



# PREVALENCE OF INTESTINAL PROTOZOA IN DEER IN MIDDLE OF IRAQ

Hind Dyaia Hadi<sup>1\*</sup>, Zainab Al. Makawi<sup>1</sup> and Tara Aso Taher<sup>2</sup>

<sup>1</sup>Natural History Research and Museum Center, University of Baghdad, Iraq.

<sup>2</sup>Department of Biology, College of Science for Women, University of Baghdad, Iraq.

## Abstract

To study the prevalence of intestinal protozoa of deer, 50 fecal samples were collected from Baghdad and its district, forty samples were found to be infected with intestinal protozoa and overall prevalence infection rate was 50% of *Cryptosporidium* sp., (30%) *Eimeria* sp. and (30%) *Entamoeba* sp. cumulatively, a total of 23 combinations of coinfections with other intestinal parasite (protozoa + Helminth) between detected infections were reported among these parasites (*Eimeria* sp. + *Rabditae*) (*Eimeria* sp. + *Toxocara* sp.) (*Cryptosporidium* + *Rabditae*) and (*Entamoeba* sp. + *Toxocara* sp.). This study is a survey documentation on multiple intestinal protozoa of deer in middle of Iraq.

**Key words** : Deer, wild animals, *Eimeria* sp., *Entamoeba* sp., *Cryptosporidium* sp., *Toxocara* sp.

## Introduction

Deer is the world-wide economic value of ruminant wildlife. Monitoring of diseases in wild animals has recently become a element necessary to prevent nature and humans. The presence of parasites in an animal resulting in decreased condition, decreased body weight gains and reproductive disorders, in particular in young humans, Furthermore, parasites affect animal products (meat and skin) quality and ultimately death (Fox, 2000). However, overpopulation could result to coccidiosis outbreaks in young wild animals with high mortality rates. There has been an increase in the population of roe deers in Galicia over the last two years (Acevedo *et al.*, 2005). Because livestock are taken into consideration to be the main reservoirs of *Cryptosporidium* oocysts, accurate information on the prevalence and species of *Cryptosporidium* is critical to assessing the risk to global health from the zoonotic transmission of *Cryptosporidium* via drinkable water. Reports on *Cryptosporidium* prevalence, however, and in particular *C. parvum* in livestock and wildlife are highly variable (De Waele *et al.*, 2012 and Smith *et al.*, 2014) And although it has been reported that wildlife contributes to the loading of cryptosporidium in surface waters

(Chalmers *et al.*, 2010). In most of the world's zoological gardens, different types of parasites were recorded in captive and wild animals, such as *Entamoeba* sp. (Opra *et al.*, 2010). Parasitic disease outbreaks in a limited space among farmed deer and intensive management practices mean that they are more severely infested with parasites than wild deer. (Vengust, 2003).

## Materials and Methods

The study included 50 deer animals of different sex and age. Fecal samples were collected from some area middle of Iraq, samples collected from with no diarrheal animals and it put in clean sterile containers. Gender, age, location and date of collection were reported on the tube and sample examination, all fecal samples were focused by flotation techniques (Urquhart *et al.*, 2000) and shethera solution (Chermette and Boufassa, 1988). Staining procedures methods were employed in this study modified ziehl – Neelsen staining technique as described by (Garcia *et al.*, 1983), and examine under 40x and 100x as well as using ocular micrometer to detect the size of infected phases of protozoa.

## Results and Discussion

During this study, a total of 50 fecal sample in deer

\*Author for correspondence : E-mail : hinddhiaa86@gmail.com

were examined, of which 40(80%) were found to be infected with one or more species of intestinal protozoa.

A total of three species of intestinal protozoa parasites (cyst/oocyst) were identified, namely, *Cryptosporidium* sp. (50%), *Eimeria* sp, (30%), *Entamoeba* sp (30%). (Table 1 and Table 2). (Fig. 1, 2 and 3).

**Table 1:** Total infection with of intestinal protozoa in deer.

| Animal | No.of Samples examined | No.of positive | Percentage (%) |
|--------|------------------------|----------------|----------------|
| Deer   | 50                     | 40             | 80             |

**Table 2:** Prevalence of intestinal protozoa species in deer.

| Intestinal protozoa       | No.of Samples examined | No.of positive | Percentage (%) |
|---------------------------|------------------------|----------------|----------------|
| <i>Cryptosporidium</i> sp | 50                     | 25             | 50             |
| <i>Eimeria</i> sp         | 50                     | 15             | 30             |
| <i>Entamoeba</i> sp       | 50                     | 15             | 30             |

**Table 3:** Prevalence of mixed infection intestinal protozoa with other parasite in deer.

| Mix infection                                  | No.of Samples examined | No. of case | Percentage (%) |
|--|------------------------|-------------|----------------|
| <i>Eimeria</i> sp+ <i>Rabditae</i> larva       | 50                     | 10          | 20             |
| <i>Eimeria</i> sp+ <i>Toxocara</i> sp          | 50                     | 5           | 10             |
| <i>Cryptosporidium</i> + <i>Rabditae</i> larva | 50                     | 5           | 10             |
| <i>Entamoeba</i> sp+ <i>Toxocara</i> sp        | 50                     | 3           | 6              |
| Total  | 50                     | 23          | 46             |

Results indicated mixed infection intestinal protozoa with helminth in deer. In this study the prevalence (*Eimeria* sp+*Rabditae*) infection rate (20%) was found higher than infection with (*Entamoeba* sp+*Toxocara* sp) were (6%). (Table 3) (Fig. 4 and 5)



**Fig. 1:** *Cryptosporidium* oocyst sp.

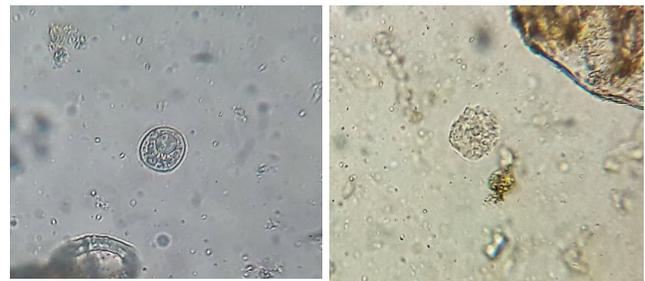


**Fig. 2:** *Eimeria* sp.

50 fecal samples in deer were examined in the current study, of which 40 (80 %) were found to be infected, these findings support earlier reports (Pacon, 1994; Santin *et al.*, 2004 and Pilarczyk *et al.*, 2005).

This study is in agreement with Tan *et al.*, (2017) who recorded gastrointestinal (GI) infections with parasites (22.9%) in deer and Barmon *et al.*, (2014) who recorded prevalence of gastrointestinal parasites of deer was (69.29%).

Based on the current study, the high prevalence of gastrointestinal parasites indicates



**Fig. 3:** Left and right *Entamoeba* trophozoite sp.

that it may have some detrimental effects on the health of these captive deers. It means that the gastrointestinal infection had an undetermined number of deers without showing any physiological outward signs of infection. It is very important in zoonotic terms because these animals can serve as reservoir of hosts for gastro intestinal parasites that are pathogenic to man (Nayak, 2016).

Similarly, Kanungo *et al.*, (2010) reported that the majority of deer noted a higher rate of mixed infection. Khatun *et al.*, (2014) reported 35.6% positive for helminth infection and 24% positive for protozoan infections, which is much lower than the current study, while Khasid *et al.*, (2003) reported 96.13% positive for



**Fig. 4:** Toxocara egg sp.



**Fig. 5:** Rabditae larva.

wild animal parasites and 100% helminthic infection was observed in spotted deer.

This was more or less similar to the Parasani *et al.*, (2001) report, which revealed that in the zoo of Rajkot Municipal Corporation, 50 % animals were positive for helminth infections and 18.8 % for protozoa. Lim *et al.*, (2008) reported 34.5 % positive for helminths and 21.8 percent %for protozoa, which is much lower than the current study for helminth infections but higher for protozoan infections.

Either as a single or mixed infection recorded at a higher rate in captive deer during this investigation shows that, effective control measures to reduce their environmental contamination are highly necessary. Frequent removal of dung and treatment of infected animals are very important gastrointestinal parasite control strategies. In addition, the practice of husbandry, the routine monitoring of parasites and the use of selective anthelmintics may be crucial for the control of parasitic gastrointestinal infections (Nayak, 2016).

Our study gives an overview of deer parasites in the middle of Iraq, but much more studies on livestock in the area and wild herbivores are needed to evaluate the dynamics of parasite transmission. Therefore, further research can also be carried out to maintain the ecological balance and to evaluate the financial losses due to parasitic deer illnesses (Barmon *et al.*, 2014).

## References

- Acevedo, P., M. Delibes-Mateos and M.A. Escudero (2005). Environmental constraints in the colonization sequence of roe deer (*Capreolus capreolus*, Linnaeus, 1758) across the Iberian Mountains, Spain. *J. Biogeogr.*, **32**: 1671-1680.
- Barmon, B.C., N. Begum, S.S. Labony, U.K. Kundu, A.R. Dey and T.R. Dey (2014). Study of gastro-intestinal parasites of deer at Char Kukri Mukri in Bhola District, Bangl. *J. Vet. Med.*, **2(1)**: 27-33
- Chalmers, R.M., G. Robinson, K. Elwin, S.J. Hadfield, E. Thomas and J. Watkins (2010). Detection of *Cryptosporidium* species and sources of contamination with *Cryptosporidium hominis* during a waterborne outbreak in North West Wales. *Journal of water and health*, **8(2)**: 311-25.
- Chermett, R. and Q.S. Boufassa (1988). *Cryptosporidiosis: A cosmopolitan diseases in animals and man* (2nded). Office international Des. Epizodies. Paris, France, 122.
- De Waele, V., M. Berzano, N. Speybroeck, D. Berkvens, G.M. Mulcahy and T.M. Murphy (2012). Peri-parturient rise of *Cryptosporidium* oocysts in cows: new insights provided by duplex quantitative real-time PCR. *Vet. Parasitol.*, **189(2-4)**: 366-8.
- Fox, M.T. (2000). Pathophysiology of gastrointestinal nematode parasitism in ruminants-an update. *Acta. Parasitology*, **45**: 253.
- Garcia, L.S., D.A. Bruchner, T.C. Brewer and R.Y. Shimizu (1983). Techniques for the recovery and identification of *Cryptosporidium* oocysts from stool specimens. *J. Clin. Microbiol.*, **18**: 185-190.
- Kanungo, S., A. Das, G.M. Das and Shakif-ul-Azam (2010). Prevalence of gastro-intestinal helminthiasis in captive deer of Bangladesh. *Wayamba Journal of Animal Science*, **2**: 42-45.
- Kasid, K.P., G.B. Srihande and G.R. Bhojne (2003). Incidence of gastro-intestinal helminthes in captive wild animals at different locations. *Zoo's Print Journal*, **18(3)**: 1053-1054.
- Khatun, M.M., N. Begum, M.A.I. Mamun, M.M.H. Mondal and M.S.U. Azam (2014). Coprological study of gastrointestinal parasites of captive animals at Rangpur Recreational Garden and Zoo in Bangladesh. *Journal of Threatened Taxa*, **6(8)**: 6142-6147.
- Lim, Y.A.L., R. Ngui, J. Shukri, M. Rohela and N.H.R. Mat (2008). Intestinal parasites in various animals at a zoo in Malaysia. *Veterinary Parasitology*, **157**: 154-159.

- Nayak, T. (2016). Studies on prevalence of gastrointestinal parasites in captive Spotted Deer in and around Bhubaneswar. Department Of Veterinary Parasitology College Of Veterinary Science And Animal Husbandry Orissa University Of Agriculture And Technology.
- Opara, M.N., C.T. Osuji and J.A. Opara (2010). Gastrointestinal parasitism In captive Animals At The Zoological Garden, Nekede Owerri, Southeast Nigeria Report and Opinion, 2 (5): 21-28.
- Pacon, J. (1994). Parasites of mouflons, stags and roe-deer from the Lower Silesia region. *Wild Parasitology*, 40: 279-92.
- Parsani, H.R., R.R. Momin, M.G. Maradin and S. Veer (2001). A survey of gastro-intestinal parasites of captive animals at Rajkot Municipal Corporation Zoo, Rajkot, Gujrat. *Zoo's Print Journal*, 16(10): 604-606.
- Pilarczyk, B., A. Balicka-Ramisz, A. Ramisz and S. Lachowska (2005). The occurrence of intestinal parasites of roe deer and red deer in the Western Pomerania voivodeship. *Wiadomosci Parazytologiczne*, 51: 307-310.
- Santin, D.M., J.M. Alunda, E.P. Hoberg and C.D.L. Fuente (2004). Abomasal parasites in wild sympatric cervids, red deer, *Cervus elaphus* and fallow deer (*Dama dama*), from three localities across Central and Western Spain: relationship to host density and park management. *Journal of Parasitology*, 90: 1378-1386.
- Smith, R.P., F.A. Clifton-Hadley, T. Cheney and M. Giles (2014). Prevalence and molecular typing of *Cryptosporidium* in dairy cattle in England and Wales and examination of potential on-farm transmission routes. *Vet. Parasitol.*, 204(3-4): 111-9.
- Tan, T.K., P. Chandrawathani, V.L. Low, B. Premaalatha, S.C. Lee, K.H. Chua, R.S.K. Sharma, N. Romano, S.T. Tay, N.H.N. Quaza and Y.A.L. Lim (2017). Occurrence of gastrointestinal parasites among small ruminants in Malaysia: highlighting *Dicrocoelium* infection in goats. *Tropical Biomedicine*, 34(4): 963-969.
- Urquhart, G., J. Armour, J.L. Duncan and F.W. Dunn (2000). *Vet. Parasitology. Longman Scientific Technical.*, 4-96.
- Vengust, G. (2003). Comparison of the parasitic fauna of fallow deer (*Dama dama*) from two enclosures in Slovenia. *Slovenia Veterinary Research*, 40: 27-31.