOSR. *Journal of pharmacy and biological sciences*, **9(4)**: 1-6
- Vamanu, C.I., M.R. Cimpan, P.J. Hol, S. Sornes, S.A. Lie and N.R. Gjerdet (2008). Induction of cell death by TiO<sub>2</sub> nanoparticles: Studies on a human monoblastoid cell line. *Toxicol. in Vitro.*, **22**: 1689-96.
- Wang, J.X., Y.B. Fan, Y. Cao, Q.H. Hu and *et al.*, (2009). TiO<sub>2</sub> Nanoparticle Translocation and 2 Potential Toxicological Effect in Rats after Intraarticular injection. *Biomaterials*, **30**: 4590-4600.
- Wang, J.X. (2007). *Toxicology Letters*, **168(2)**: 176-85.
- Warheit, D.B., R.A. Hoke, C. Finlay, E.M. Donner, K.L. Reed, and C.M. Sayes (2007). Development of a base of toxicity tests using ultrafine TiO<sub>2</sub> particles as a component of nanoparticle risk management. *Toxicol. Lett.*, **171**: 99-110.
- Warheit, D.B., T.R. Webb, K.L. Reed, S. Frerichs and C.M. Sayes (2007). *Toxicology*, **230**: (Copyright (C) 2013 American Chemical Society (ACS). All Rights Reserved.), 90-104.
- Warrier, P.K. (1994). Chinesse Medicinal Plants, A compendium of 500 species. Orient Longman; Kottakkal., **5**: 180-185.
- Weir, A., P. Westerhoff, L. Fabriciu, Hristovski and N. Von Goetz (2012). *Environmental science & technology*, **46(4)**: 2242-2250.
- Zhang, X.D., D. Wu, X. Shen, P.X. Liu, F.Y. Fan and S.J. Fan (2012). In vivo Renal Clearance, Biodistribution, Toxicity of Gold Nanoclusters. *Biomaterials*, **33**: 4628-4638.
- Zhao, J., N. Li, S. Wang, X. Zhao and *et al.*, (2010). The Mechanism of Oxidative Damage in the Nephrotoxicity of Mice Caused by Nano-Anatase Tio. *J. Exp. Nanosci.*, **5** : 447-462.