MOLECULAR DETECTION OF INTEGRON CLASS 1 GENE IN PROTEUS MIRABILIS ISOLATED FROM DIABETIC FOOT INFECTIONS

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Abstract

The study aim to isolation and identification of aerobic bacteria from diabetic foot ulcers in diabetic patients, investigation the resistance of bacteria to antibiotic agent. The present study included 50 specimens collected from patients suffering from diabetic foot ulcer for both sexes with an age ranged between (20-70) years. The results revealed that the frequency among males more than 40(80%) female 10(20%). The results showed that G-ve bacteria revealed a high rate 4 5(90%), 5(11%) belonged to E. coli, then P. aeruginosa 10(22%) followed by K.pneumonia with 13(28%) and proteus mirabillis 17(37%). Out of the 50 specimens, only 5(11%) isolates belong to S. aureus. Among bacterial isolates, proteus mirabilis 17(37%) was isolated and diagnosed from clinical specimens of diabetic foot ulcer and detect the antibacterial resistance to some antibacterial agent phenotypically. On the other hand to study the sensitivity of isolates to different antibiotics were tested by Kirby-Bauer disc diffusion method. The date revealed that the antimicrobial resistance rates of the P. mirabilis isolates to Colistin (100%), Levofoxacin (60%), Gentamicin (60%), Trimethoprim- sulfamethoxazole (60%) and Piperacillin-tazobactam (60%) were all high. The moderate resistance rate was observed for Ciprofloxacin (50%). The lowest resistance rate was observed for Ceftazidime (30%), Amikacin (20%), Tobramycin (20%), Ticarcillin (20%), Cefepime (10%), Meropenem (0%) and Imipenem (0%) and Aztreonam (0%). In molecular research, 15(88%) of Integron gene was used to detect Integron gene. These data indicated a high prevalence of Class 1 integrons among Foot Diabetes Ulcer patients with P. mirabilis isolates. Based on these findings, integrons that play an important role in transmitting resistance genes to the clinical bacteria as possible.

Key words: P.mirabilis, diabetic foot infection, Integrons gene, class I.

Introduction

Diabetes mellitus (DM) is a chronic condition caused by hereditary and/or acquired insulin development deficiency induced by the pancreas, or by insulin ineffectiveness. Such a deficiency contributes to increased bloodglucose concentrations, which in turn damages many of the body’s systems, especially the blood vessels and nerves and makes up the most common diabetes-rely (Spichler et al., 2015). Diabetic foot ulcers are one of the major causes of death and morbidity among diabetes sufferers. It involves damage to all tissue, necrosis or gangrene layers that typically occur on the feet’s sole due to peripheral neuropathy or peripheral arterial disease (PAD) in patients with diabetes. (Wang et al., 2012).

The genus Proteus is a Gram-negative bacillus that belongs to the Enterobacteriaceae family. Members of the genus Proteus are widespread in the environment and the gastrointestinal tract of human and animals (Hegazy, 2016; Chen et al., 2017). Proteus is known as a nosocomial, opportunistic pathogen and is more common in community-acquired infections. Proteus mirabilis and
have been reported to cause wounds infections, respiratory tract infections and both community-acquired and catheter-associated urinary tract infections (UTI) (Trivedi et al., 2015).

Bacterial resistance to antimicrobial agents is a serious problem worldwide with regard to treatment of infectious diseases. Understanding of the molecular basis of how resistance genes are acquired and transmitted may contribute to the creation of new antimicrobial strategies. The spread of antibiotic resistance is usually associated with either the clonal spread of an epidemic strain or through independent acquisition of the resistance genes on plasmids, transposons or integrons (Martínez et al., 2007). Naturally occurring gene expression elements called “integrons” have been described as vehicles for the acquisition of resistance genes carried by mobile elements.

Integrons are one of the mobile genetic elements which can carry genes of resistance to different antibiotics, which contain integrase gene, two conserve areas of 

Integrons can be divided into two major groups: resistance integrons and super-integrons. Although not independently mobile, integrons are widespread versatile DNA elements and can be divided into two distinct subsets: the mobile integrons and the chromosomal integron (Corrêa et al., 2014). Class I integral gene cassettes normally do not have their own promoters and their transcription depends on the promoters Pc and P2 (Sunde, 2005).

These structures have also been found to be involved in the genetic reassortment of resistance determinants frequently observed in multiple-antibiotic-resistant bacterial pathogens (Martínez et al., 2007).

The aim of the present study to perform a molecular analysis of Integron gene class 1 and to determine their antibiotic resistance pattern as well as comparing these recent findings with those reported in our previous study.

### Materials and Methods

#### Specimens collection and bacterial identification

Collection 50 samples swab specimens from diabetic foot infection ulcer. The specimens were transported by sterile transport swabs to the bacteriology laboratory for culture on MacConkey agar, Mannitol agar and blood agar, the plates were incubated at 37°C for 24 hours then a single pure isolated colony was transferred to trypticase soya agar for the preservation and to carry out other biochemical tests and VITEK 2 compcat system that confirmed the identification of isolates.

#### Genomic and plasmid (DNA Extraction)

Genomic and plasmid DNA was successfully extracted from P. mirabilis isolates by boiling method, briefly, colonies were suspended in 100 microliters of sterile distilled water and boiled at 100°C in the water bath for 15 minutes than rapidly cooled at -20°C for one hour, then centrifugation and the supernatant were preserved for the used in the amplification-processes (Ali et al., 2009). The concentration and purity of the extracted DNA was determined using spectrophotometry.

### Table 1: Primers used in this study.

<table>
<thead>
<tr>
<th>Primer Type</th>
<th>Primer sequence (5'-3')</th>
<th>Amplicon size (bp)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>IntI</td>
<td>F: CAGTGGACATAAGCCGTTC</td>
<td>160</td>
<td>Zareei, Y. et al., 2014</td>
</tr>
<tr>
<td></td>
<td>R: CCCGAGGCATAGACTGTA</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

### Table 2: PCR programs of primers that apply in the thermocycler.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Initial denaturation</th>
<th>No. of cycles</th>
<th>Denaturation</th>
<th>Annealing</th>
<th>Extension</th>
<th>Final extension</th>
</tr>
</thead>
<tbody>
<tr>
<td>IntI</td>
<td>94°C for 5 min</td>
<td>35</td>
<td>94°C for 30 sec</td>
<td>55°C for 30 sec</td>
<td>72°C for 1 min</td>
<td>72°C for 7 min</td>
</tr>
</tbody>
</table>
DNA were calculated directly by spectrophotometry, the purity of DNA extracted varied between (1.8-2). Gel electrophoresis validated and analyzed the extracted DNA.

**Molecular Identification**

The PCR assay was performed to detect the integron genes for *P. mirabilis* shown in table 2. Alpha DNA Company, Canada developed these primers as shown in table 1. Agarose gel electrophoresis was observed to the amplified PCR products by staining with ethidium bromide. The product of electrophoresis was detected by the use of a gel documentation device. The positive findings were separated when sample base pairs of the DNA band were equal to the target product size (Bartlett and Stirling, 1998). Finally, the gel was shot using a documentation method for the Biometra gel.

**Results and Discussion**

**Demographic profile of patients with diabetic foot ulcer**

The research was carried out on 50 specimens of suspected patients’ diabetic foot wounds between October, 2019 and February, 2020. Results showed that 50 clinical specimens were distributed according to the report patient’s age ranged from (20-70) years old. The lowest incidence was among the (20-30) and more than (60) age group 3.3% and 1.6% respectively, while the highest incidence was among the (51-60) age group (39.1%).

In 50 patients, 40(80%) were male and 10(20%) were female as in fig. 2. Increasing infected male more than female frequency similar with the result of Bansal *et al.*, (2008) they observed that male more frequency than female (78.64% vs 21.36%) from the total 270 diabetic patients (180 male and 90 female) suffering from foot infections. Predominance of males over female patients as shown in the study can be explained by the fact that in our country males are exposed more to the outside environment factors. The predominance male in DFU could be linked to factors such as gender-related differences in life styles and professional roles that require the feet to tolerate more pressure as a result of work, increased level of outdoor work and poor compliance to foot care practices compared to females. The wounds healing in females more better than in male, this may be due to differences in hormones and explain that increase estrogen receptor in female increase healing wound which act as endogenous enhancers of healing process while in male increased the level of androgen was considered harmful for wound healing since androgenic species decrease repair of dermis (Wilkinson and Hardman, 2017).

**Isolation of pathogenic bacteria of diabetic foot ulcer (DFU)**

Culture investigation depending on morphological and biochemical test shows that Gram negative bacteria reveals a high rate 45(90%), 5(11%) belonged to *E. coli*, then *P. aeruginosa* 10(22%) followed by *K. pneumonia* with 13(28%) and *proteus mirabilis* 17(37%). Out of the 50 specimens, only 5(11%) isolates belong to *S.*

<table>
<thead>
<tr>
<th>Test</th>
<th><em>P. mirabilis</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Oxidase test</td>
<td>+</td>
</tr>
<tr>
<td>Catalase test</td>
<td>+</td>
</tr>
<tr>
<td>Motility</td>
<td>+</td>
</tr>
</tbody>
</table>

Table 3: The biochemical features of *P. mirabilis*.

![General structure of class 1 integrons](image1.png)

*Fig. 1: General structure of class 1 integrons.* (Gillings, 2014).
Bansal et al. (2008) observed that gram negative bacteria were isolated in 76% whereas gram positive bacteria were isolated in 24% of DFU patients. The predominance of gram negative bacteria has been noted in this study agree with (Banu et al., 2015).

**Proteus mirabilis identification**

- **Morphologically characterization:**

  The bacterial isolates collected from clinical samples were initially classified based on cultural morphology, microscopic properties and biochemical studies. Microscopically *P. mirabilis* a gram negative bacilli emerged, *P. mirabilis*’ cultural identification was based on the colonial morphology. When the *P. mirabilis* colonies were grown on the blood agar, swarming colonies appear (Fahadawi et al., 2019).

- **Biochemical tests**

  Biochemical test results that were reported in table 3 were considered to complement the initial identification of *P. mirabilis* isolates. The isolates have reported general characteristics, the isolates have been positive for oxidase and catalase testing (Khalifa et al., 2019).

  After extensive bacteriological analysis based on morphological, cultural and biochemical samples, the isolates were tentatively identified as *P. mirabilis*.

  The final identification was carried out using an automated compact VITEK-2 Compact system with 47 biochemical tests and one GN-ID card Well performed negative regulation (Appendix 2). Results show that only 17 isolates from diabetic foot wound were identified as *P. mirabilis* with confidence level of ID message was very high (Probability percentage between 88 and 98).

**Phenotypic resistance of Isolates**

- **Antibiotic Susceptibility**

  This test was conducted to all isolates by using disc diffusion test. The results were interpreted according to the recommendation of CLSI, (2019). The date revealed that the antimicrobial resistance rates of the *P. mirabilis* isolates to Colistin (100%), Levofloxacin (60%), Gentamicin (60%), Trimethoprim-sulfamethoxazole (60%) and Piperacillin-tazobactam (60%) were all high. The moderate resistance rate was observed for Ciprofloxacin (50%). The lowest resistance rate was observed for Ceftazidime (30%), Amikacin (20%), Tobramycin (20%), Ticarcillin (20%), Cefepime (10%), Meropenem (0%) and Impenem (0%) and Aztreonam (0%). Some Proteus mirabilis isolates show an elevation in resistance to imipenem due to several reasons: loss of outer membrane porins, reduced PBP1 a expression or

![Fig. 2: Total patients of diabetic foot ulcer according to gender.](image-url)
reduced PBP2 imipenem binding (Girlich et al., 2014). That the production of imipenem resistant in Proteus mirabilis is due to a lack of OMP 24 kDa. Amikacin is an amino-glycoside antibiotic used to treat bacterial infection of different forms. This works by binding to the ribosomal subunit of the bacterial 30s which causes misreading of mRNA and leaving the Bacterium which cannot synthesize proteins that are vital to its development. Ciprofloxacin is a synthetic chemotherapeutic antibiotic of the class of fluroquinolone drugs and is a part of a broad variety of antimicrobial agents that inhibits bacterial DNA and protein synthesis and is considered an effective medication for urinary care tract infection. A Study in Taiwan by Wang et al., (2014) reported that the susceptibility to this antimicrobial decreased (from 80.1% to 53.8%). That is considered roughly in agreement to the outcome of this report, on resistant growth. Because of their synergistic impact on bacteria, trimethoprim and sulfamethoxazole work together. It is antimicrobial bactericidal agent for gram positive as well as gram negative bacteria abroad. Trimethoprim is a diaminopyrimidine, while sulfamethoxazole is a sulfonamide and co-trimoxazole inhibits the synthesis of tetrahydrofolic acid, essential for the synthesis of bacterial nucleic acid, along with two components of the drug that inhibit specific steps in the pathway of folate synthesis (Ramlakhan et al., 2014). In this study P. mirabilis isolates developed high resistance (72%) against trimethoprim-sulfamethoxazole, which was approximately confirmeds with Nahar et al., (2014) reported a resistant percentage of (66.7%).

### Molecular detection of class 1 integron gene

Integrons are mobile genetics elements that could be important in the dissemination and accumulation of resistance genes in bacteria that usually located within transposon conjugative plasmids (Mazel, 2006). Integrons are genetic elements with a specific functional configuration that has evolved in bacteria, catching and expressing cassettes of exogenous genes through site-specific recombination (Xiao et al., 2019). The result showed that the integron gene was detected in 15(88%) p. mirabilis isolates as in fig. 3.

Class 1 integrons are the most common and widely distributed among Gram-negative bacteria isolated from clinical samples and often are associated with lateral transfer of antibacterial resistance genes (Al-Sanouri et al., 2008). Integrons are mobile genetic elements that could be important in the dissemination and accumulation of resistance genes in bacteria. Integrons are usually located within transposons or conjugative plasmids (Mazel, 2006). In a previous study, 65% of isolates harbored class 1 integron (Bashir et al., 2015), whereas in another study class 1 integron was found in 49% of uropathogenic isolates (Ajiboye et al., 2009). However, these strains needs to be sequenceds by 16S rRNA or RAPD-typing for further studies as recommended by Salih and Shafeek, 2019; Banoon et al., 2019).

### Conclusions

1. The high frequency of specimens collection from patients infected with diabetic foot ulcer G-ve isolated are higher than G +ve.
2. Male is more frequent than Female

3. Most samples isolated from clinical infections had Integrons gene as one of the important mechanisms for acquisition and dissemination of antibiotics resistance mechanisms in *P. mirabilis*.

References


Xiao, L., X. Wang, N. Kong, M. Cao, L. Zhang, Q. Wei and W.